

基因體學

Genomics



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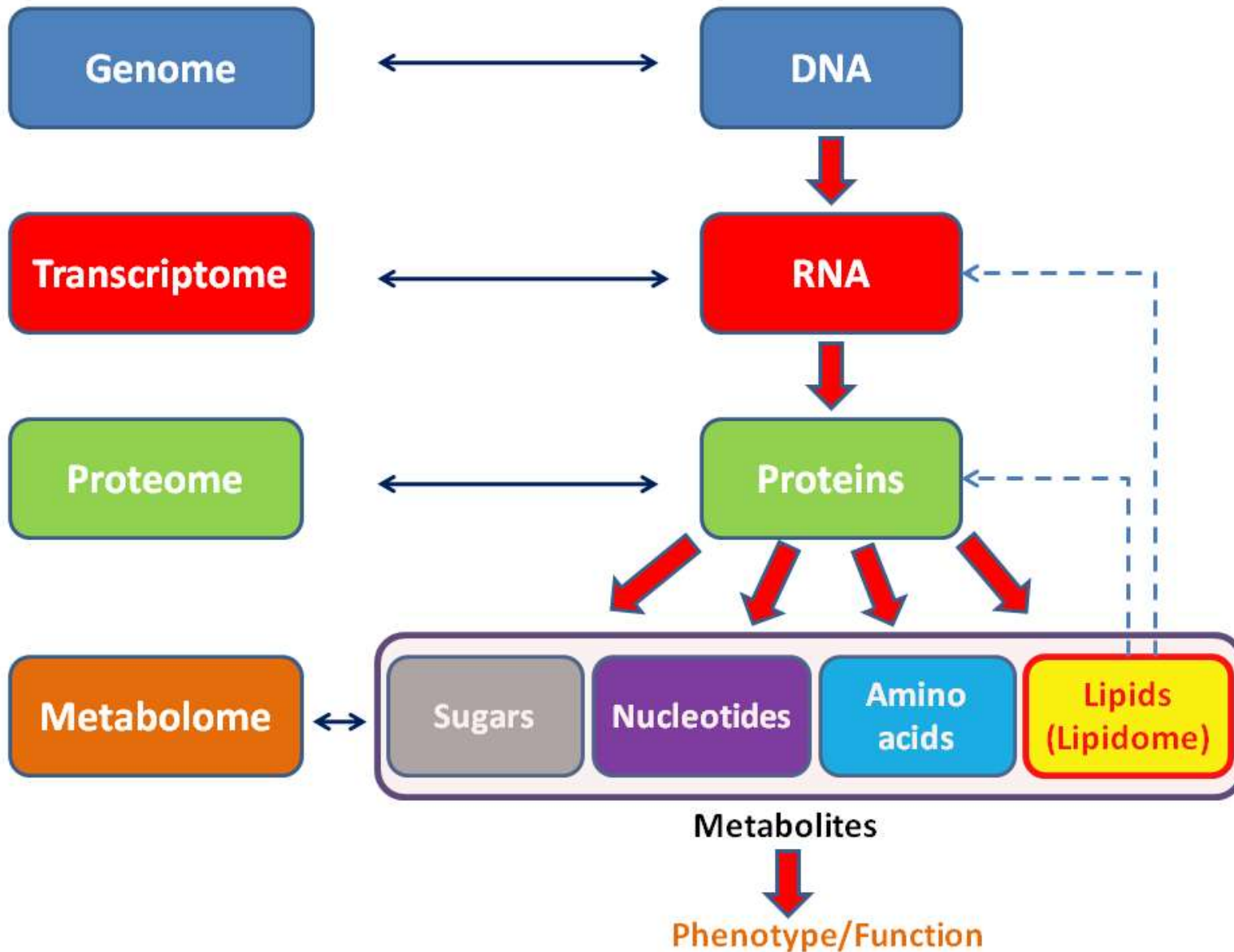
主題大綱



- Genomics (基因體學)
- History of DNA sequencing (DNA定序歷史)
- The first-generation DNA sequencing (第一代DNA定序)
- Next-generation sequencing (NGS,次世代定序)
- CRISPR/cas9 genome editing (基因編輯技術)
- Genome analysis (基因體分析)

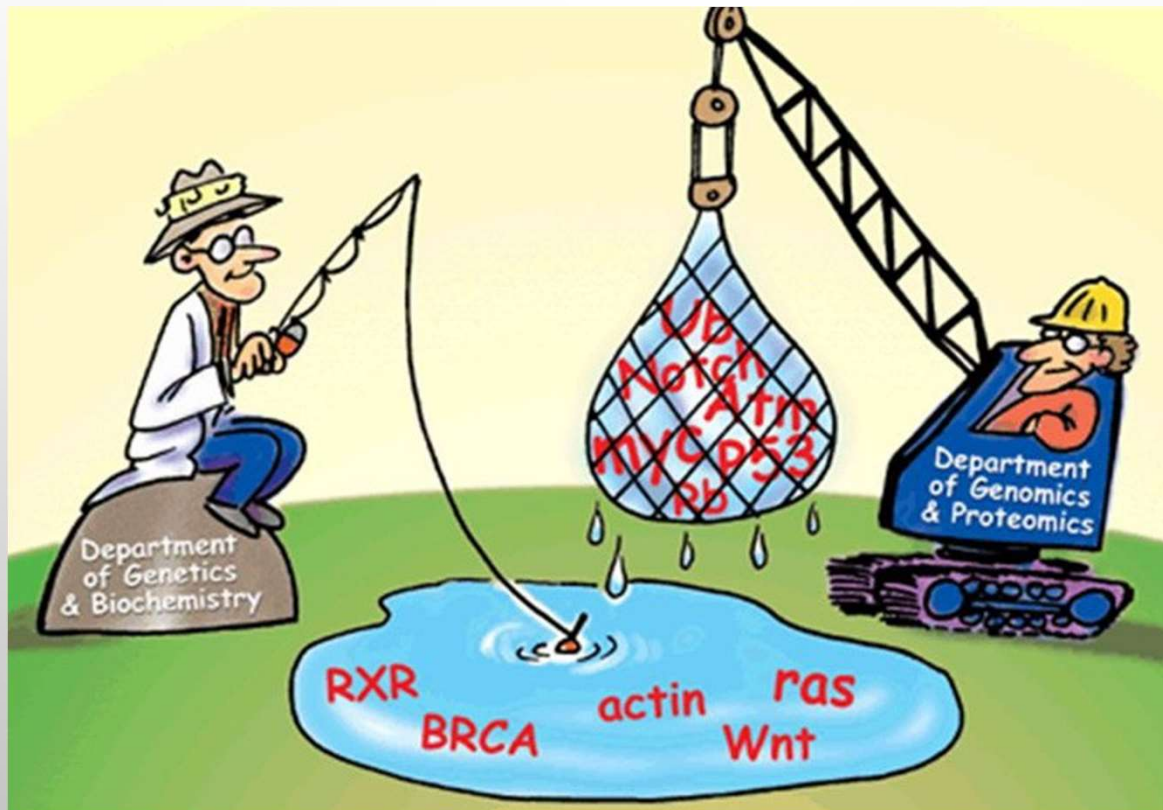
Genomics (基因體學)

The "omics" revolution



基因體學 (Genomics)

- 基因體(genome)：細胞內所有的DNA，包含核DNA(nuclear DNA)、葉綠體DNA、粒腺體DNA

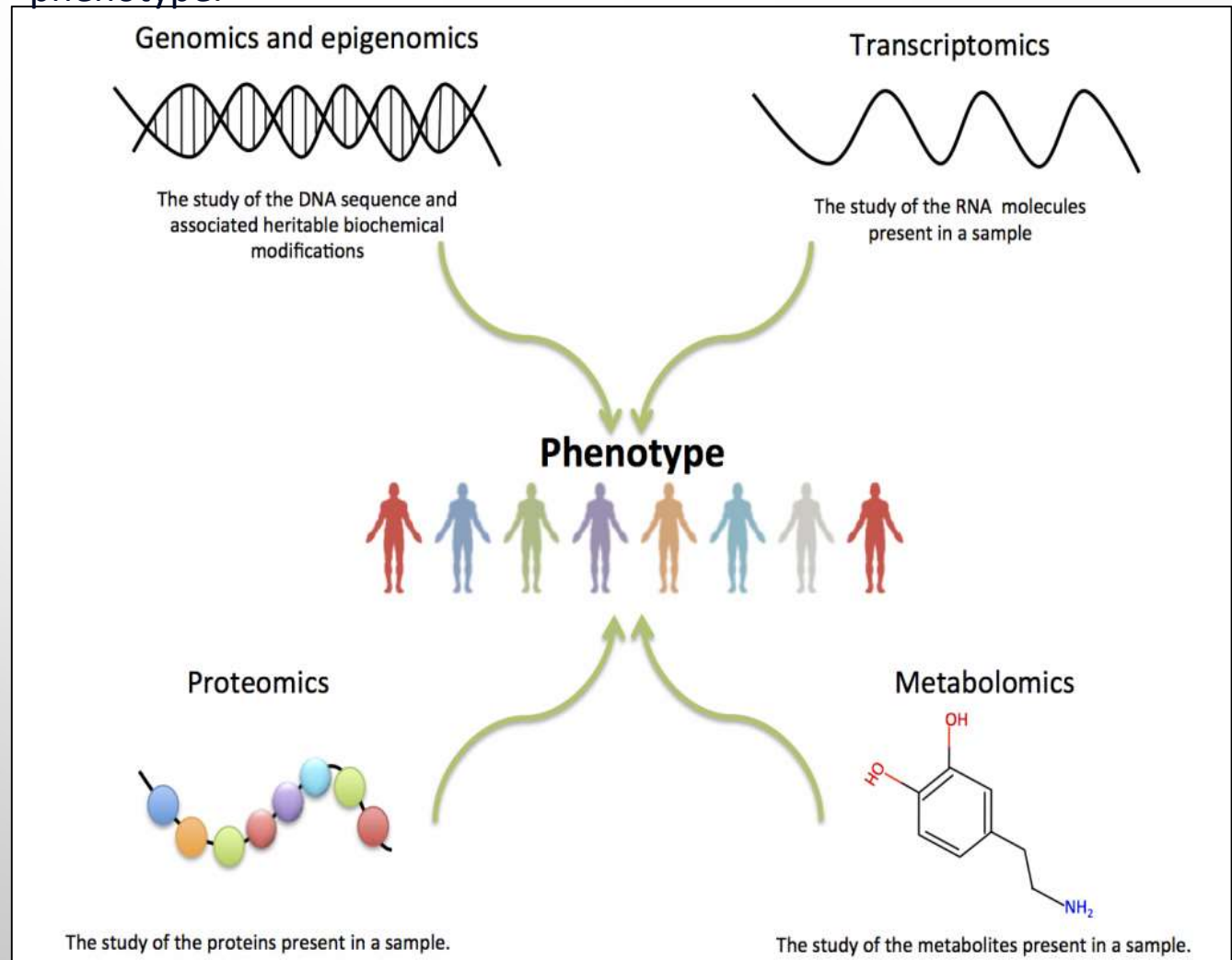


高通量(high-throughput)
策略研究基因功能

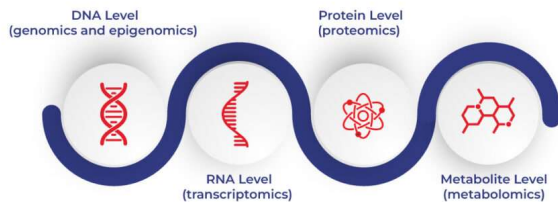
功能基因體學 (Functional genomics)

The goal of functional genomics is to determine how the individual components of a biological system work together to produce a particular phenotype.

- DNA level
(genomics and epigenomics)
- RNA level
(transcriptomics)
- Protein level
(proteomics)
- Metabolite level
(metabolomics)



Levels of Focus of Functional Genomics

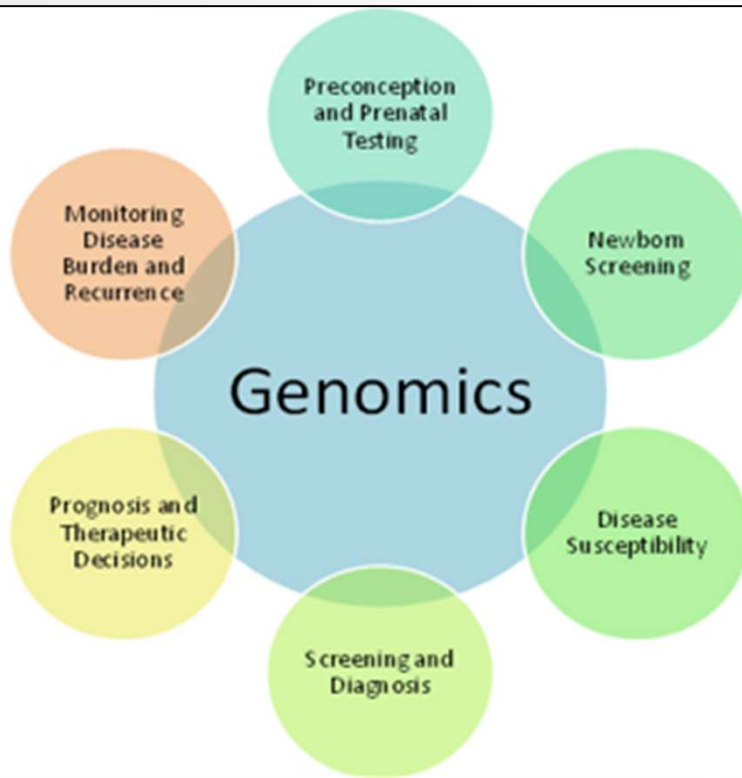


基因體學Applications

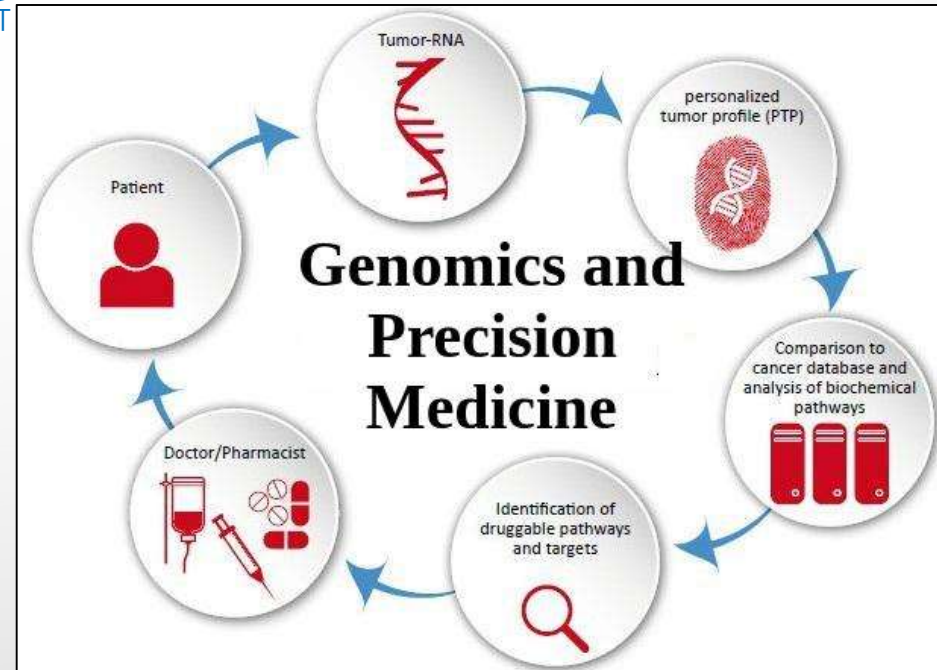
1. Gene function
2. Precision medicine
3. Genome-wide association study(GWAS)全基因組關聯分析
4. Metagenomics – 微生物菌相分析
5. Epigenomics – DNA 甲基化分析

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單一核甘酸多型性的分析



<https://www.genome.gov/minc/toolkit/why-genomics>



<https://www.openpr.com/news/2757743/upcoming-opportunities-in-genomics-and-precision-medicine>



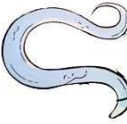



基因體大小

- 單位(bp / base pair)

1 bp = 1 bp , 1 kb = 1,000 bp,

1GB = 1,000,000,000 bp, 1MB = 1,000,000 bp

- 基因體大小

Species	<i>Porcine circovirus</i>	<i>Escherichia coli</i>	<i>Caenorhabditis elegans</i>	<i>Drosophila melanogaster</i>	<i>Homo sapiens</i>	<i>Amoeba dubia</i>
Genome Size	1759 bp	4.6 MB	100 MB	130 MB	3.2 GB	670 GB
Common Name	 Virus	 Bacteria	 Nematode	 Fruit fly	 Human	 Ameoba

可表現蛋白基
因數目

3

4288

19,000

13,600

~ 20,000

?

C值謎(C-value enigma): 生物的C值 (基因組大小) 並不與生物複雜程度相關的現象

Ex.例如植物與原生動物，可能具有比人類更大的基因組

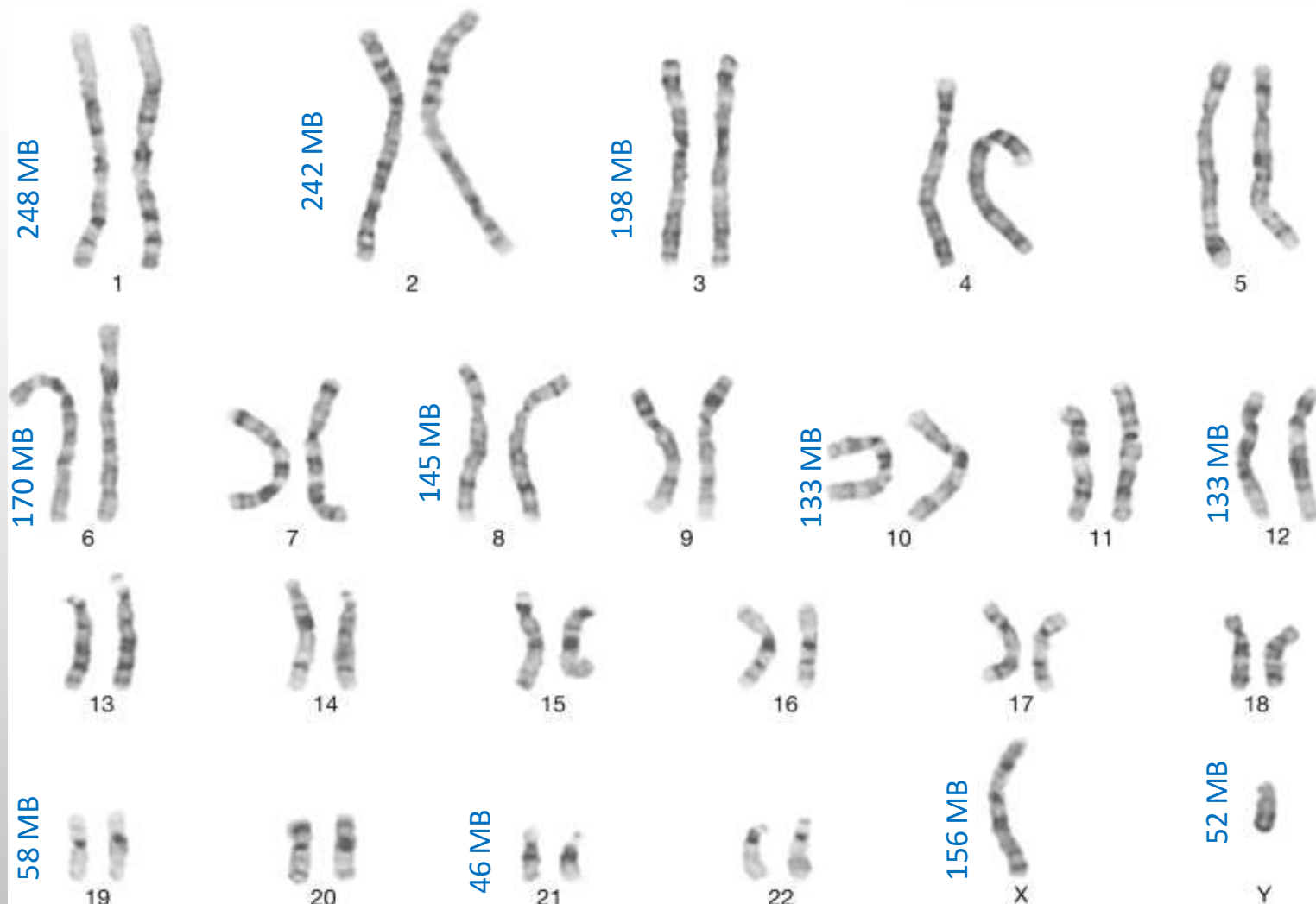
G值謎(G-value paradox) : 生物的G值 (基因數量) 並不與生物複雜程度相關的現象

Ex.顯微線蟲*Caenorhabditis elegans*由一千個細胞組成，但具有與人類大致相同數量的基因

8

Chromosome (染色體)

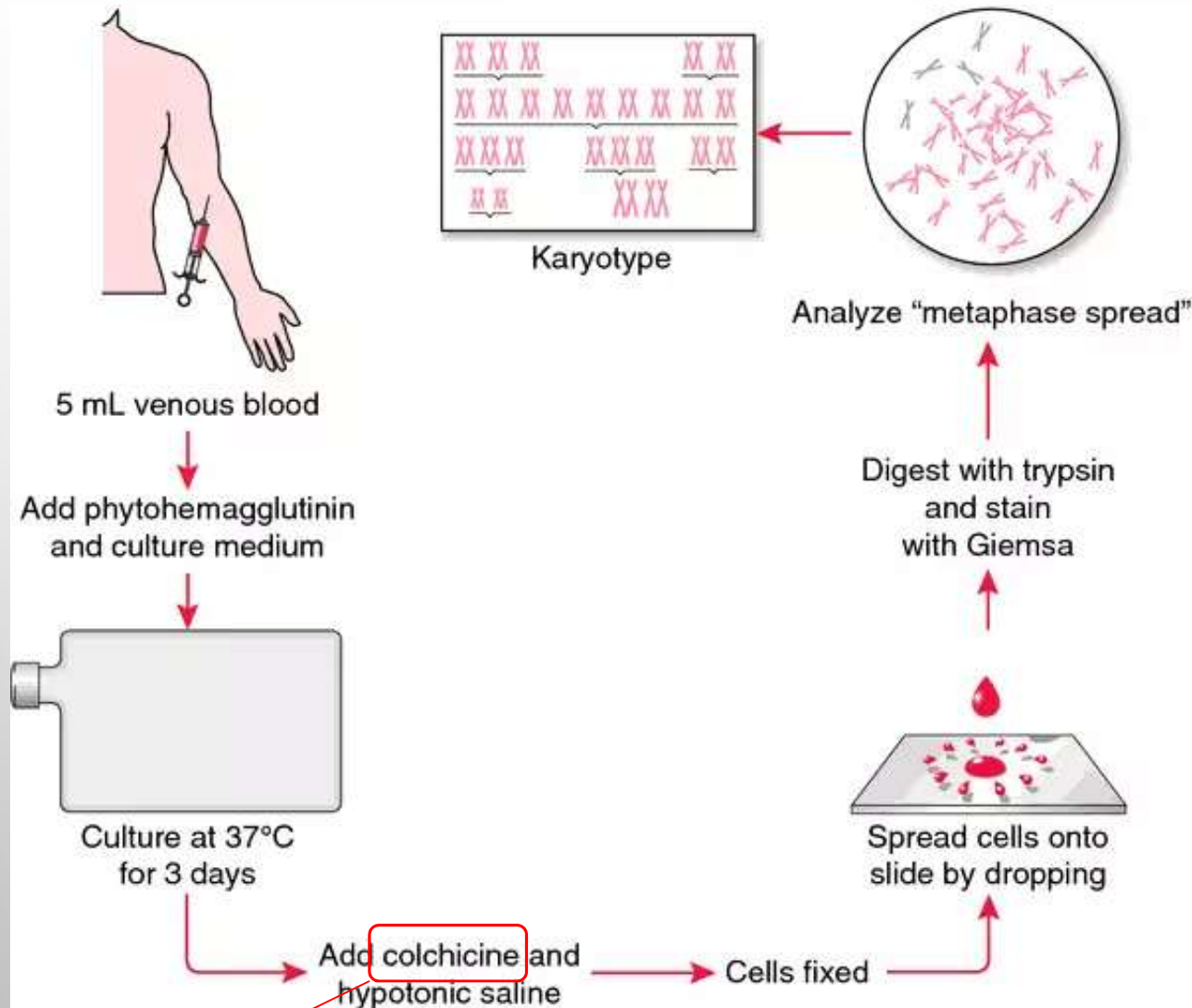
Total: 3,234.83 Mb



A.B.S Giersch, in Pathobiology of Human Disease, 2014

人類染色體

Preparation of a karyotype



在有絲分裂過程中，它不會幹擾染色體的複製，但會阻止紡錘體纖維的形成。在中期，它抑制微管的組裝，因此不會發生紡錘體形成。

. From Mueller and Young, 2001.

染色體異常疾病

疾病	發生率	檢出率	陽性預測率
唐氏症 (21 號 3 條 染色體症) Down syndrome	約 1/600~1/1,000 	>99%	>99%
愛德華氏症 (18 號 3 條 染色體症) Edwards syndrome	約 1/3,000~1/6,000 	>99%	>98 %
巴陶氏症 (13 號 3 條 染色體症) Patau syndrome	約 1/5,000-10,000 	>99%	>99%

染色體條帶技術

- 利用染劑使染色體呈現各自獨特條帶形態，藉以區別染色體的不同

技術	方法	亮帶	暗帶
G 帶	胰蛋白酶 + Giemsa	GC rich	AT rich
R 帶	熱磷酸鹽處理 + Giemsa	AT rich	GC rich
Q 帶	Quinacrine(螢光染料)	GC rich	AT rich
C 帶	熱鹼 $Ba(OH)_2$ + Giemsa	著絲點以外	著絲點

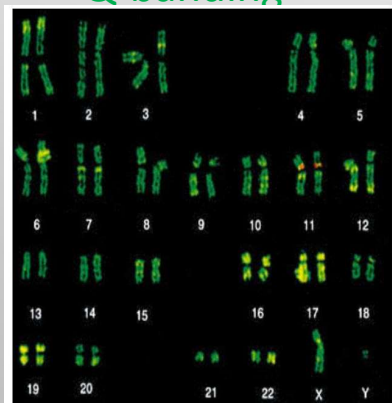
G-banding



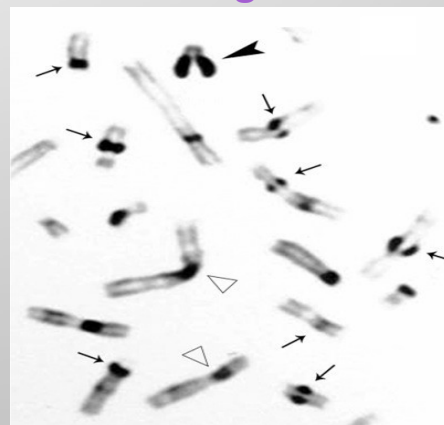
R-banding



Q-banding



C-banding

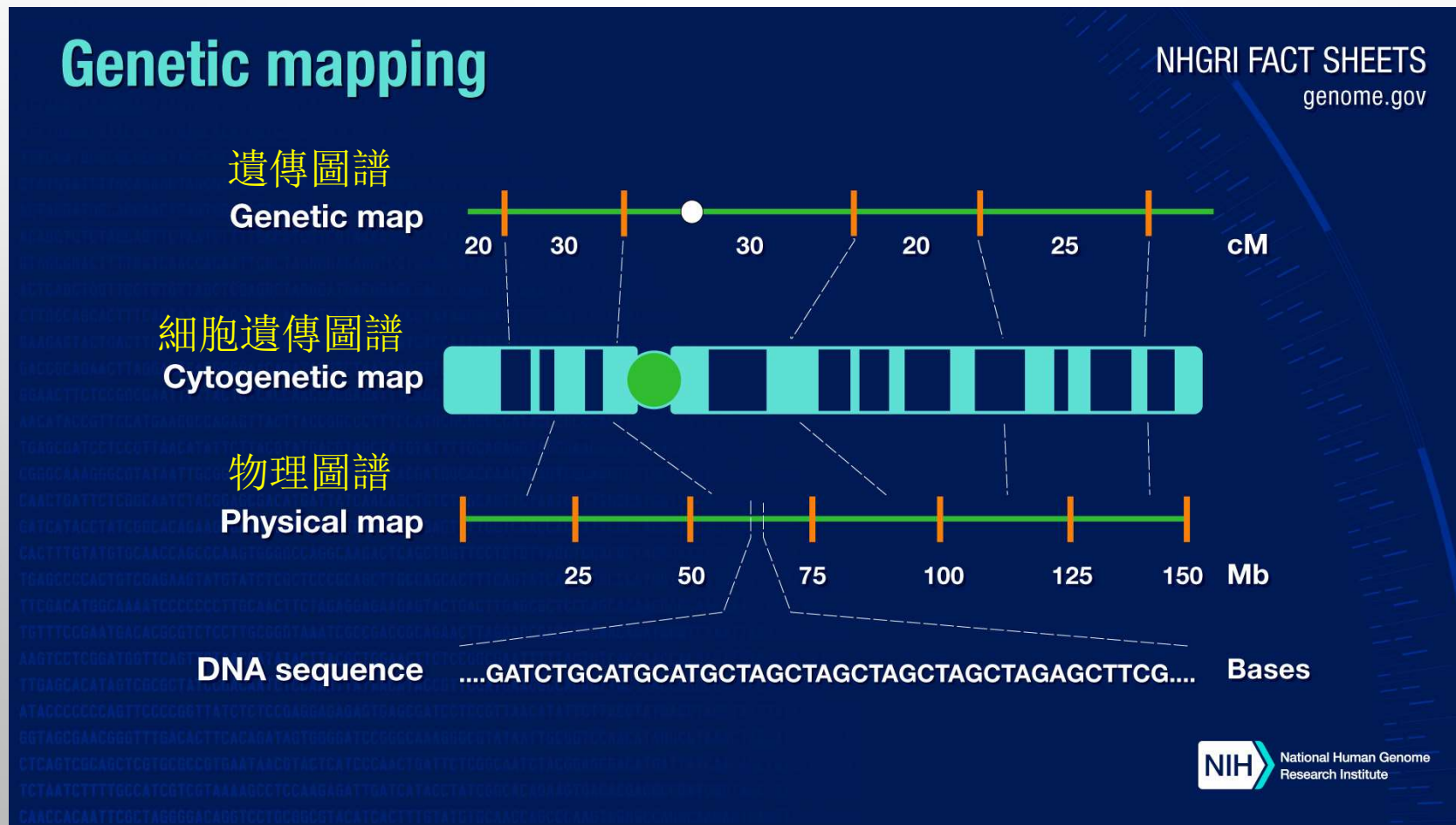


基因體圖譜 (Genome map)

Cytogenetic map (細胞遺傳圖譜): 由染色體染色而來，沒有單位，以區域劃分

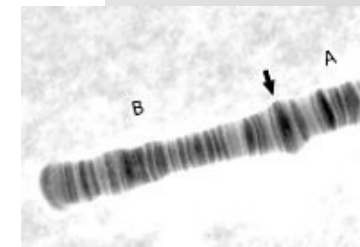
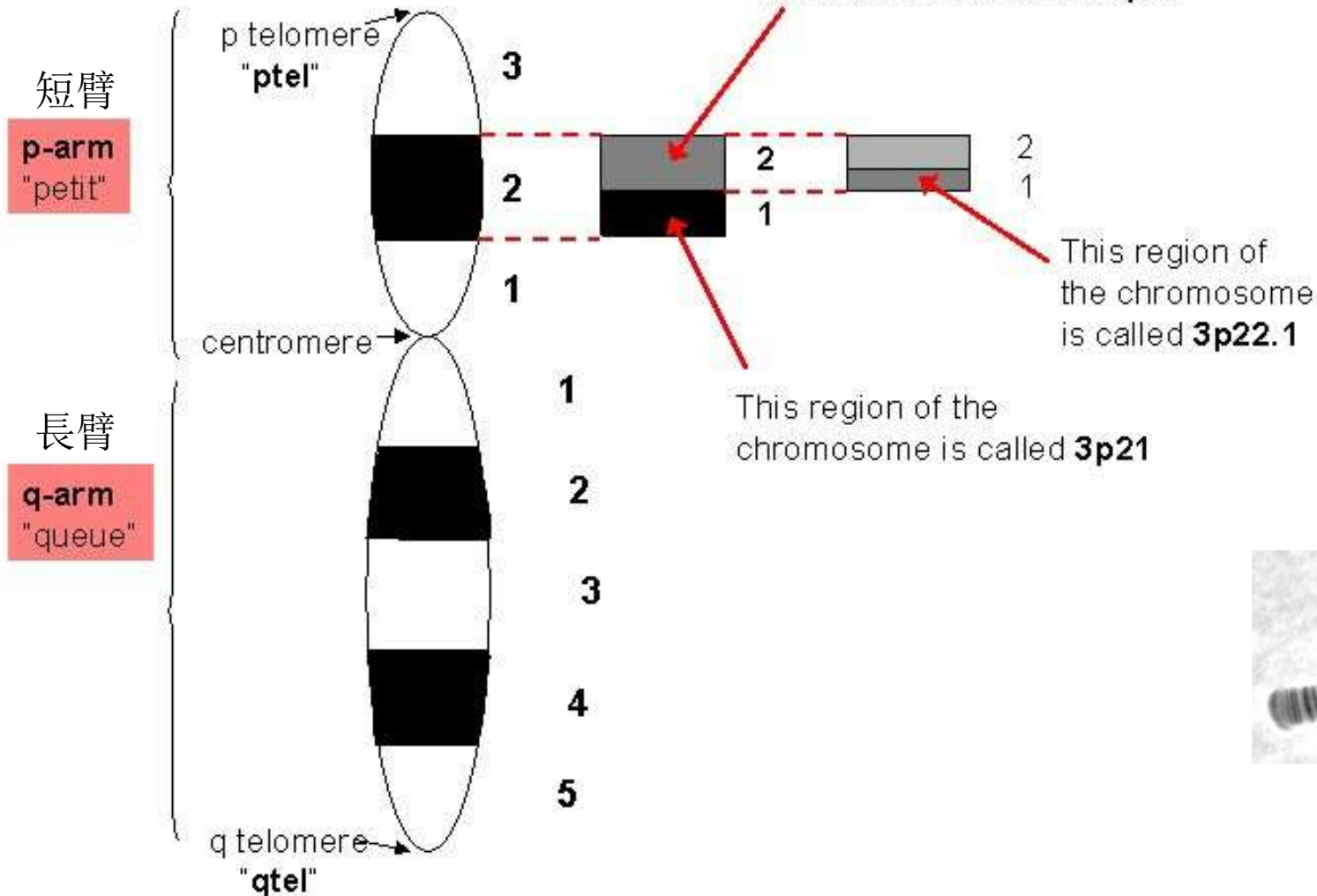
Genetic map (遺傳圖譜): 由互換率計算而來，單位cM (centimorgan)

Physical map (物理圖譜): 由序列定序而來, 單位bp



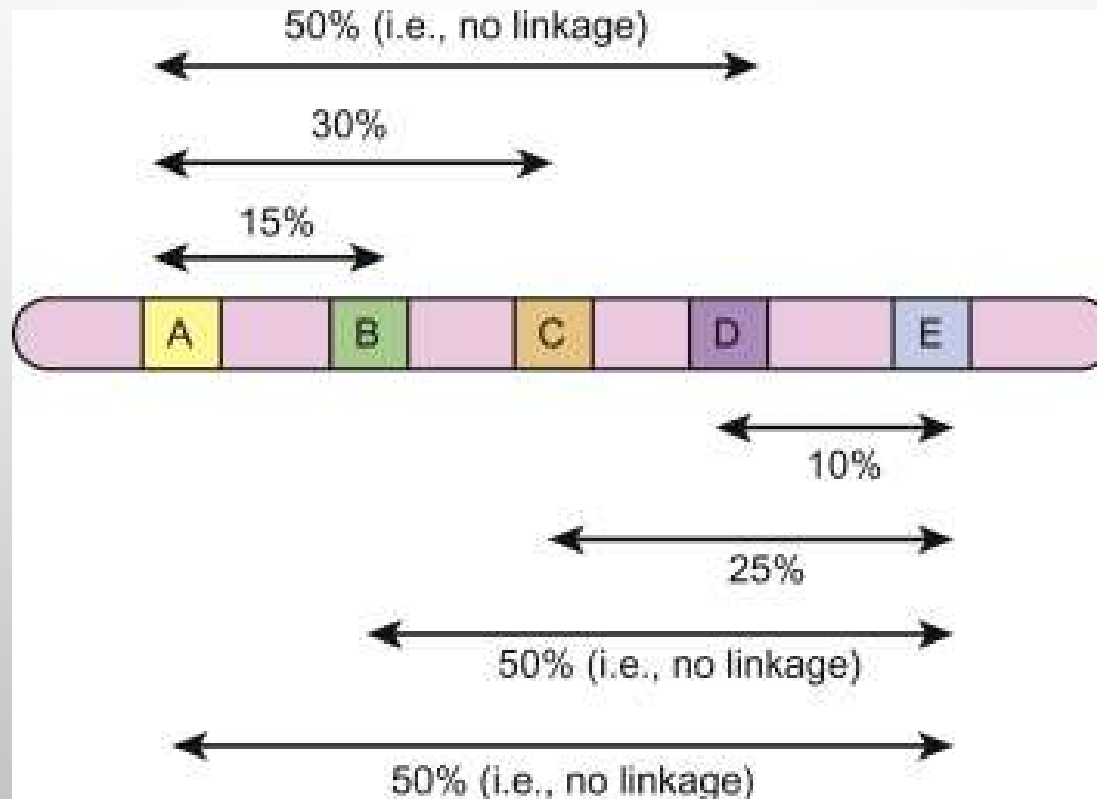
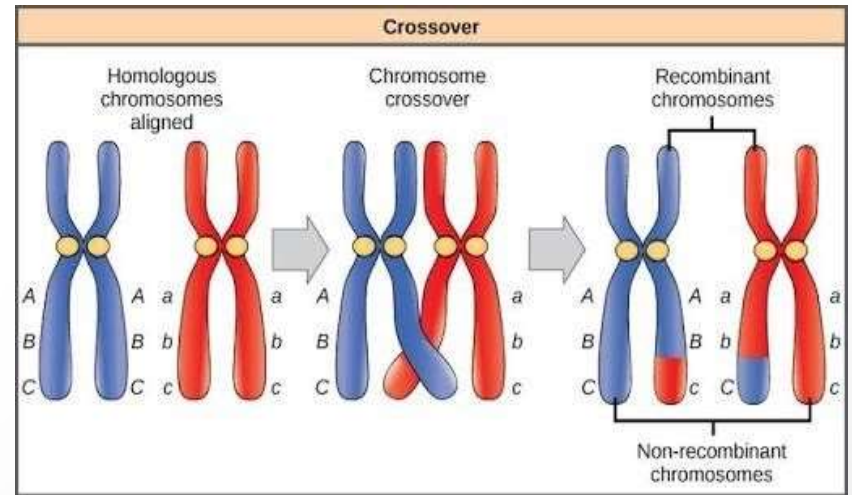
細胞遺傳圖譜 (Cytogenetic map)

Chromosome 3:

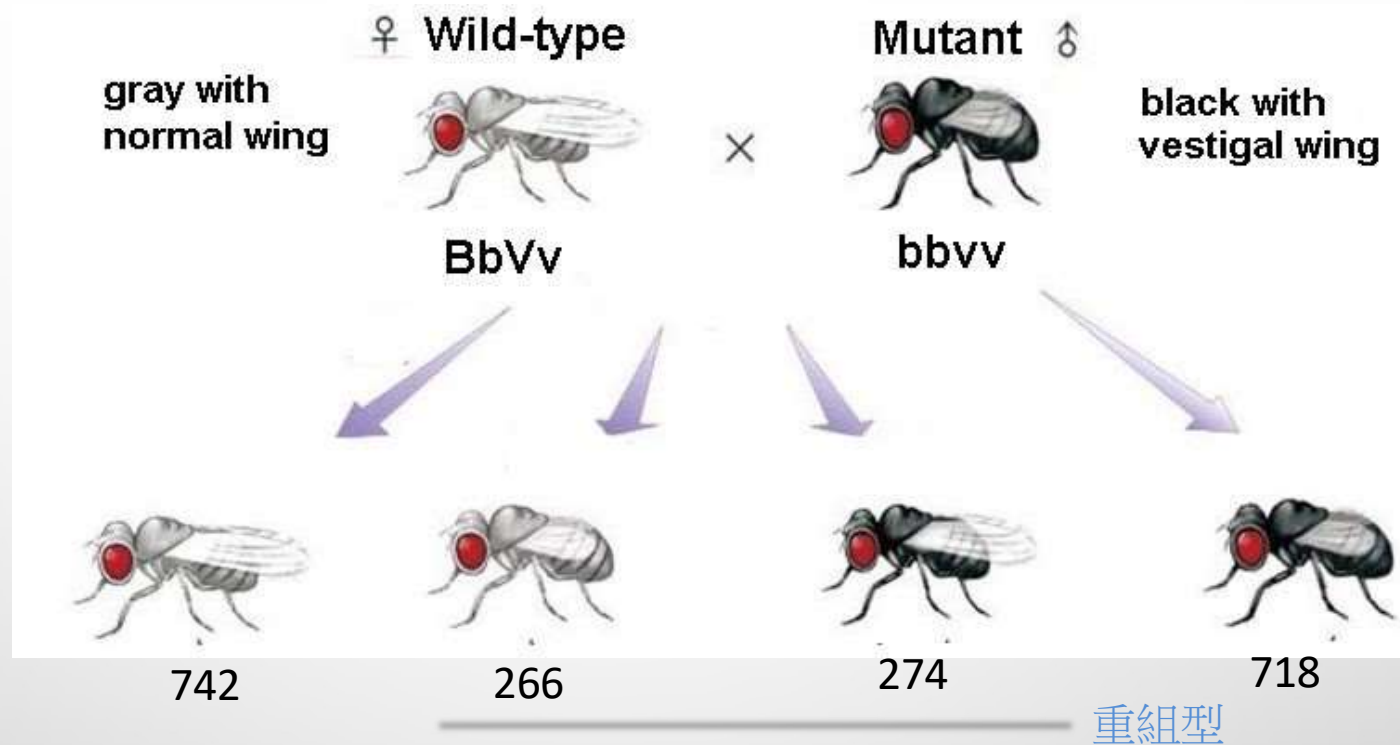


遺傳圖譜 (Genetic map)

- 1% 重組率 = 1 cm (centimorgan)
- 重組率 < 50% --> "連鎖"
- 重組率 = 0% --> "完全連鎖"
- 在人類細胞中 1 cM 約 1 Mb



染色體重組 (CHROMOSOME RECOMBINATION)



B, V 非連鎖基因之期望值

500
1/4

500
1/4

500
1/4

500
1/4

B, V 為完全連鎖之期望值

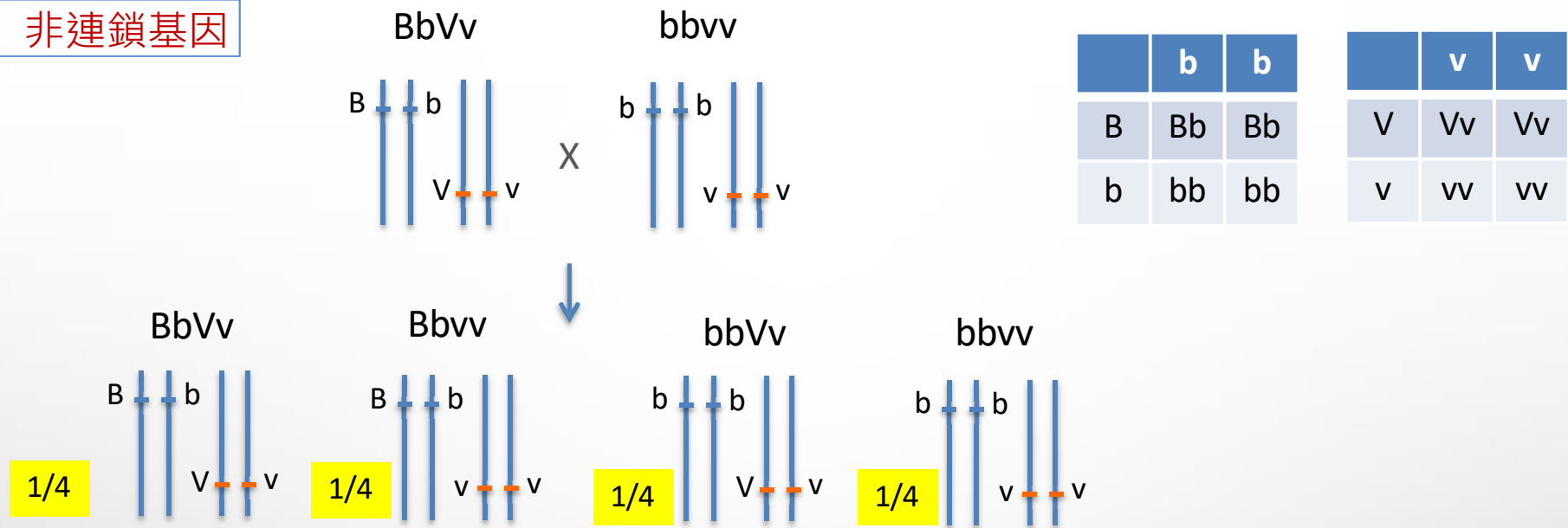
1000
1/2

0

0

1000
1/2

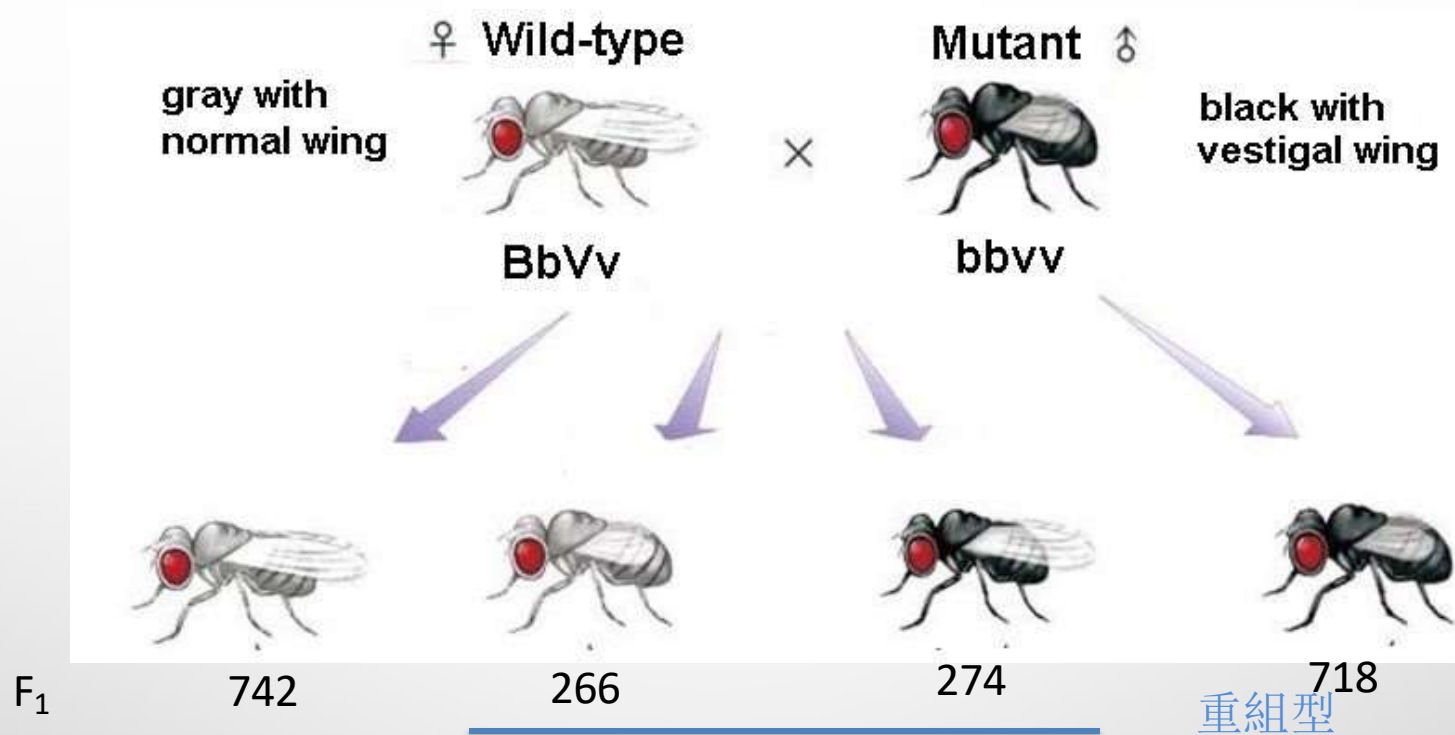
非連鎖基因



基因連鎖



RECOMBINATION RATE (重組率)



重組率 = 重組型 / 所有子代
= (266 + 274) / 2000 = 0.27
兩基因距離為 **27 centimorgan (cM)**

- *重組率最高為50%
- *重組率的精準度和樣品數量成正相關

DNA 載體 (DNA Vector)

載體	承載量	宿主細胞
人類人造染體(HAC) <small>發表於1997年</small>	6000 - 10000 Kb	human cell
酵母人造染色體(YAC) <small>於1983年建立</small>	100 - 3000 kb	Yeast
細菌人造染色體(BAC)	150 ~ 350 kb	E. Coli
噬菌體載體(PAC)	100- 300 kb	E. Coli
黏質體(Cosmid, 噬菌體載體/質體之複合體)	35-45 kb	E. Coli
質體(plasmid)	<= 15kb	E. Coli

如果以一倍的覆蓋率計算，人類基因組(3,200,000 kb) 需要

320 HAC

1,066 YAC

9,142 BAC

10,666 PAC

71,111 Cosmid

213,333 plasmid

* 通常一個基因組庫的要求為6倍覆蓋率以上

19

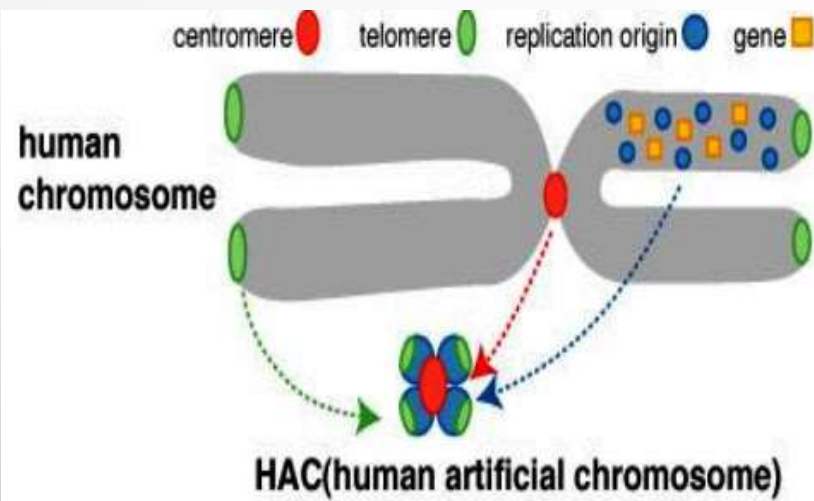


<https://www.youtube.com/watch?v=buAIHv6YOHQ>

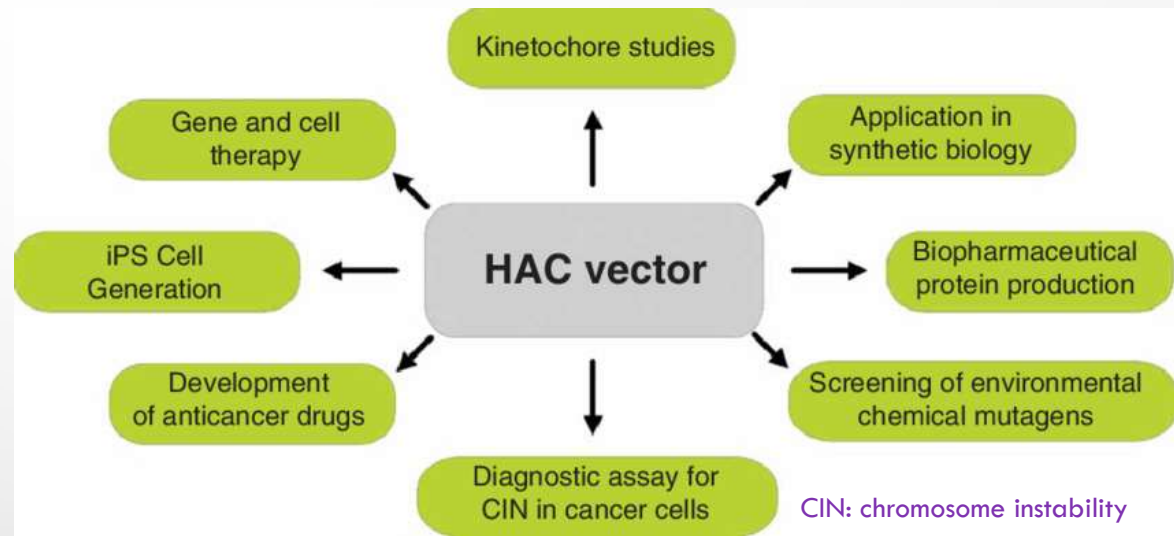
<https://en.wikipedia.org/wiki/Cosmid>

人類人造染色體(Human Artificial Chromosome,HAC)

人類人工染色體是一種小型染色體，可作為載體搭載一些基因，並作為人類細胞中額外的染色體，使這些基因表現於人類體內。



<https://plantbreeding2010.blogspot.com/2022/11/a-brief-account-of-yac-bac-hac.html>



•DOI: [10.1517/17425247.2014.882314](https://doi.org/10.1517/17425247.2014.882314)

HACs have been used in the development of approaches to the reprogramming of cells into iPSCs, creation of transgenic animals, and the generation of experimental models for the treatment of genetic diseases. HACs have been also extensively used to study chromosome functions and chromosomal instability.



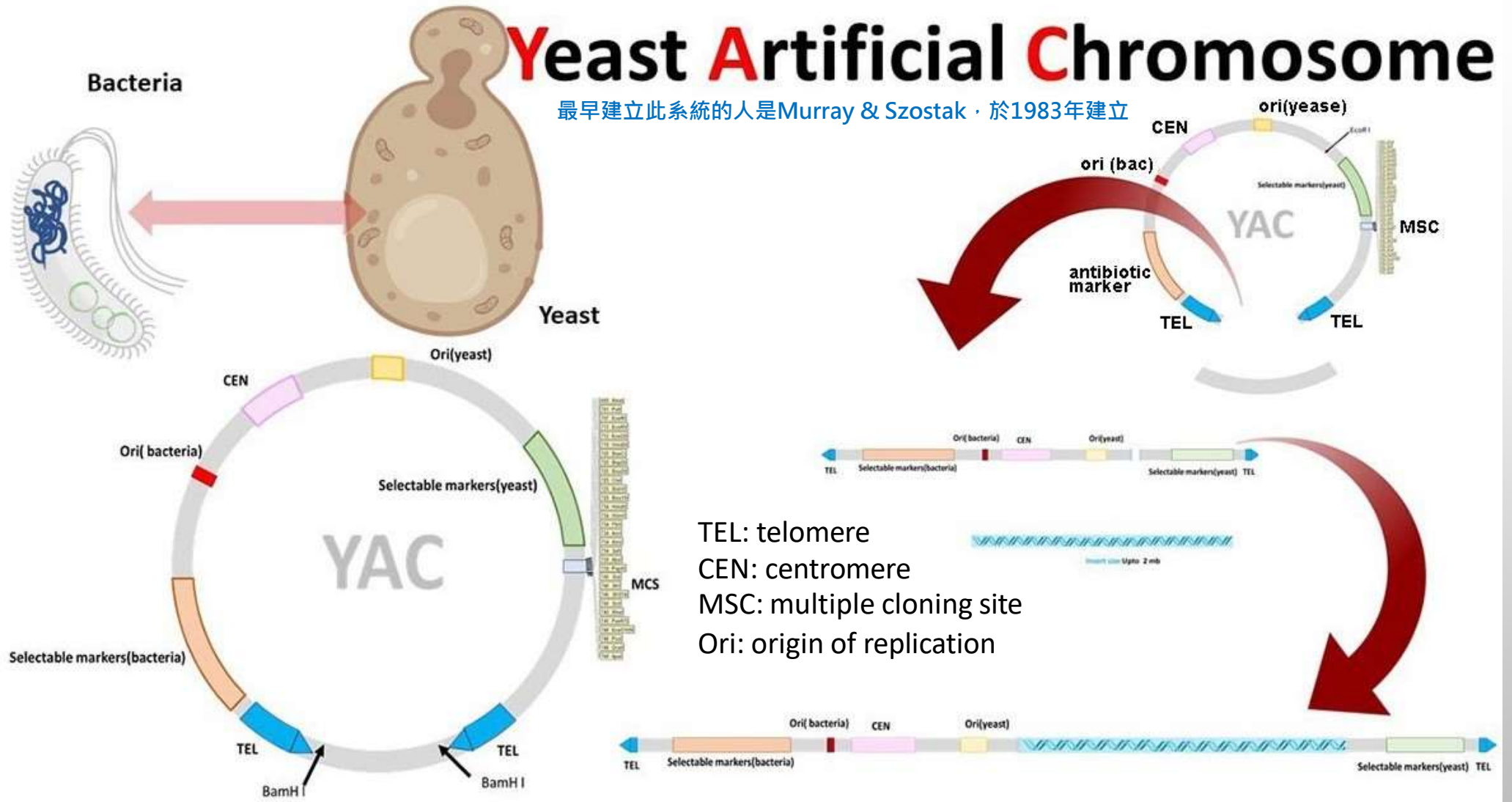
人類人工染色體 成功植入老鼠

<https://www.youtube.com/watch?v=tNQdTzOQtel>

酵母人造染色體(Yeast Artificial Chromosome, YAC)

Yeast Artificial Chromosome

最早建立此系統的人是Murray & Szostak · 於1983年建立



Capacity: 100 -3,000 kb

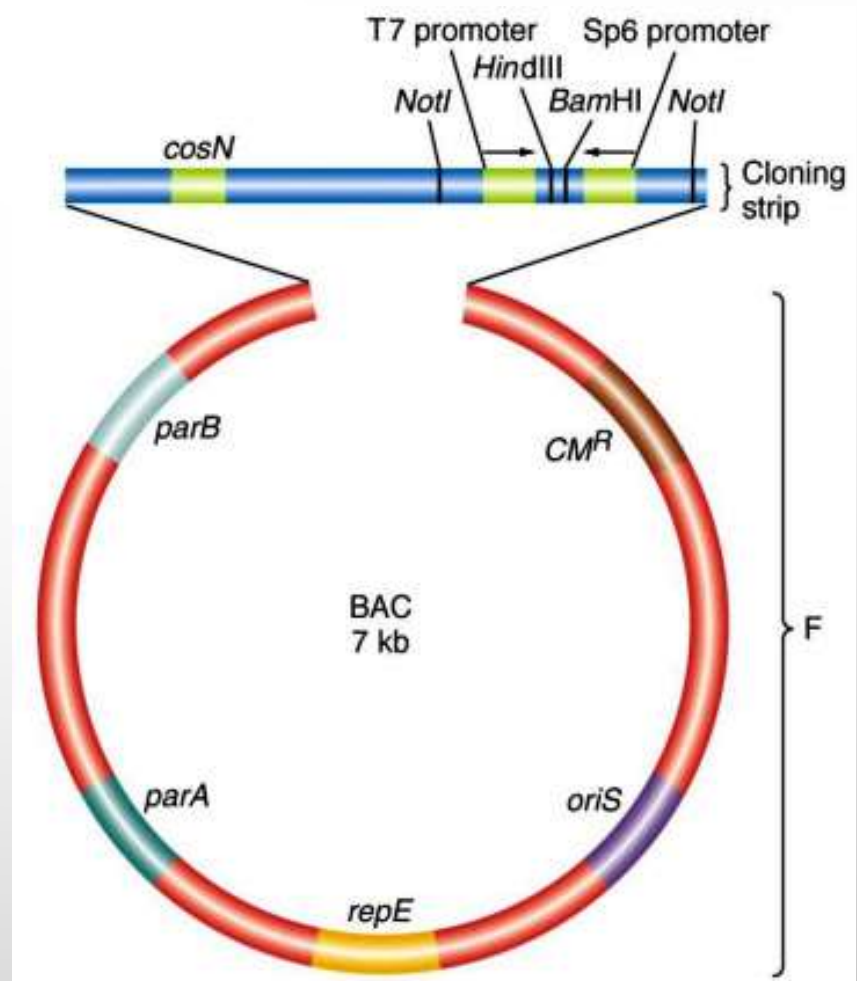
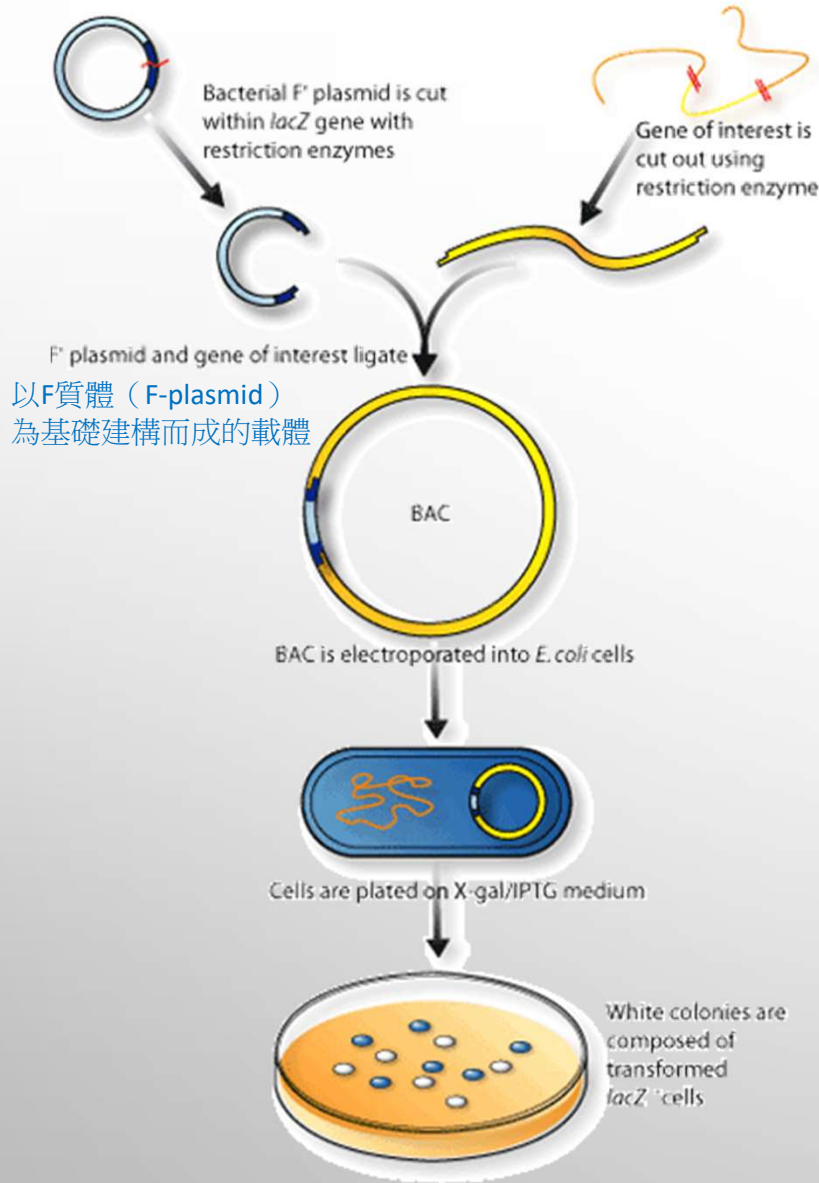


<https://www.youtube.com/watch?v=ZvLPTNYpQX8>

缺點: 其大小與內源的染色體的大小相近，就很難從中分離出來，不利於進一步分析。

優點: 為高容量選殖載體。酵母細胞比大腸桿菌對不穩定的、重複的和極端的 DNA 有更強的容忍性。由於高等真核生物的基因大多數是多外顯子結構並且有長的內含子，大型基因組片段可通過 YAC 載體轉移到動物或動物細胞系中，進行功能研究。

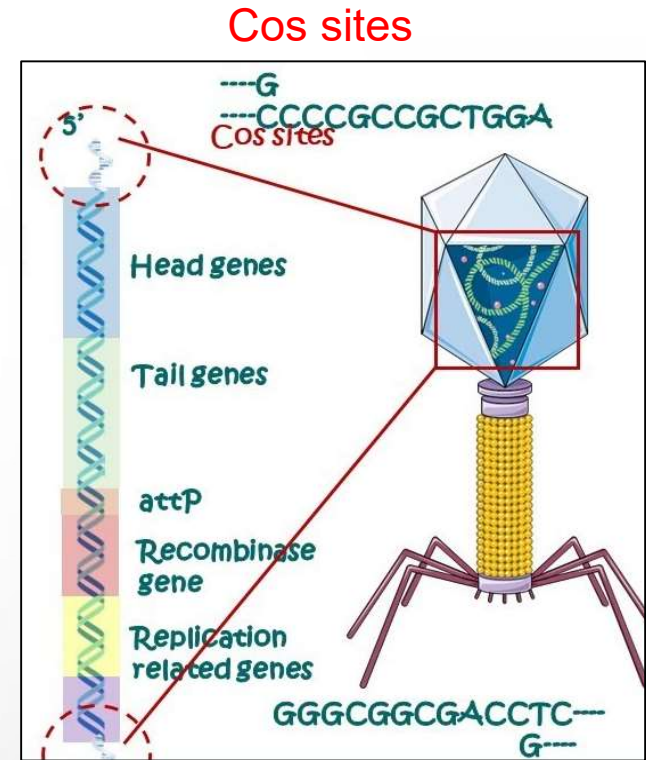
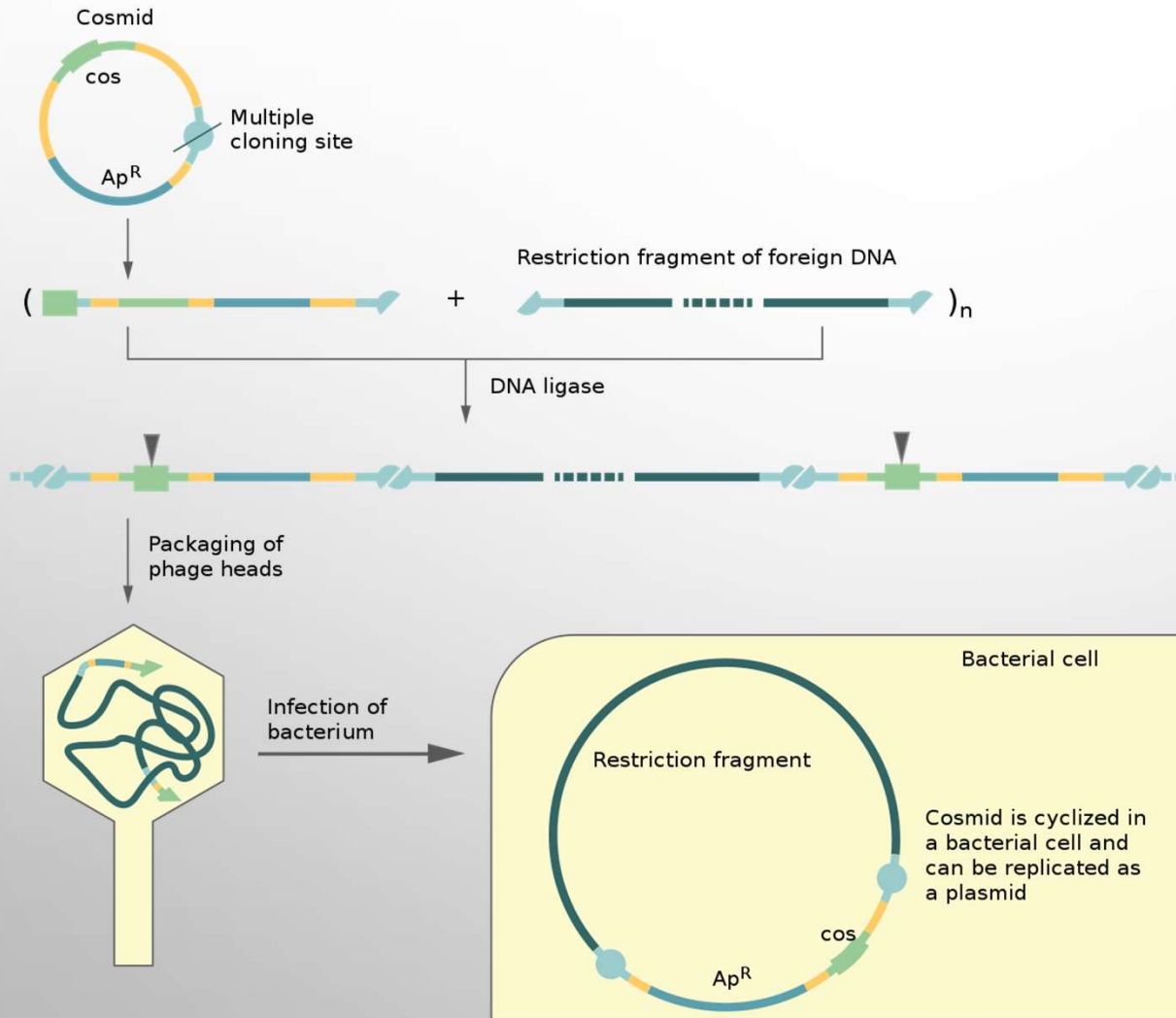
細菌人造染色體(Bacterial Artificial Chromosome, BAC)



<https://plantbreeding2010.blogspot.com/2022/11/a-brief-account-of-yac-bac-hac.html>

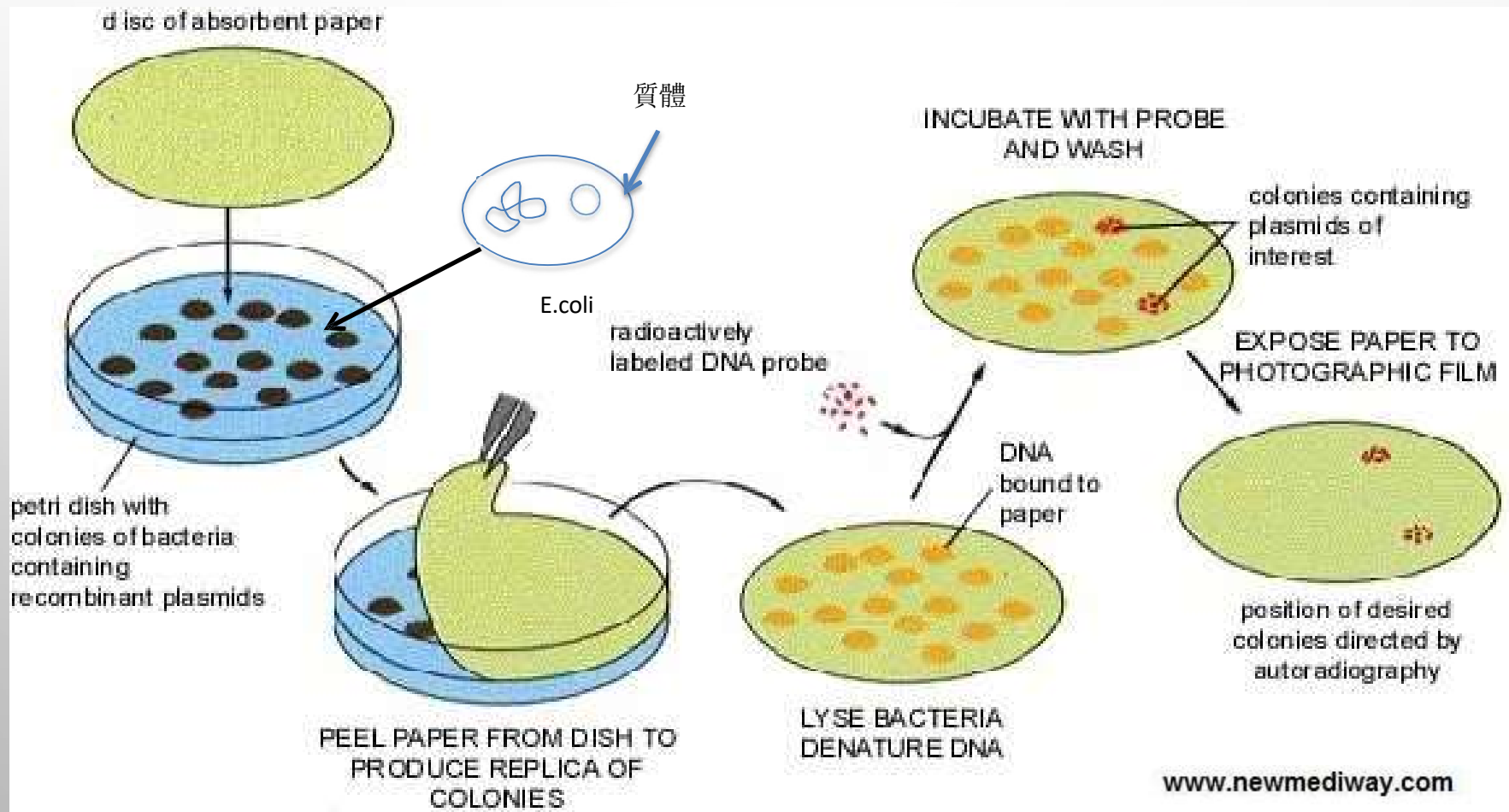
黏質體(Cosmid)

黏質體是一種以λ噬菌體中的cos sequences所建構而成的質體，是常用的選殖載體之一，可用於建構基因組庫。



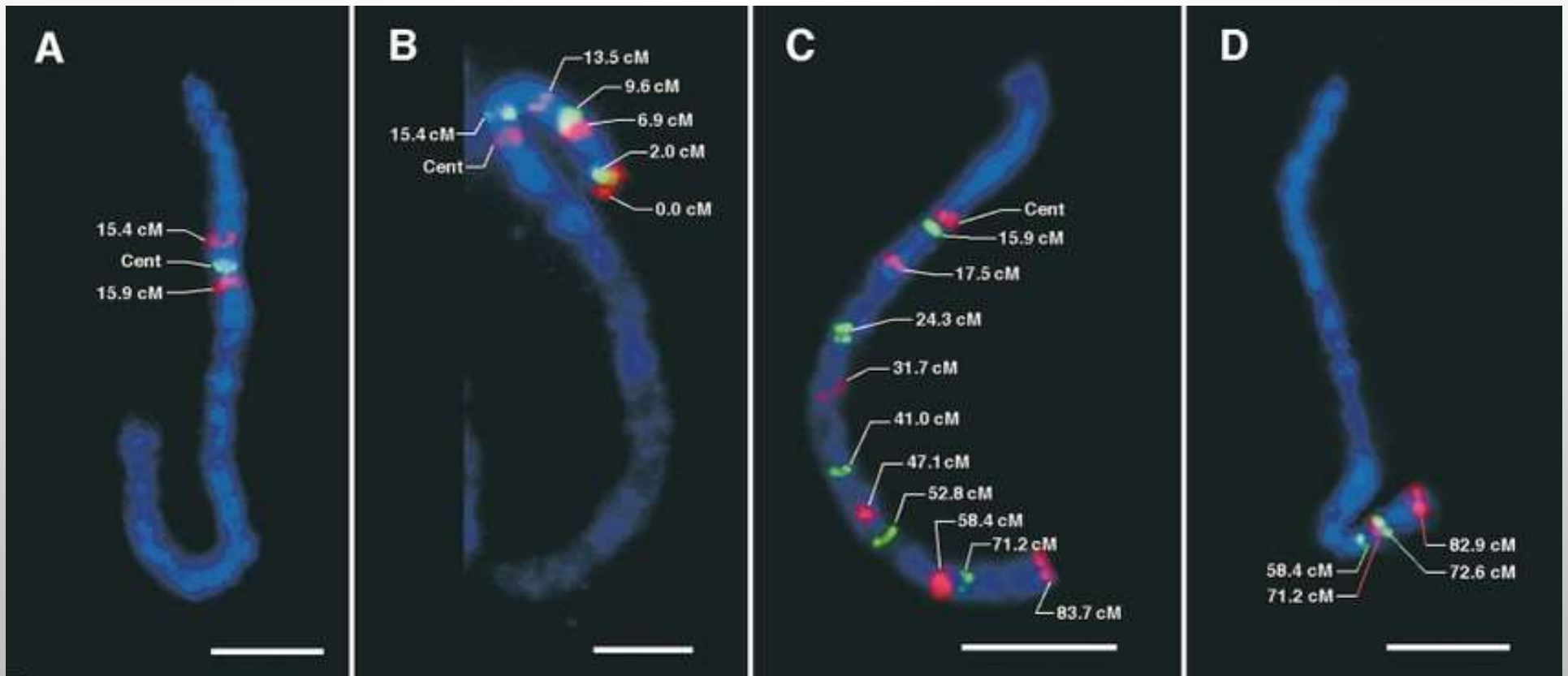
菌落雜交(Colony Hybridization)

- 主要原理為單股DNA可和放射性探針(probe,單股DNA)結合
- 目的為尋找含有特定序列的菌落
- 培養菌落 → 拓印至硝化纖維膜 → 鹼破壞打破細胞並使DNA變性 → 放射性探針雜合 → 訊號偵測



螢光原位雜合FISH (Fluorescence in situ hybridization)

- 主要原理為單股DNA可和螢光標定探針(probe,單股DNA)結合
- 目的為確認目標序列在染色體上的位置
- 細胞固定於玻片→以formamide將染色體變性→螢光標定探針雜合→螢光顯微鏡觀察



History of DNA sequencing

History of DNA sequencing

1869 – Discovery of DNA

1909 – Chemical characterization

1953 – Structure of DNA solved

1977 – Sanger sequencing invented
– Sequencing by degradation (Maxam-Gilbert)
– First genome sequenced – Φ X174 (5 kb)
Enterobacteria phage phiX174

1986 – First automated sequencing machine

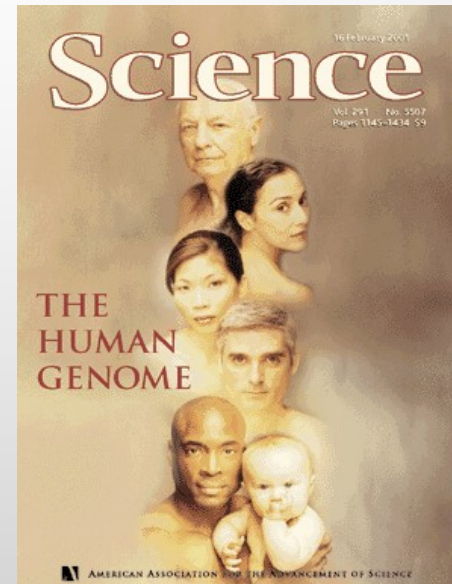
1990 – Human Genome Project started

1992 – First “sequencing factory” at

The Institute for Genomic Research (TIGR)

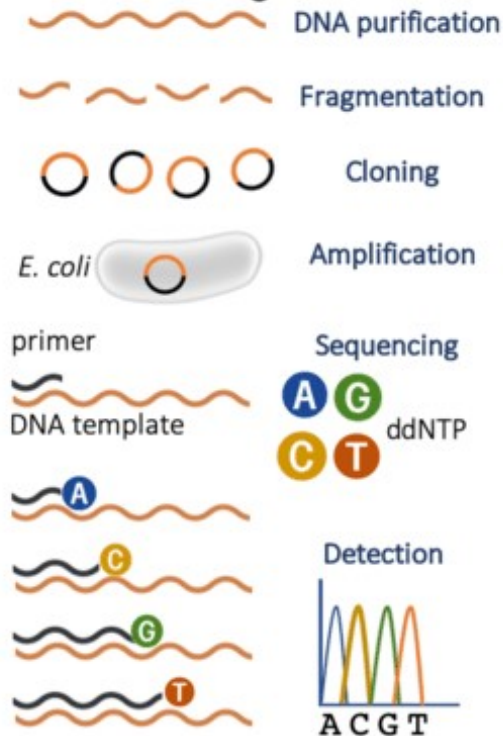


- 1995 – First bacterial genome – *H. influenzae* (1.8 Mb)
- 1996 – Pyrosequencing
- 1998 – First animal genome – *C. elegans* (97 Mb)
- 2003 – Completion of Human Genome Project (3 Gb)
- 2005 – First “next-generation” sequencing instrument
- 2011-The “third generation” sequencer
- 2013– >10,000 genome sequences in NCBI database



First generation sequencing (chain termination)

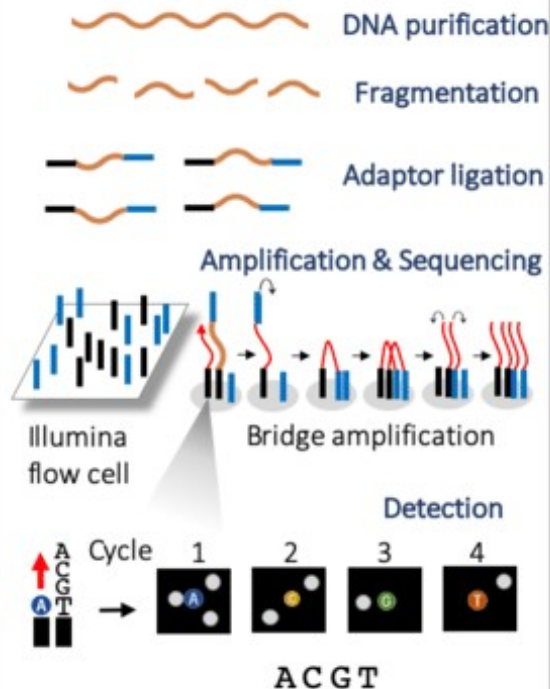
Sanger



- Sequencing by synthesis
- Read length: 300-1000 nucleotides
- Accuracy: >99%
- Real-time analysis: No
- Output data: low output

Second generation sequencing (massively parallel)

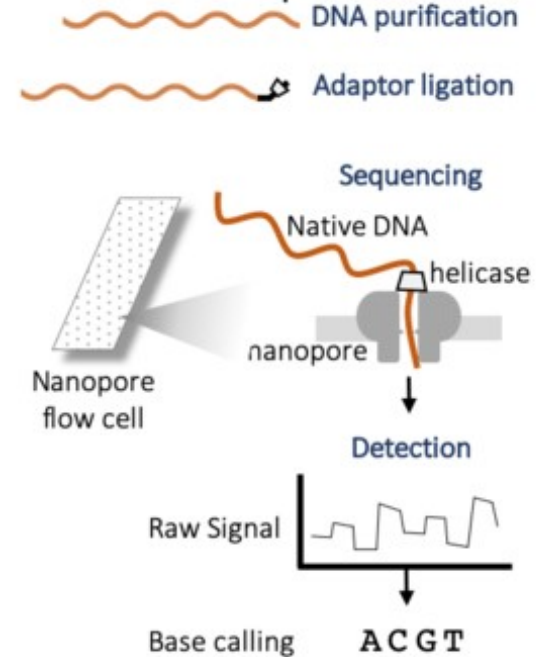
Illumina



- Sequencing by synthesis
- Read length: 36-600 nucleotides
- Accuracy: >99%
- Real-time analysis: No
- Output data: high output

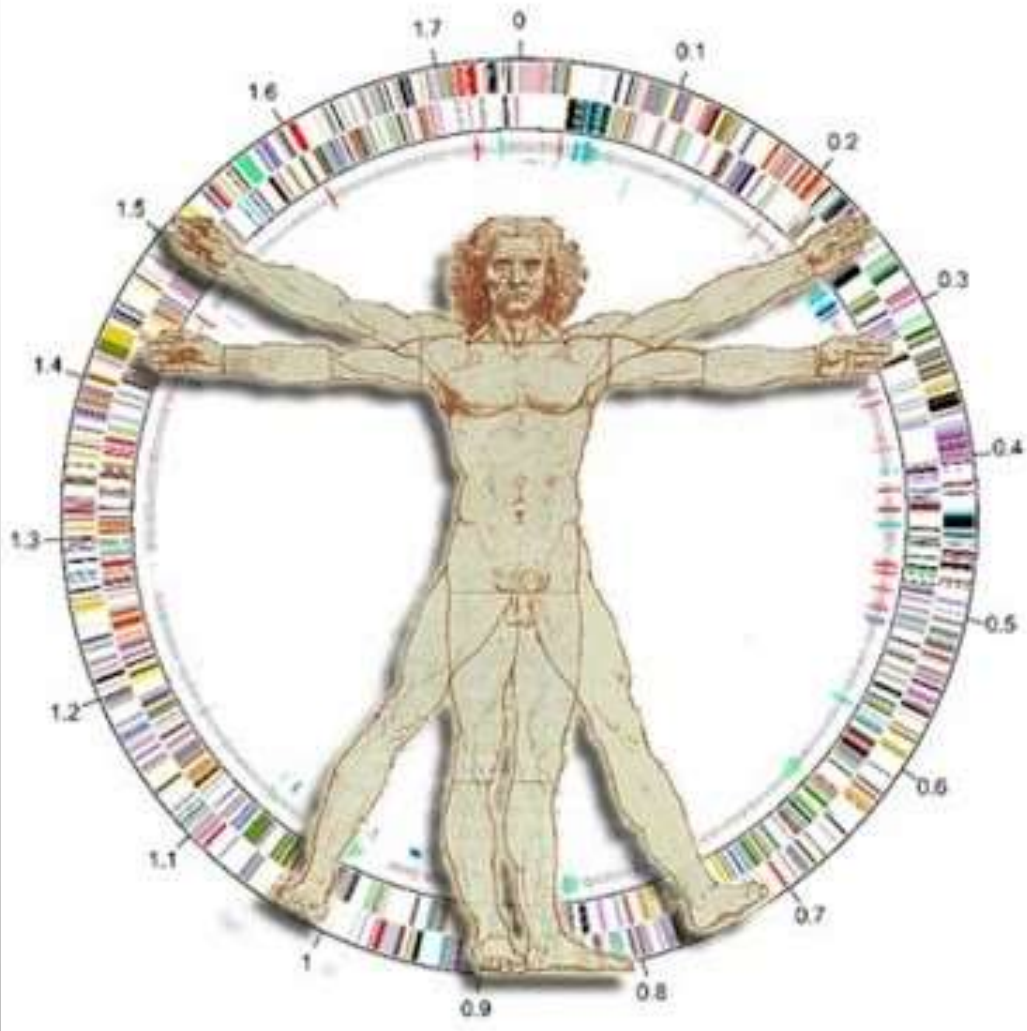
Third generation sequencing (single molecule)

Nanopore



- Single molecule sequencing
- Read length: 200-2 million nucleotides
- Accuracy: ~90-95%
- Real-time analysis: Yes
- Output data: high output

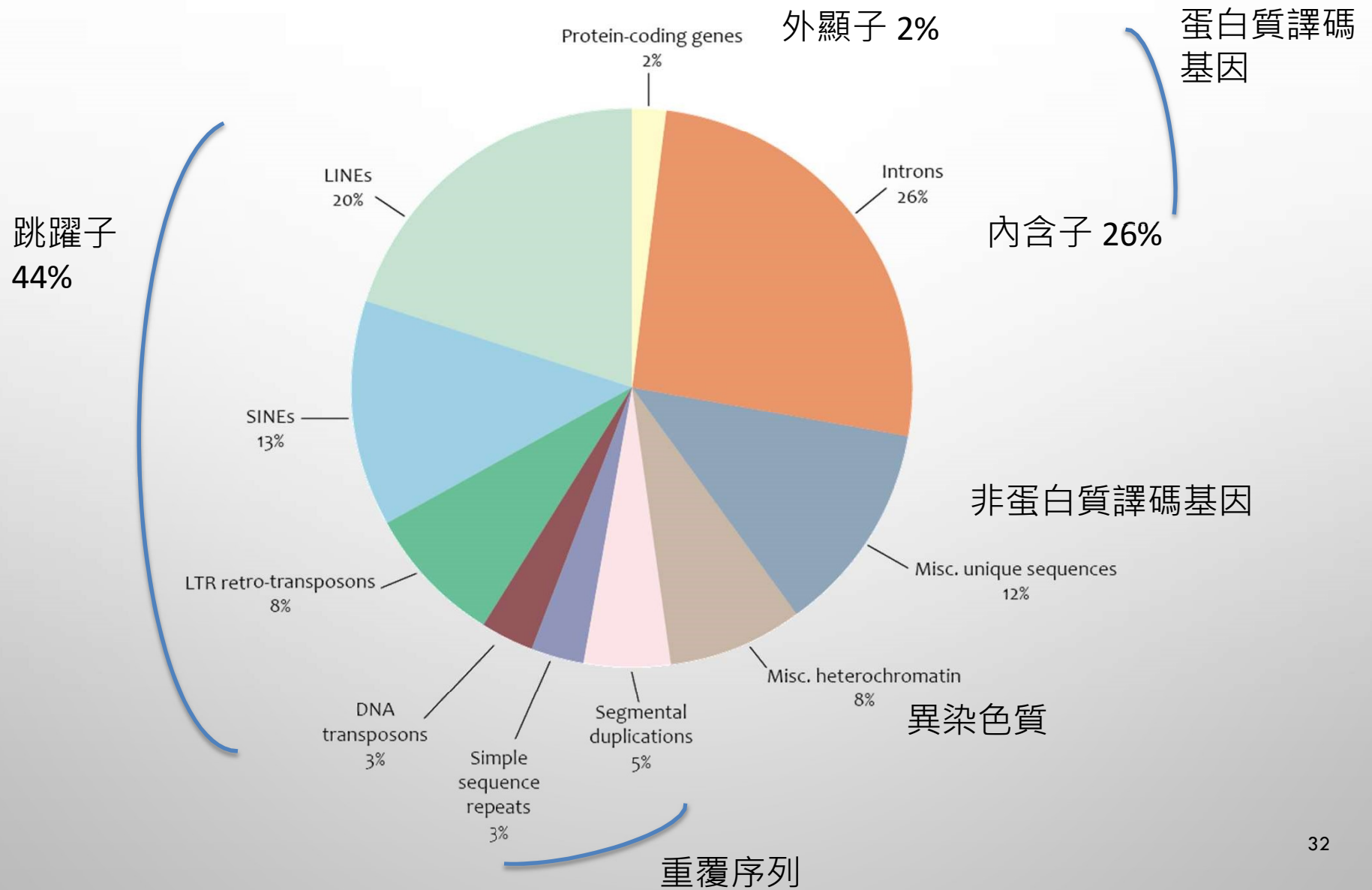
Human genome project (人類基因體計劃)



- 目的：將人類基因組序列完全定序並註解所有基因
- 1990計劃啟動
- 2003公佈草圖
- 總經費\$3,000,000,000美元
- 共18個國家參與

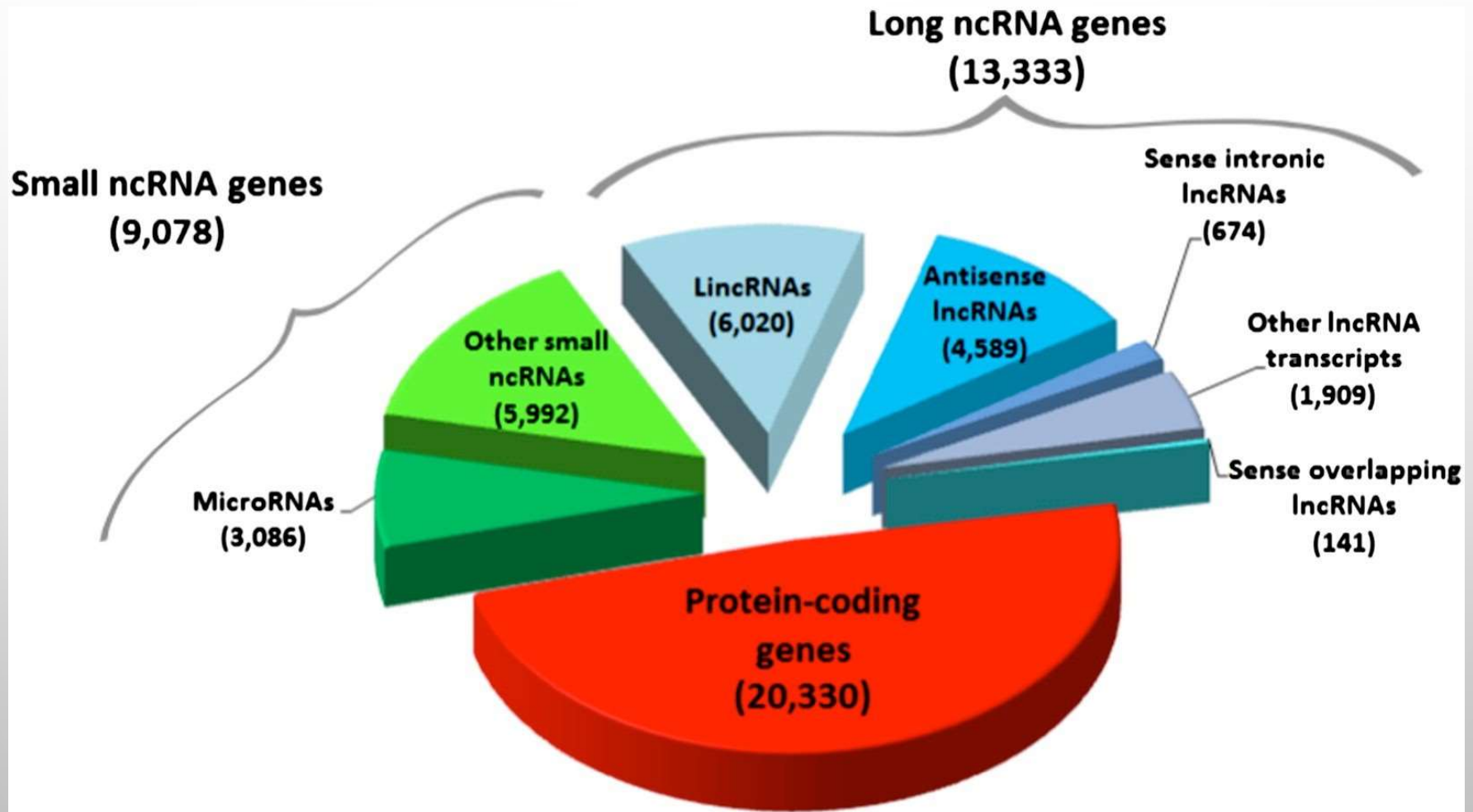
- 1984 – 科學家於美國能源部會議提出構想
- 1986 – 與會科學家再次強調該計劃重要性並討論
- 1988 – 與會科學家一致同意該計劃重要性並準備著手進行
- 1990 - 提出初步構想(為期15年, 經費美金\$3,000,000,000,採用階層式定序法)
- 1992 - 發布低解析度基因組草圖(genome map)
- 1998 – Celera公司宣佈將以霰彈槍定序法於五年內完成基因組定序, 經費\$300,000,000, 完成後將註冊所有基因
- 1999 – 第一條染色體公布(chromosome 22)
- 2000 - Celera公司宣佈已完成~97%
- 2003 - 人類基因組計劃完成(99%)
- 2022 - 人類基因組計劃完成(100%) Science 376.6588 (2022): 44-53.

人類基因體序列分析



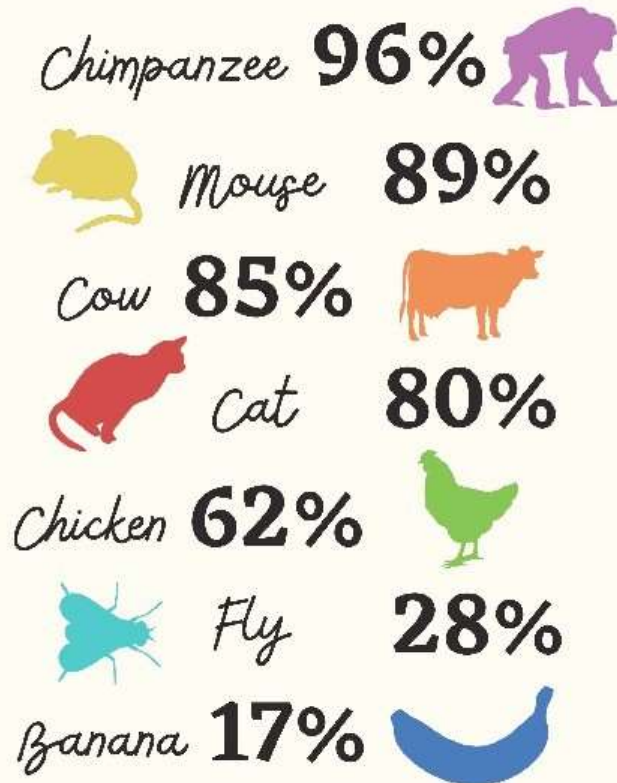
人類基因體基因註解

ncRNAs (ncRNAs genes) versus mRNAs (protein coding genes)



<https://www.researchgate.net/publication/351788403>

What portion of genes do humans share with...?



- the portion of the human genome that contains a unique ortholog in the comparison species, considering only protein coding genes
- calculated using `omadb`, a python wrapper for the OMA database
- inspired by Natasha Glover's blog post 'The Banana Conjecture'

全基因组定序(Whole Genome Sequencing)

1 Break genome into large fragments and clone

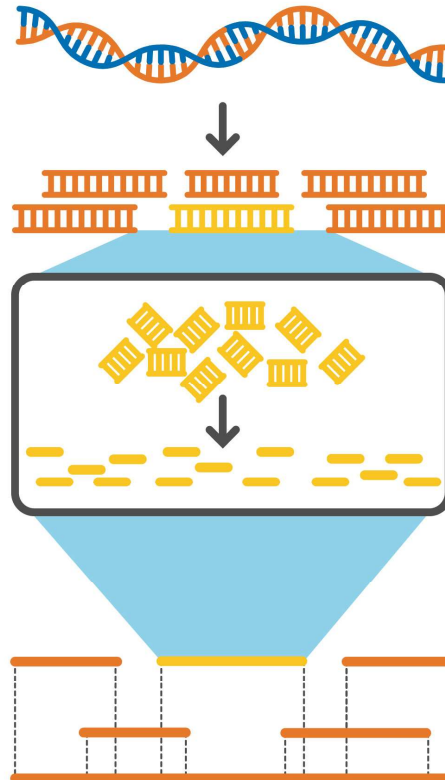
2 Break individual clone into small fragments

3 Generate thousands of sequence reads

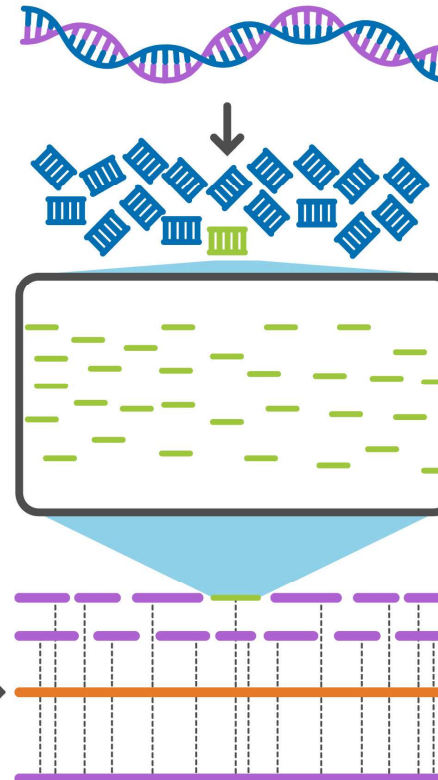
4 Assemble sequence reads for each clone

Reference genome

Reference Genome



Individual Genome



1 Break genome into small fragments

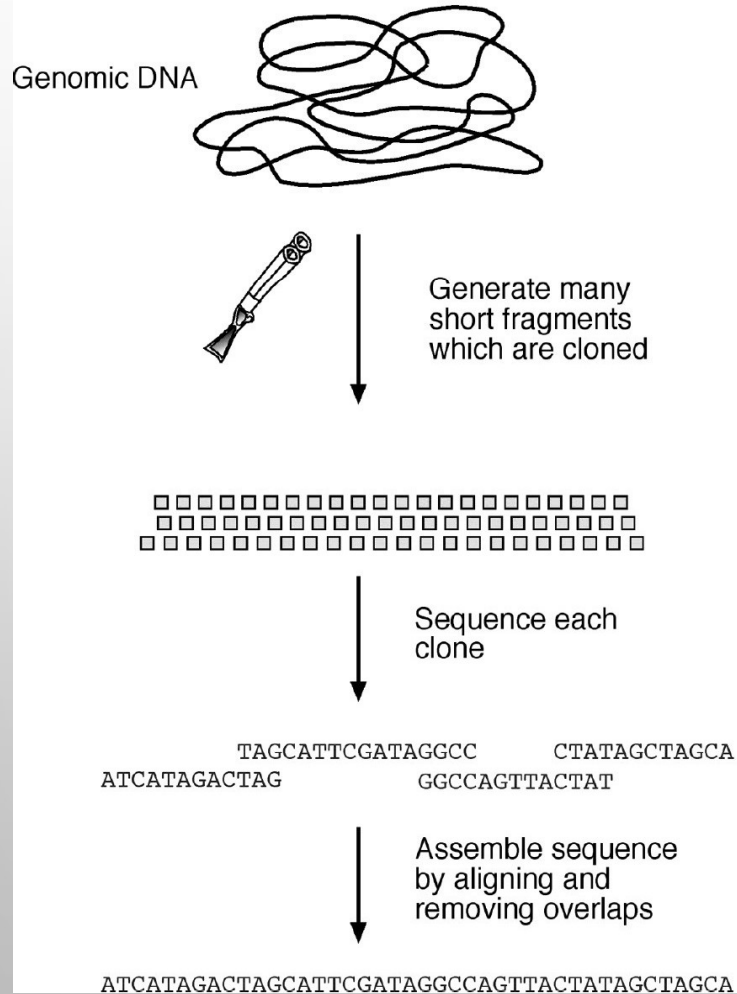
2 Generate millions of sequence reads

3 Align sequence reads into a reference genome

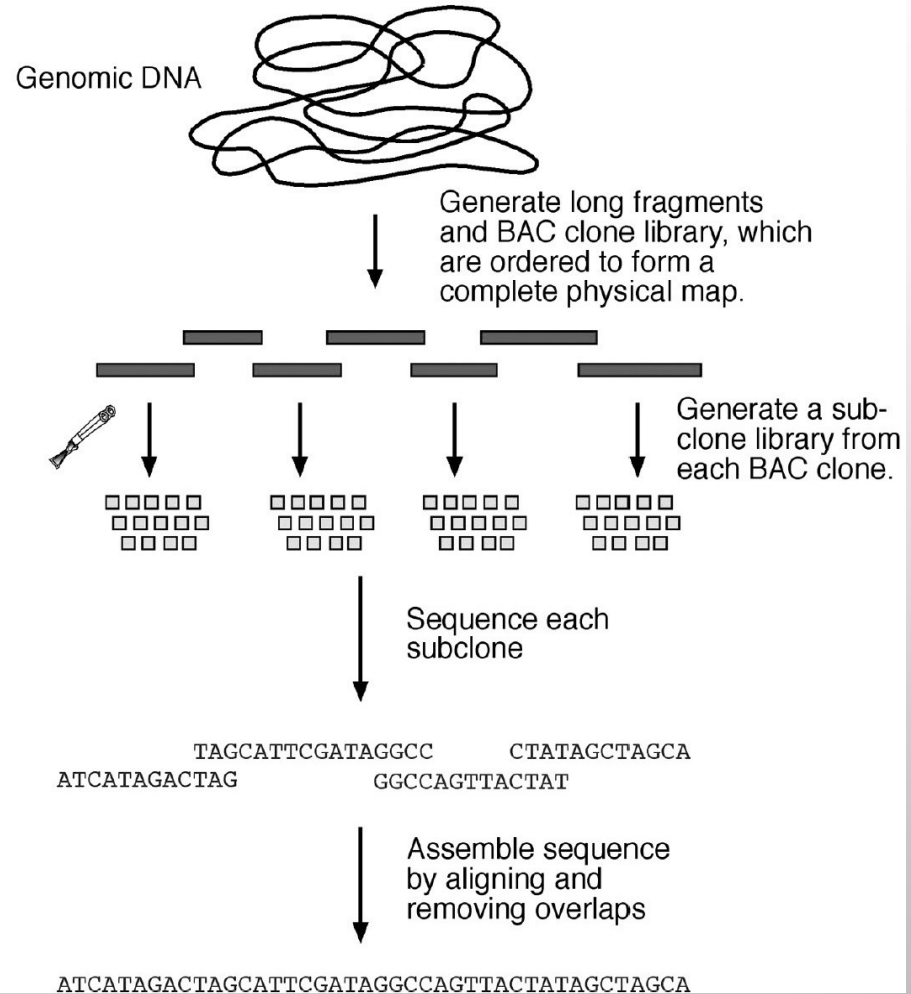
Individual genome

基因組定序策略(Genome sequencing strategies)

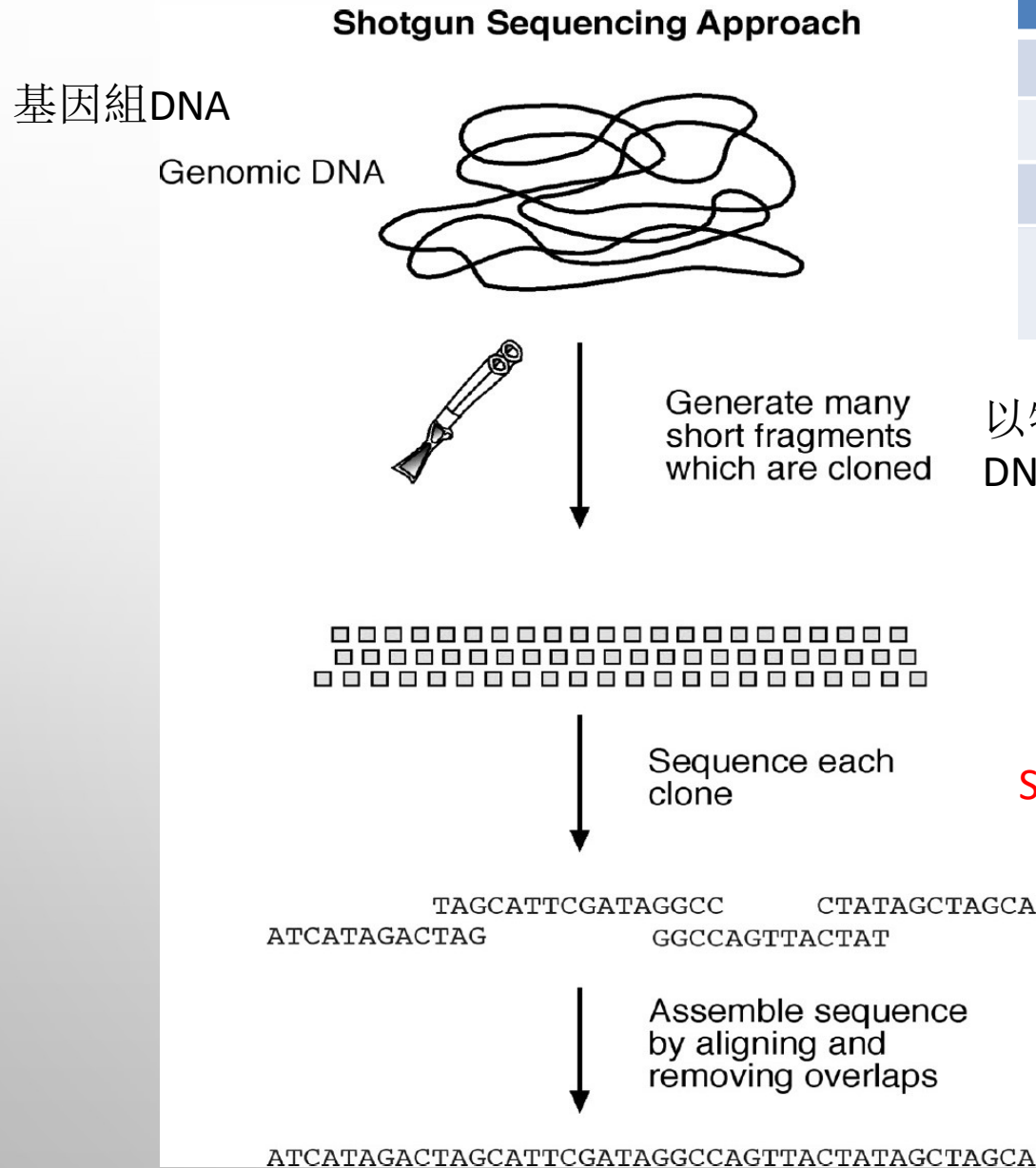
Shotgun sequencing approach (霰彈槍定序法)



Hierarchical sequencing approach (階層式定序法)



Shotgun sequencing approach (霰彈槍定序法)



	霰彈式定序
時間	短
經費	少
人力	少
空缺區域 (gap)	大

以物理、化學或酵素法將DNA打斷~500 bp

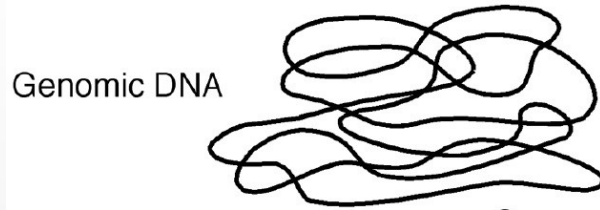
Sanger 或次世代定序

基因組序列組裝

Hierarchical sequencing approach (階層式定序法)

	階層式定序
時間	長
經費	多
人力	多
空缺區域(gap)	小

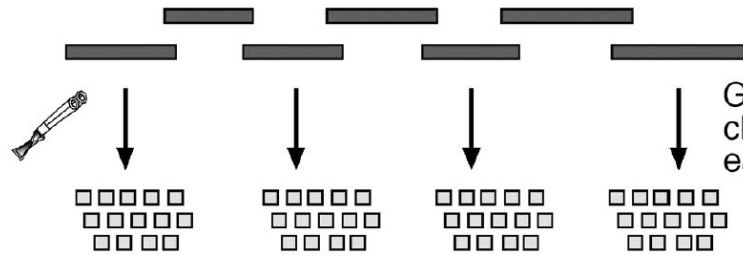
Hierarchical Sequencing Approach



Generate long fragments and BAC clone library, which are ordered to form a complete physical map.

以物理、化學或酵素法將DNA打斷 ~ 150-300 Kb, 並接在 YAC or BAC

將BAC以FISH定位在染色體



將BAC (150 kb) 片段亞克隆至質體 (plasmid, 2 kb)

以sanger定序質體DNA

Sequence each subclone

```

TAGCATTCGATAGGCC      CTATAGCTAGCA
ATCATAGACTAG          GGCCAGTTACTAT
    
```

Assemble sequence by aligning and removing overlaps

序列組裝

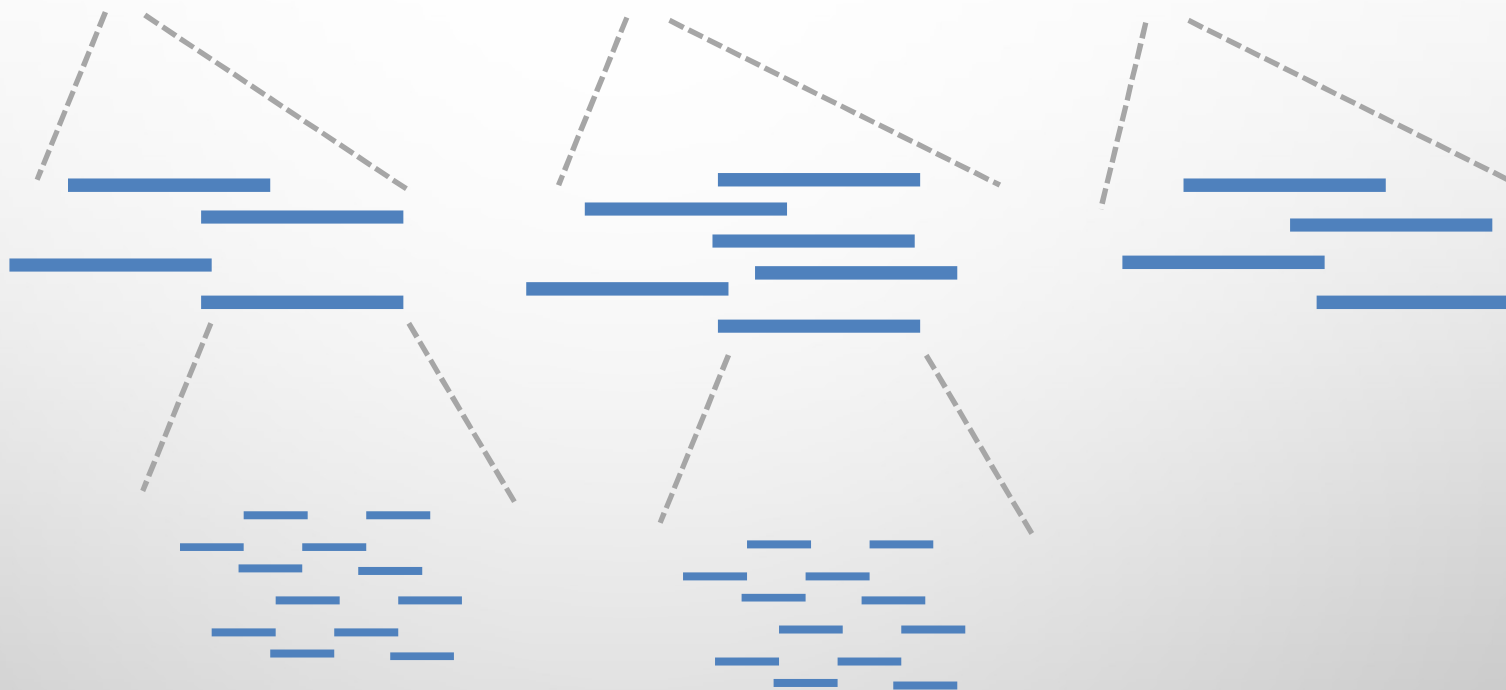
```

ATCATAGACTAGCATTCGATAGGCCAGTTACTATAGCTAGCA
    
```

基因組
DNA

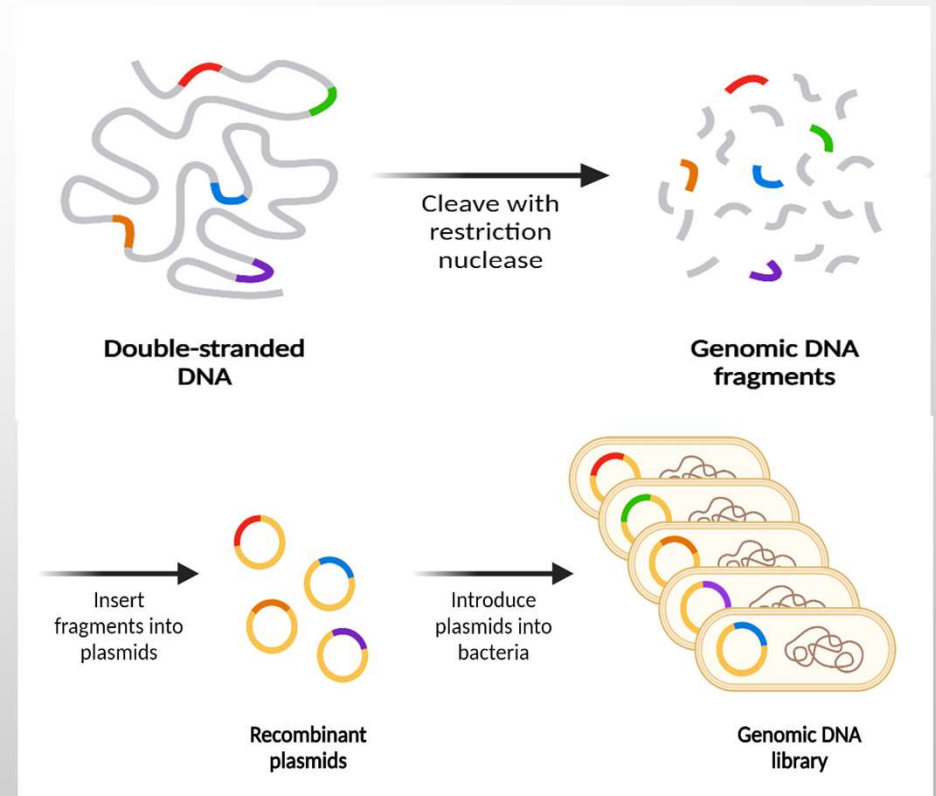
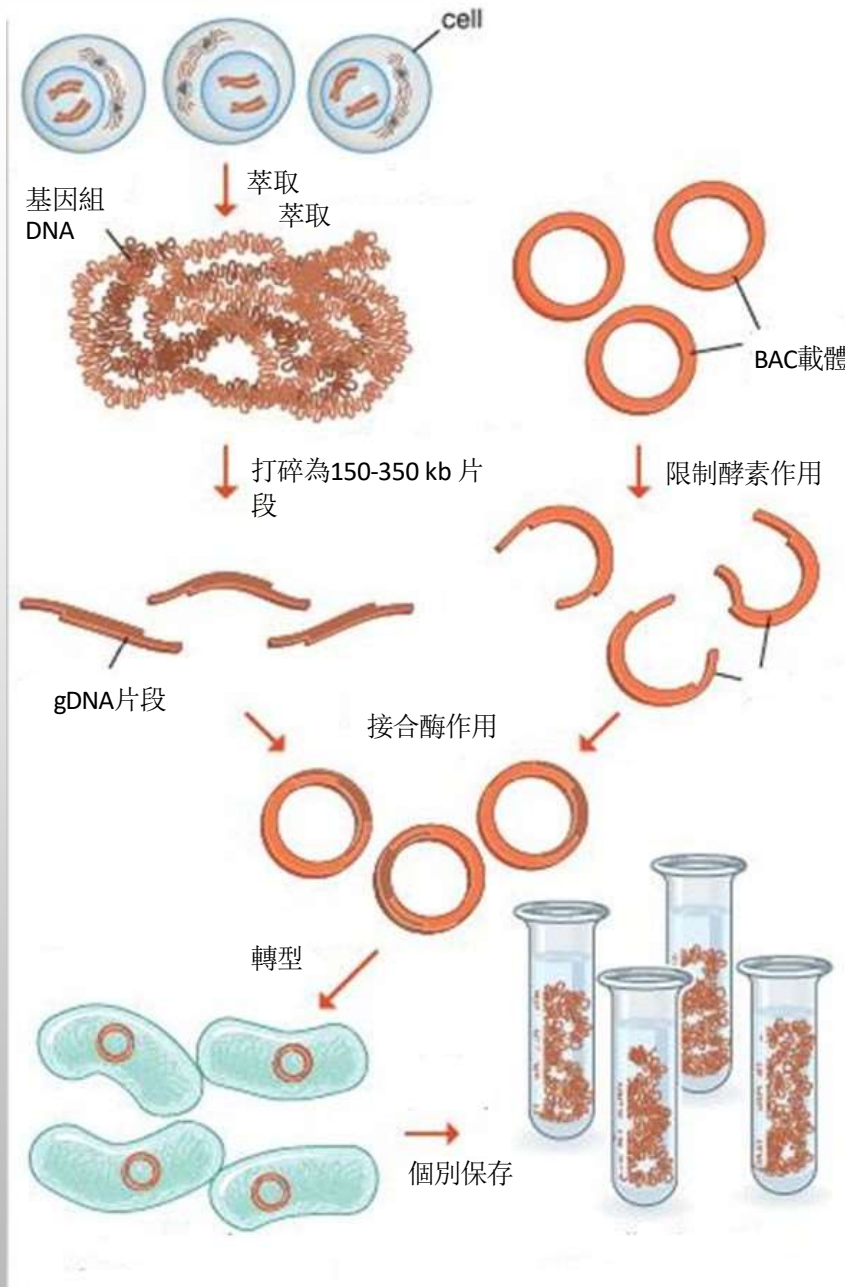


BAC | YAC
基因庫



質體
基因庫

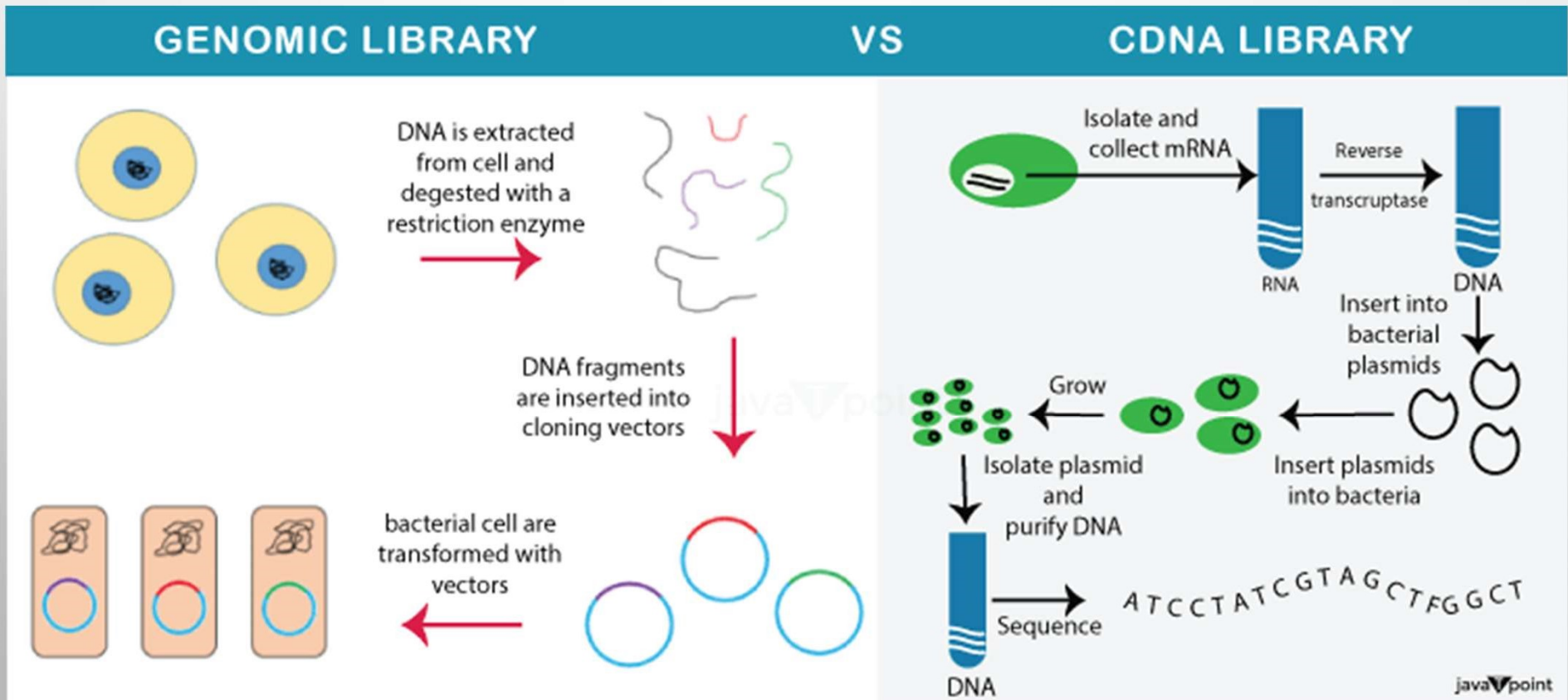
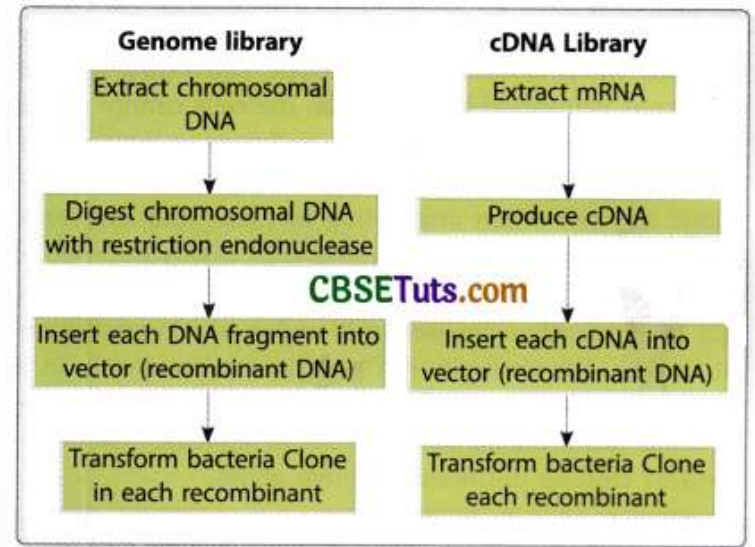
基因組庫建構 (Genomic DNA library construction)



Genomic DNA library

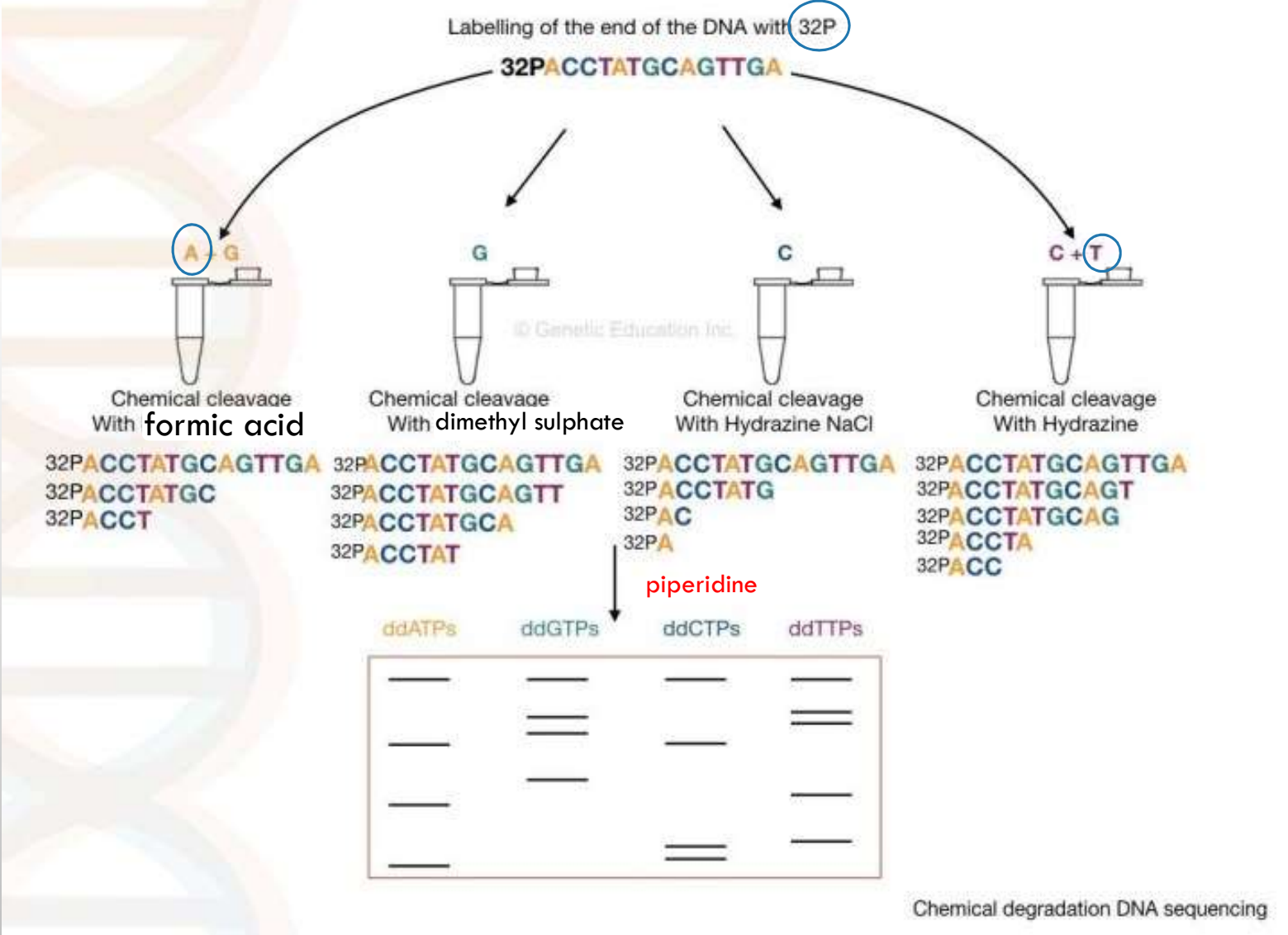
VS

cDNA library



The first-generation DNA sequencing

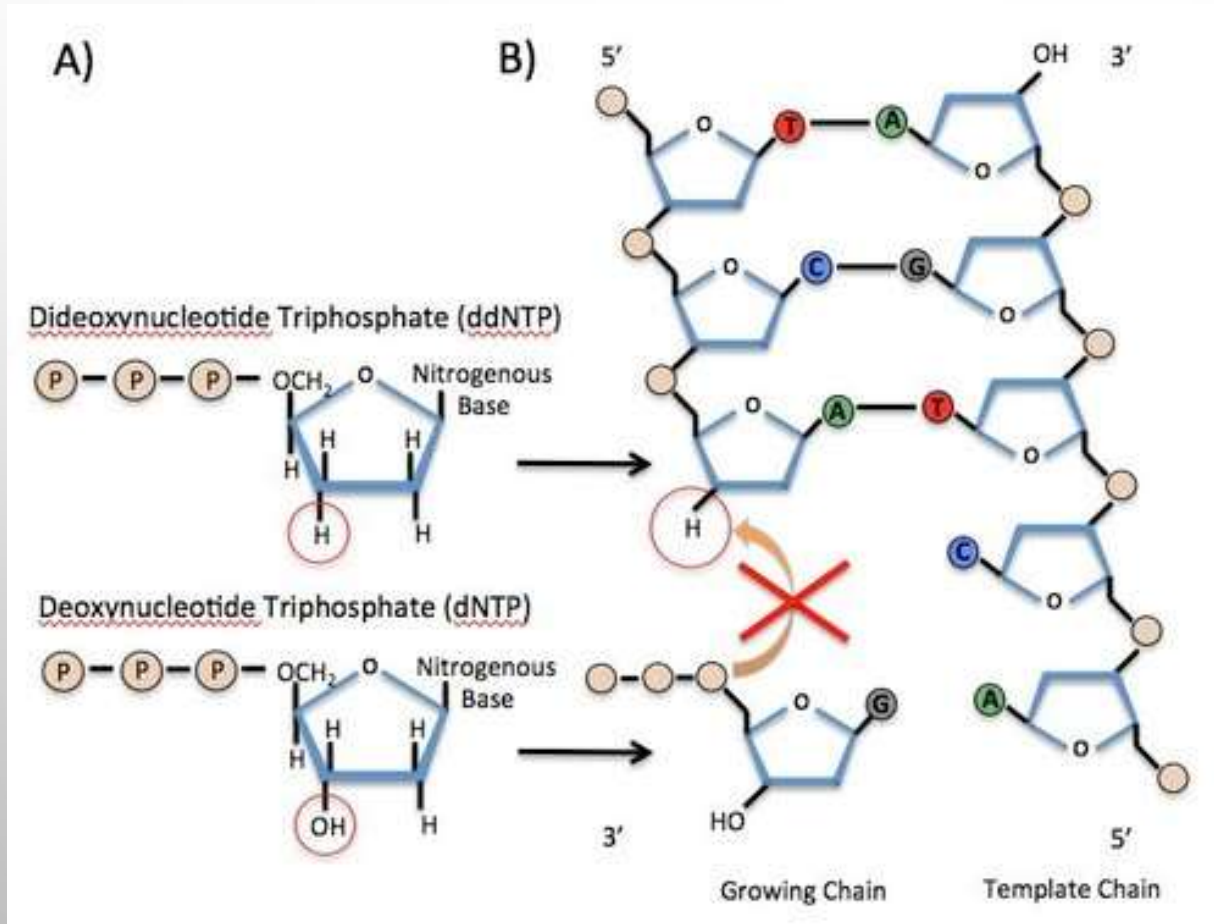
(1) Maxam Gilbert DNA sequencing



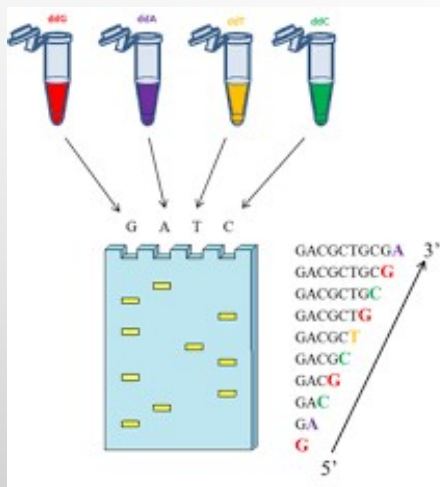
https://www.youtube.com/watch?v=_B5Dj8PL4E0

(2) SANGER SEQUENCING

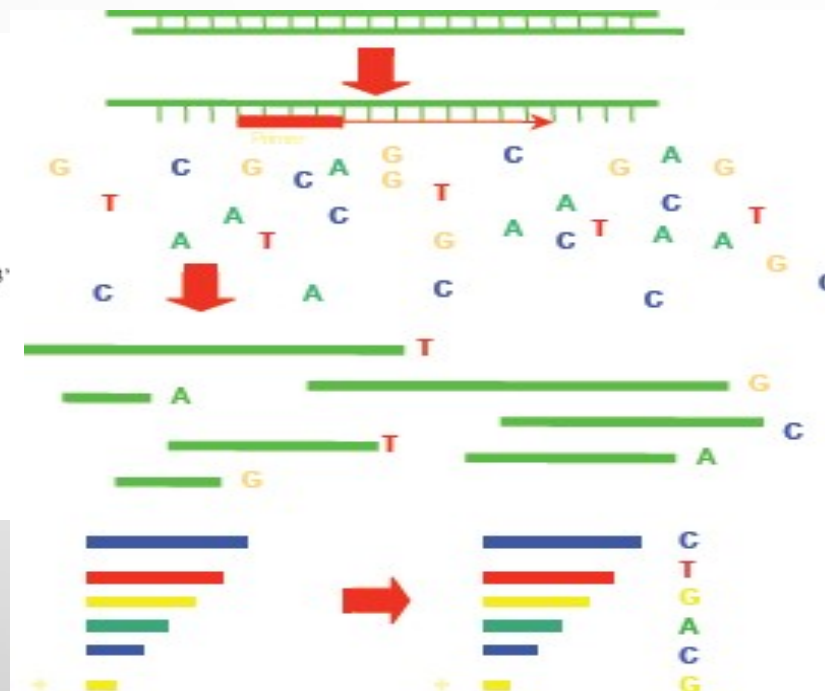
- Uses DNA polymerase
- All **four nucleotides**, plus one dideoxynucleotide (**ddNTP**)
- **Random termination** at specific bases
- Separate by gel electrophoresis



1986: 4 Reactions to 1 Lane
fluorescently labelled ddNTPs



Sequencing Reaction Products



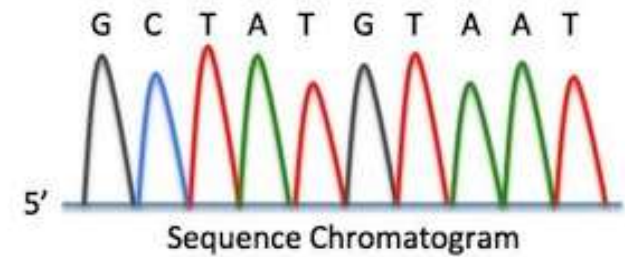
Progression of Sequencing Reaction

Template Sequence

3' GAGCAAATTCGATACATTATTGT... 5'
Primer

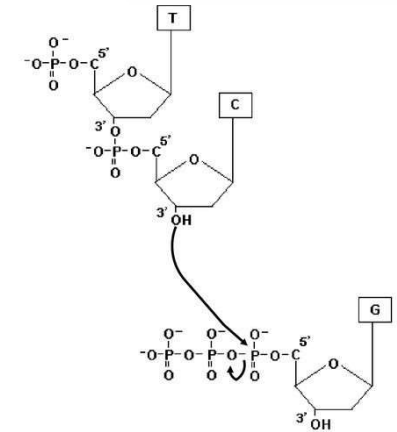
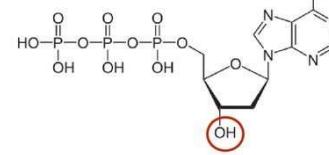
5' CTCGTTTAAG... 3'

CTCGTTTAAGG — G
CTCGTTTAAGGC — C
CTCGTTTAAGGGT — T
CTCGTTTAAGGGTA — A
CTCGTTTAAGGGTAT — T
CTCGTTTAAGGGTATG — G
CTCGTTTAAGGGTATGT — T
CTCGTTTAAGGGTATGTA — A
CTCGTTTAAGGGTATGTAA — A
CTCGTTTAAGGGTATGTAAT — T



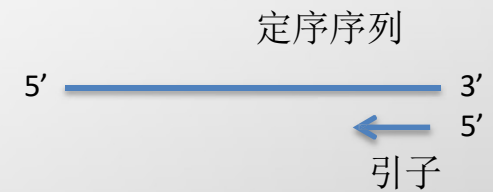
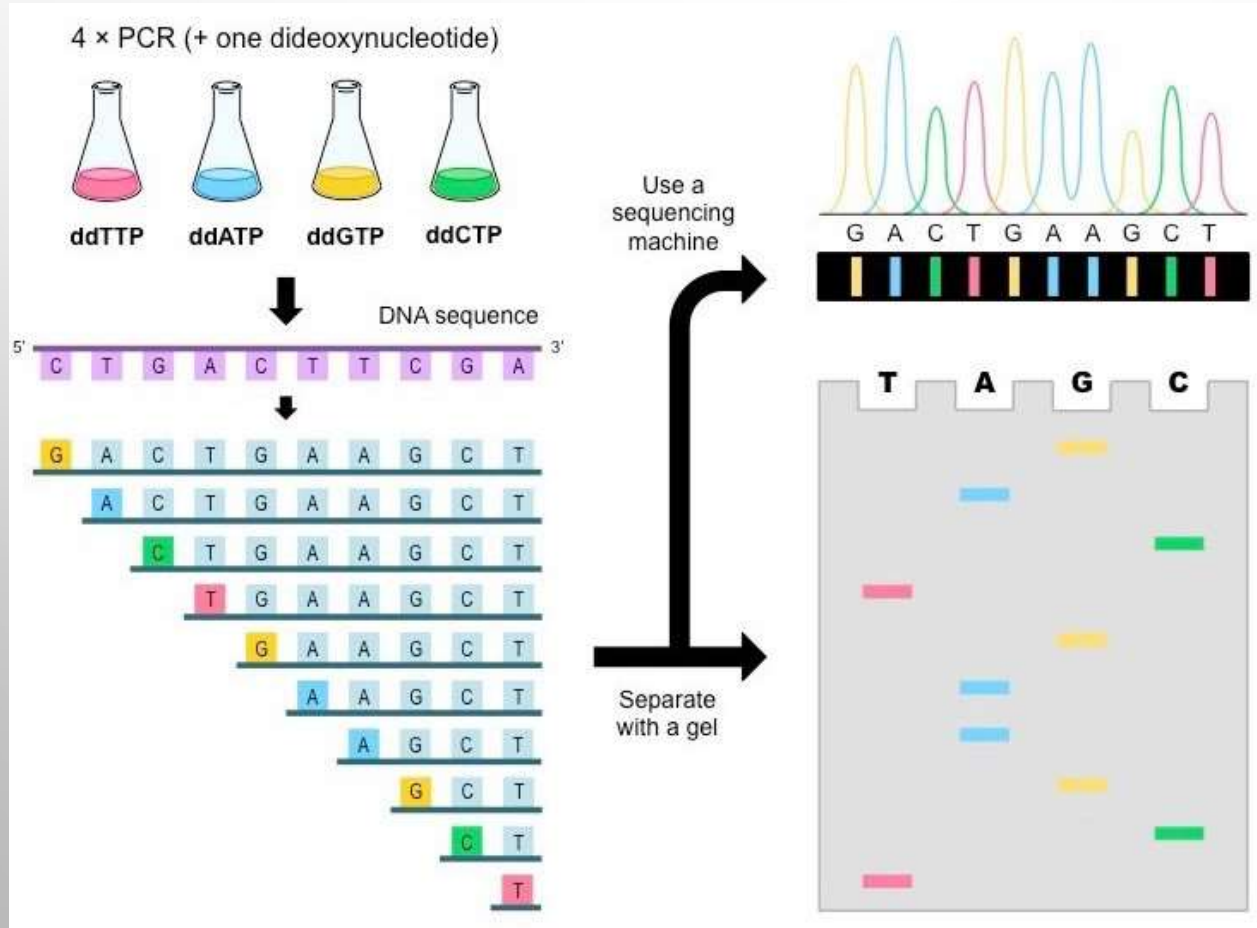
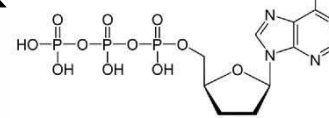
- 藉由螢光標定ddNTP(雙脫氧核苷酸,五碳糖缺乏3端OH基)使PCR反應停止
- 定序長度 500-1200 bp

deoxynucleotide (dNTP)



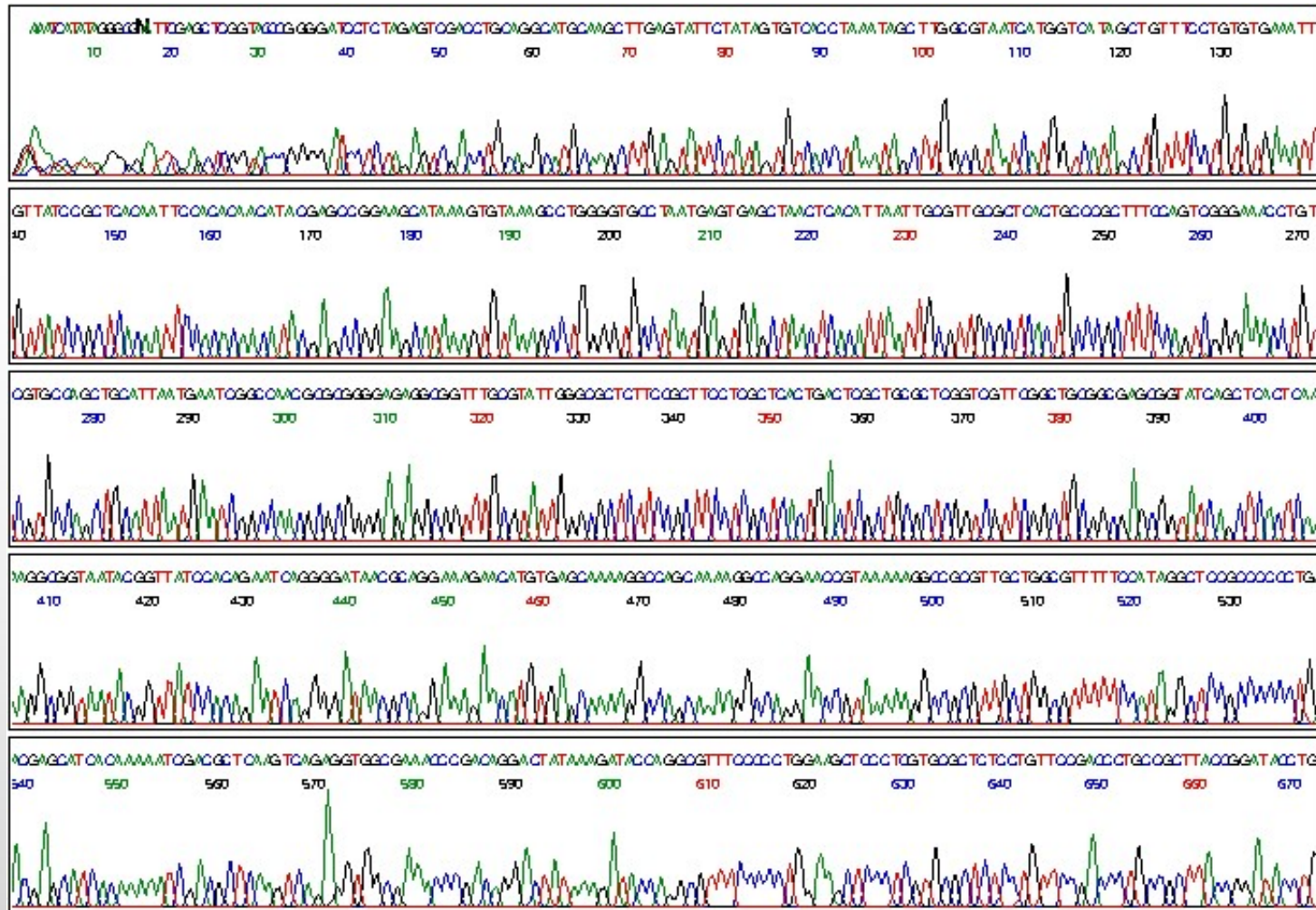
SANGER SEQ

dideoxynucleotide (ddNTP)



PCR

跑膠, 螢光分析



<https://www.youtube.com/watch?v=FvHRio1yyhQ>



<https://www.youtube.com/watch?v=iqAmkNSu3ol>



Next-generation sequencing (NGS)

First Generation

Sanger Sequencing
Maxam and Gilbert
Sanger Chain-termination

- Infer nucleotide identity using dNTPs then visualize with electrophoresis
- 500-1000 bp fragments
- Relatively slow and expensive

Second Generation

Next Generation Sequencing

454, Solexa, Ion Torrent, Illumina

- High throughput from the parallelization of sequencing reactions
- High accuracy
- ~50-500 bp fragments
- Faster and more affordable

Third Generation

PacBio, Oxford Nanopore

- Sequence native DNA in real time with single-molecule resolution
- Traditionally lower accuracy than NGS
- Tens of kb fragments, on average

Short-read sequencing

Long-read sequencing

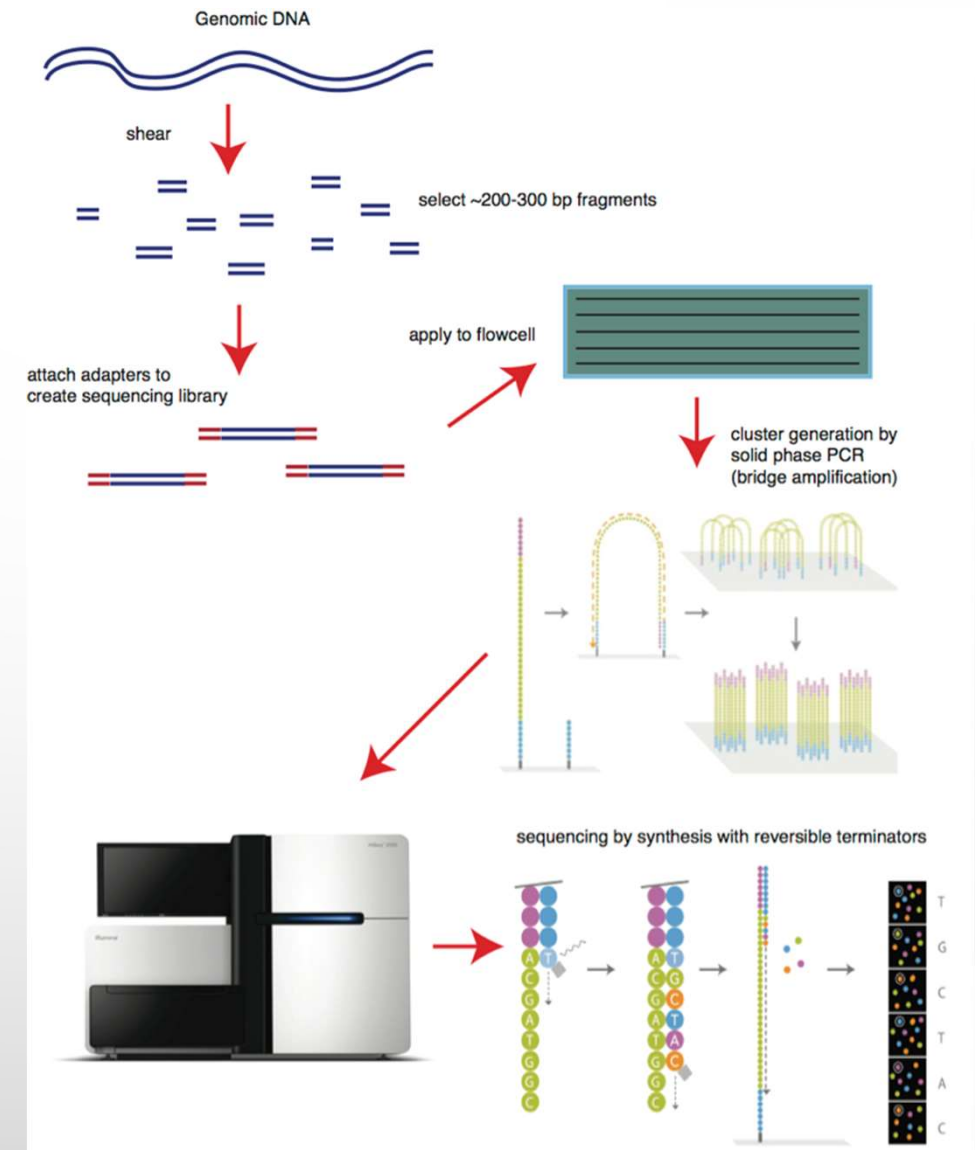
核心方法著重於直接定序單一分子

避免將 DNA 震碎成小片段；若針對 DNA 的表觀遺傳修飾，也能省略建立定序基因庫（library）的階段，故能夠避免 PCR error。



Steps to illumina sequencing

- **Library construction**
 - - Fragment, attach adapter DNA
- **Cluster generation**
 - - Add to flow cell
 - - Bridge amplification
- **Sequencing**
 - - Single base at a time, imaging
- **Data analysis**
 - - Images transformed into basecalls and 'reads'



<https://www.youtube.com/watch?v=womKfikWlxM>



<https://www.youtube.com/watch?v=fCd6B5HRaZ8>

1. **Library preparation**—the sequencing library is prepared by random fragmentation of the DNA or cDNA sample, followed by 5' and 3' adapter ligation. Alternatively, “tagmentation” combines the fragmentation and ligation reactions into a single step that greatly increases the efficiency of the library preparation process.⁹ adapter-ligated fragments are then PCR amplified and gel purified.

A. Library Preparation

Genomic DNA 

↓ Fragmentation

Adapters 

↓ Ligation

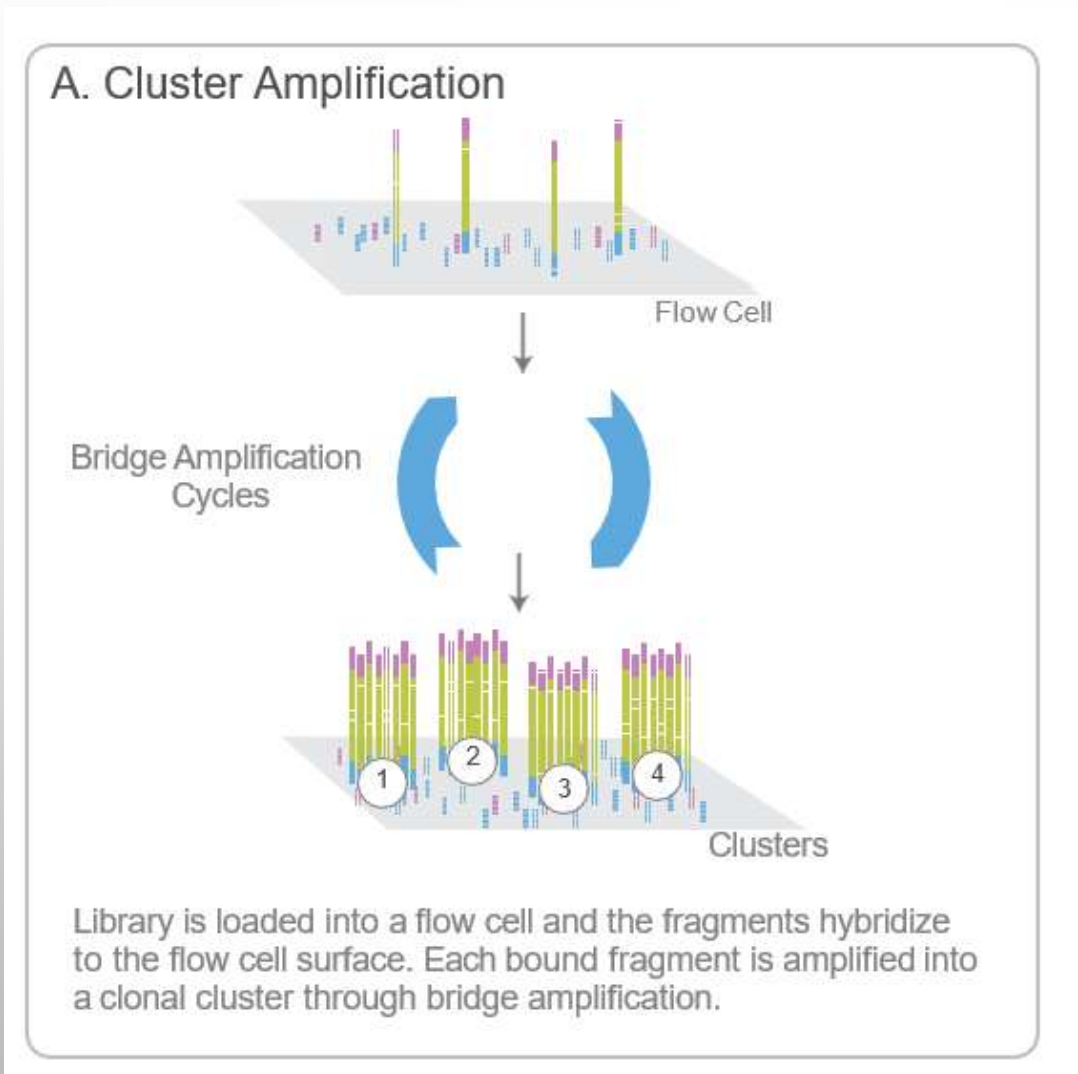
Sequencing Library 

NGS library is prepared by fragmenting a gDNA sample and ligating specialized adapters to both fragment ends.

1. 利用超聲波將DNA打斷成200-500 bp的片段大小，然後於片段兩端接上接頭(adapter)



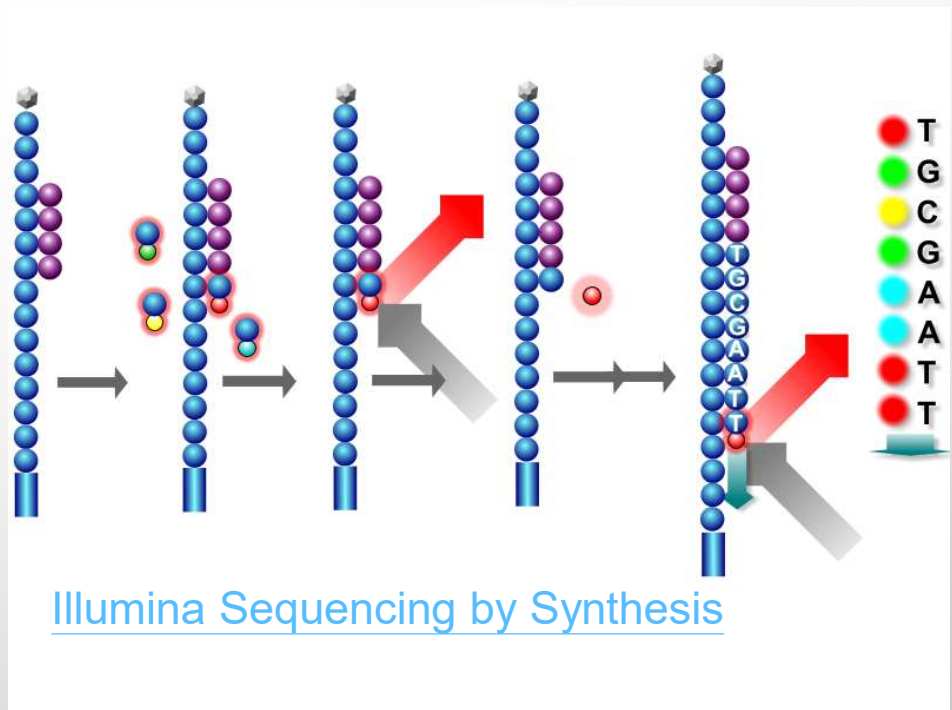
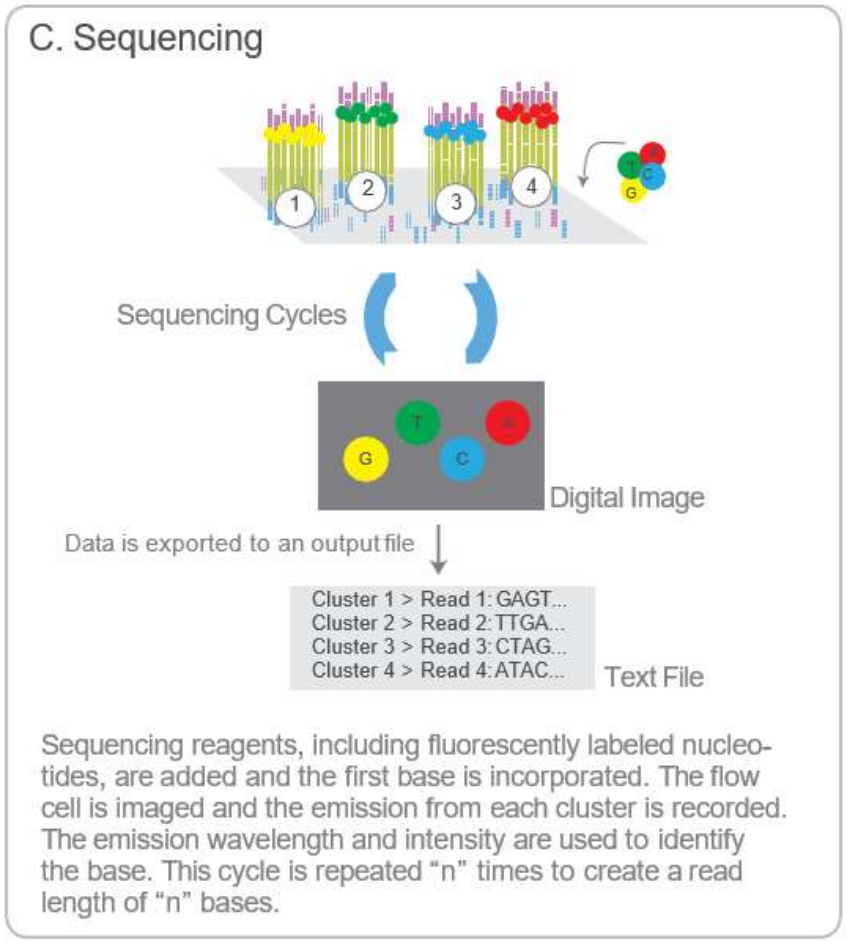
2. **Cluster generation**—for cluster generation, the library is loaded into a flow cell where fragments are captured on a lawn of surface-bound oligos complementary to the library adapters. Each fragment is then amplified into distinct, clonal clusters through bridge amplification. When cluster generation is complete, the templates are ready for sequencing.



2. 將已接接頭的DNA片段放入到表面帶有互補接頭序列的flowcell，接頭互相配對後讓DNA片段吸附於flowcell上

3. 透過橋式聚合酶連鎖反應進行DNA複製，放大訊號

3. **Sequencing**—illumina SBS technology utilizes a proprietary reversible terminator–based method that detects single bases as they are incorporated into DNA template strands. As all 4 reversible, terminator-bound dntps are present during each sequencing cycle, natural competition minimizes incorporation bias and greatly reduces raw error rates compared to other technologies. The result is highly accurate base-by-base sequencing that virtually eliminates sequence-context-specific errors, even within repetitive sequence regions and homopolymers.

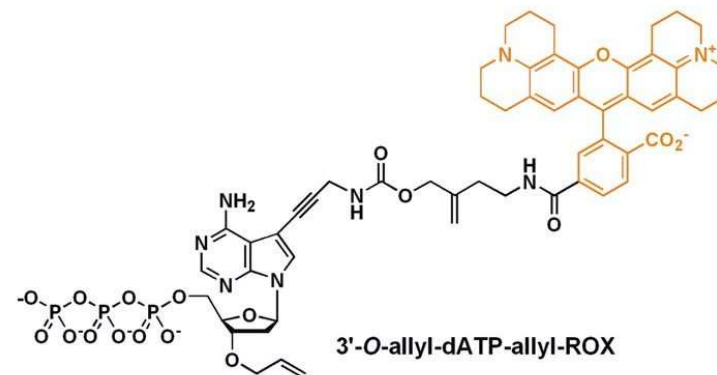
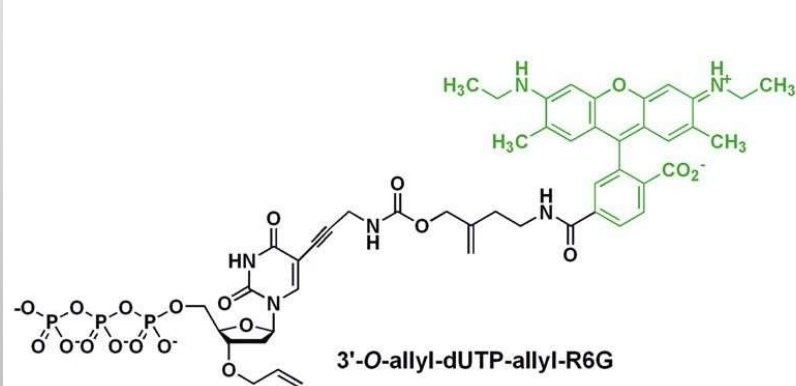
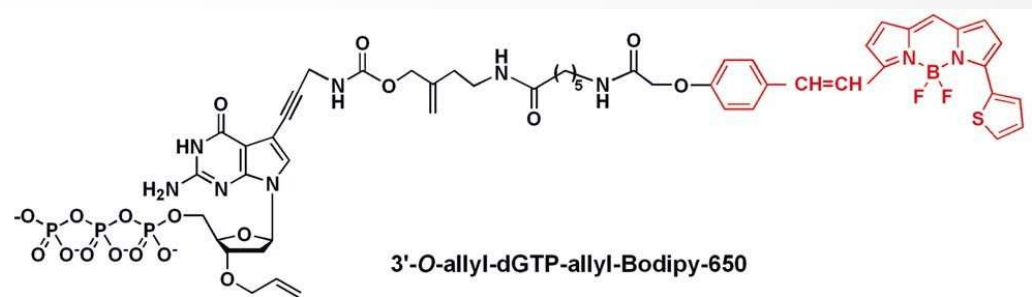
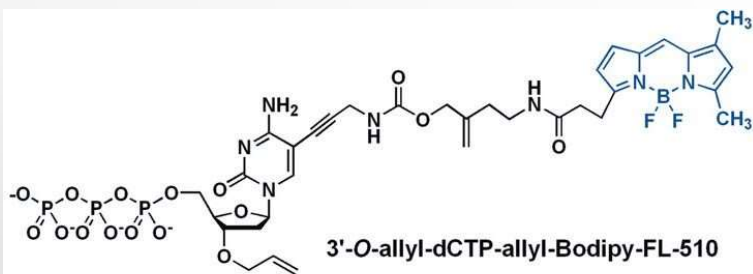


定序方法採用邊合成邊定序（如同 Sanger 定序法），加入不同已標定特定可移除螢光分子之鹼基(dNTP)及合成反應試劑，反覆讓螢光移除及偵測，達到高速且大量的DNA定序。



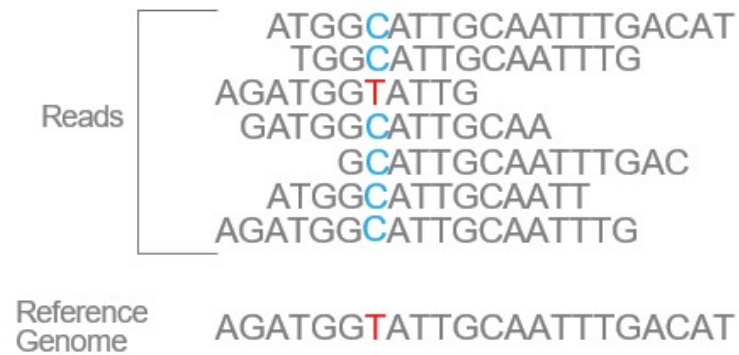
SBS — 反應用特殊核苷酸

- SBS所使用的核苷酸為特殊鹼基，上頭帶有發色基，可在激光後發出不同顏色
- 核苷酸五碳糖3端上帶有烯丙基(allyl group), 可使PCR反應無法進行
- 發色基及五碳糖3端的烯丙基可以鈹(pd)反應將其移除，使PCR反應繼續



4. **Data analysis**—during data analysis and alignment, the newly identified sequence reads are then aligned to a reference genome. Following alignment, many variations of analysis are possible such as single nucleotide polymorphism (SNP) or insertion-deletion (indel) identification, read counting for RNA methods, phylogenetic or metagenomic analysis, and more.

D. Alignment & Data Analysis

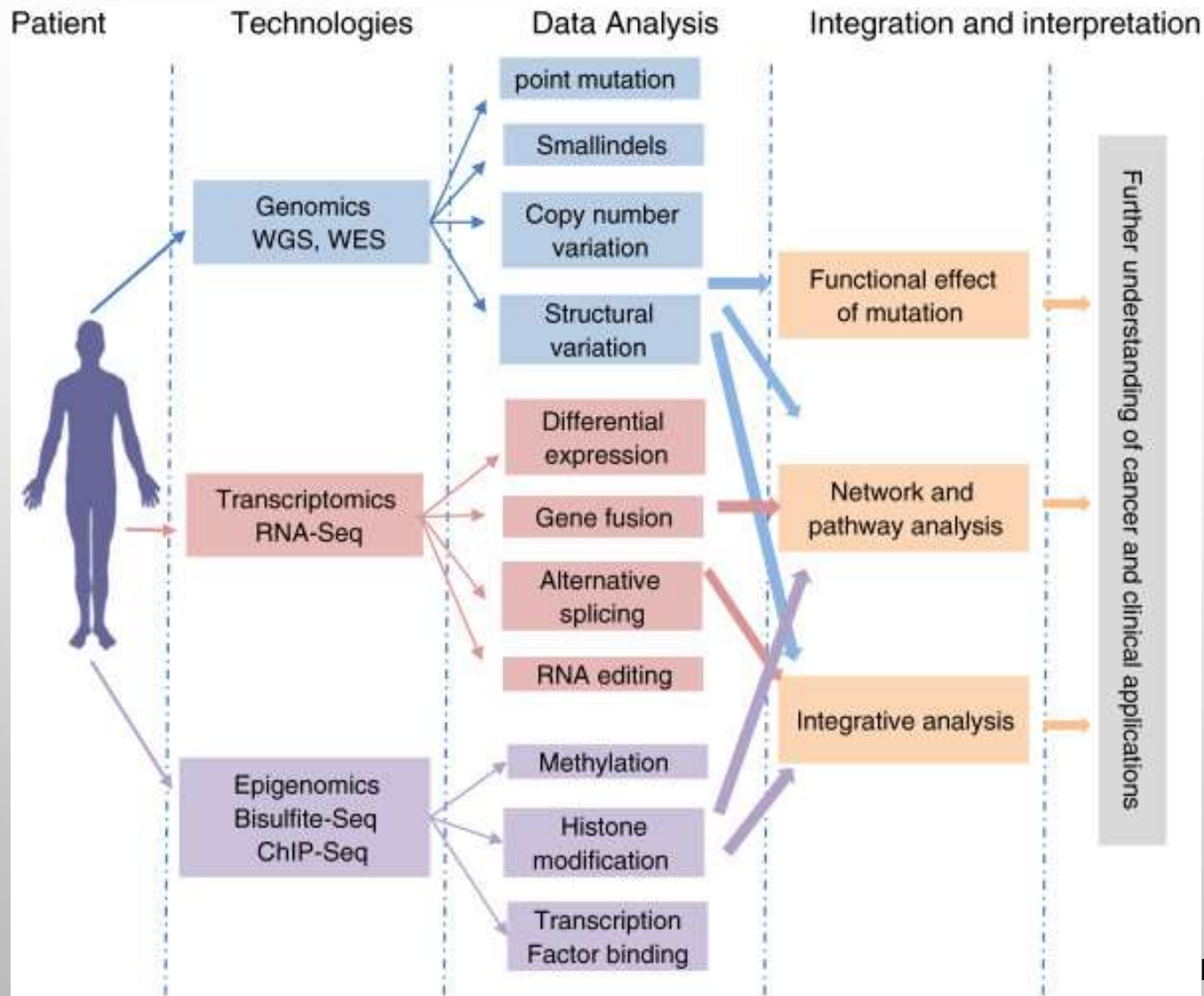


Reads are aligned to a reference sequence with bioinformatics software. After alignment, differences between the reference genome and the newly sequenced reads can be identified.



<https://www.youtube.com/watch?v=l4BAfRekohk>

NGS applications



- **DNA LEVEL:**
 - WHOLE GENOME SEQUENCING
 - GENOME DE NOVO
 - EXOME & TARGET REGION
 - CHIP SEQUENCING
- **RNA LEVEL:**
 - TRANSCRIPTOME
 - MIRNA EXPRESSION
- **SPECIAL PROJECT:**
 - EPIGENETICS (METHYLATION)
 - METAGENOMICS (菌相分析)

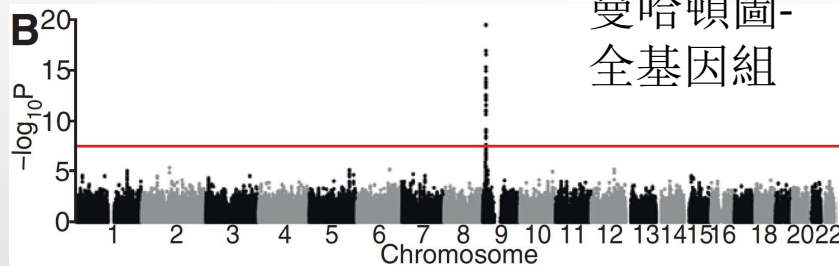
whole exome sequencing (WES)

Whole genome sequencing (WGS)

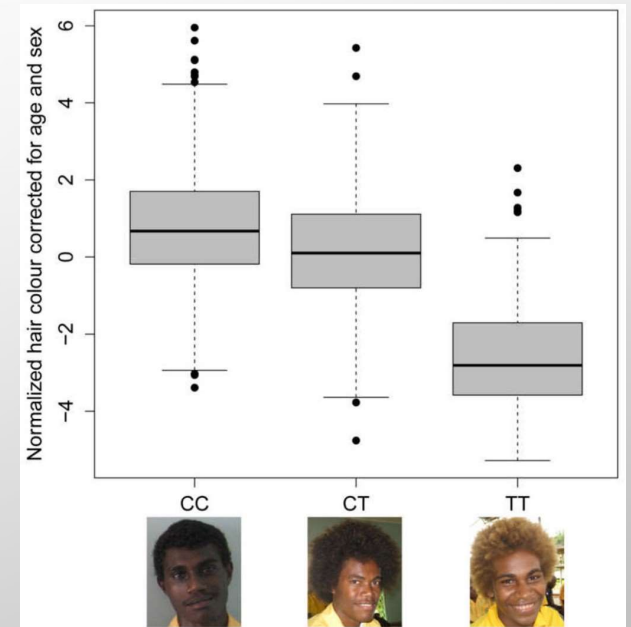
(1) Genome-wide association study, GWAS (全基因組關聯分析)

- 由全基因組中找出變異序列或基因，個體間的遺傳背景不宜差異太大
- 金髮個體：42位， 黑髮個體：43位
- 全基因組定序 → GWAS分析 → 候選基因 → 基因功能分析

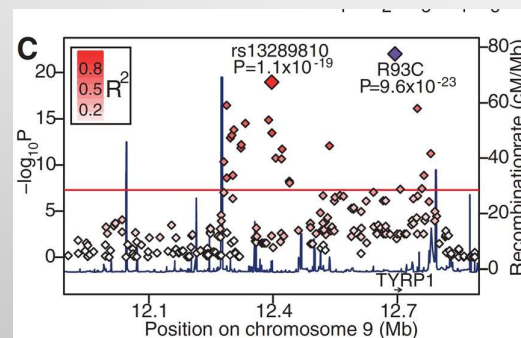
42位



R93C DNA突變



43位



19種癌症次世代基因定序納健保 最高補助3萬最快5月上路

(中央社記者陳婕翎2024/2/6 19:25)

包含**14**種實體瘤及**5**種血液腫瘤

實體瘤NGS檢測給付**14**種癌別，包含**非小細胞肺癌、三陰性乳癌、卵巢癌 / 輸卵管癌 / 原發性腹膜癌、大腸直腸癌、攝護腺癌、泌尿道上皮癌、肝內膽管癌、黑色素瘤、腸胃道間質瘤、甲狀腺癌、甲狀腺髓質癌、胰臟癌、NTRK基因融合實體腫瘤、胃癌。**

石崇良表示，實體瘤NGS檢測規劃出**3**種模式：

第1種BRCA-1與BRCA-2基因，基因片段較長，採獨立給付，預計給付健保點值**1萬點**；

第2種是各癌別最基本基因位點小於**100**個基因的小套組，給付**2萬點**；

第3種100個以上基因位點大套組，給付**3萬點**。

精準狙擊癌細胞關鍵在基因檢測!

專家：有助於以下癌症患者治療

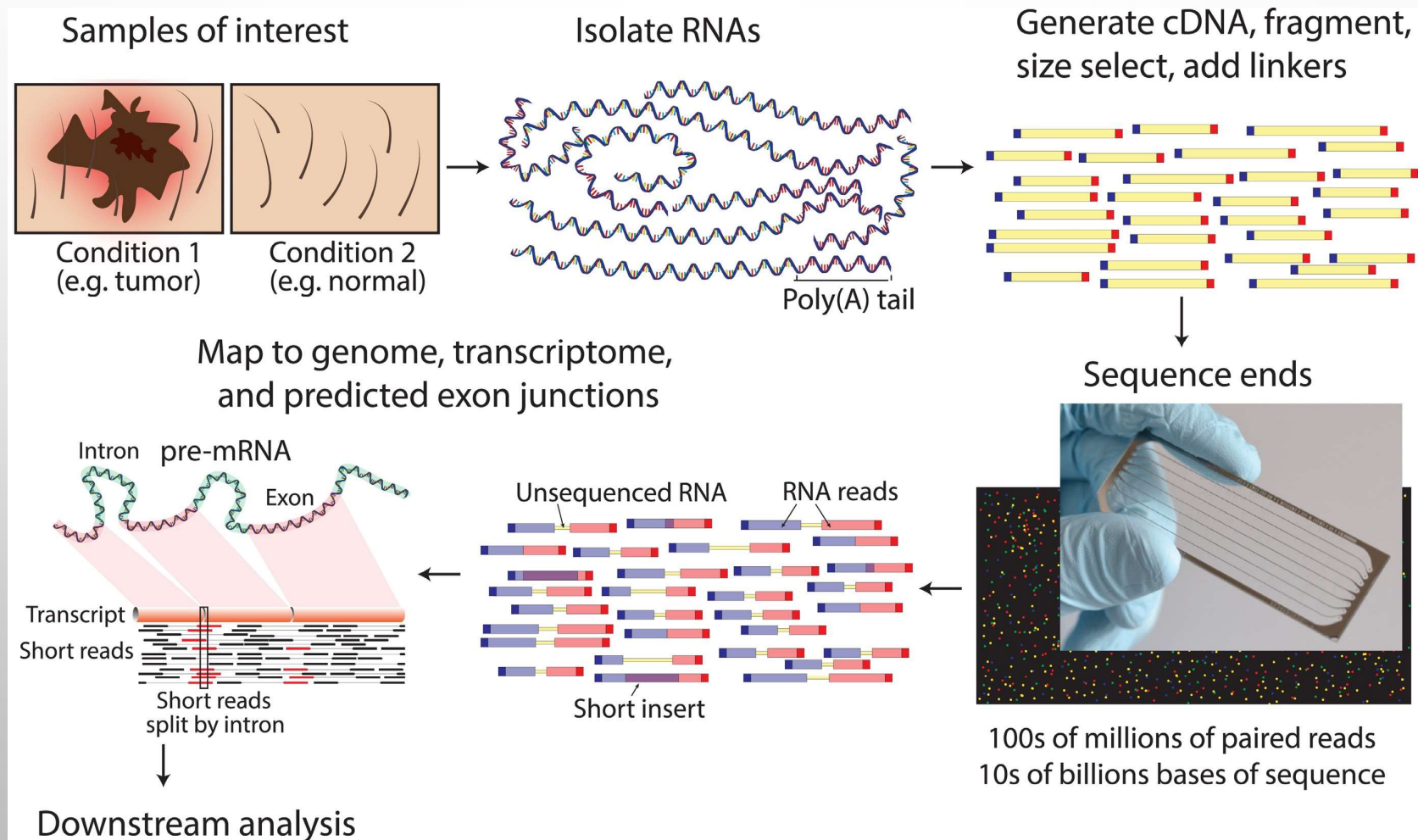
癌別	說明
肺癌	用藥空間最大，7成肺腺癌患者在接受基因檢測後可找到相對應抗癌藥物。
乳癌	約有2到3個特定變異基因，已有相對應的藥物問世，治療效果顯著。
卵巢癌	與乳癌類似，約有2到3個特定變異基因，已有相對應的藥物問世。
大腸癌	相對應新藥約2到3種，建議進行基因檢測，獲得最適合的藥物診治。

資料來源：台北醫學大學 李國遠副校長/台北慈濟醫院醫療部醫療長 何景良教授

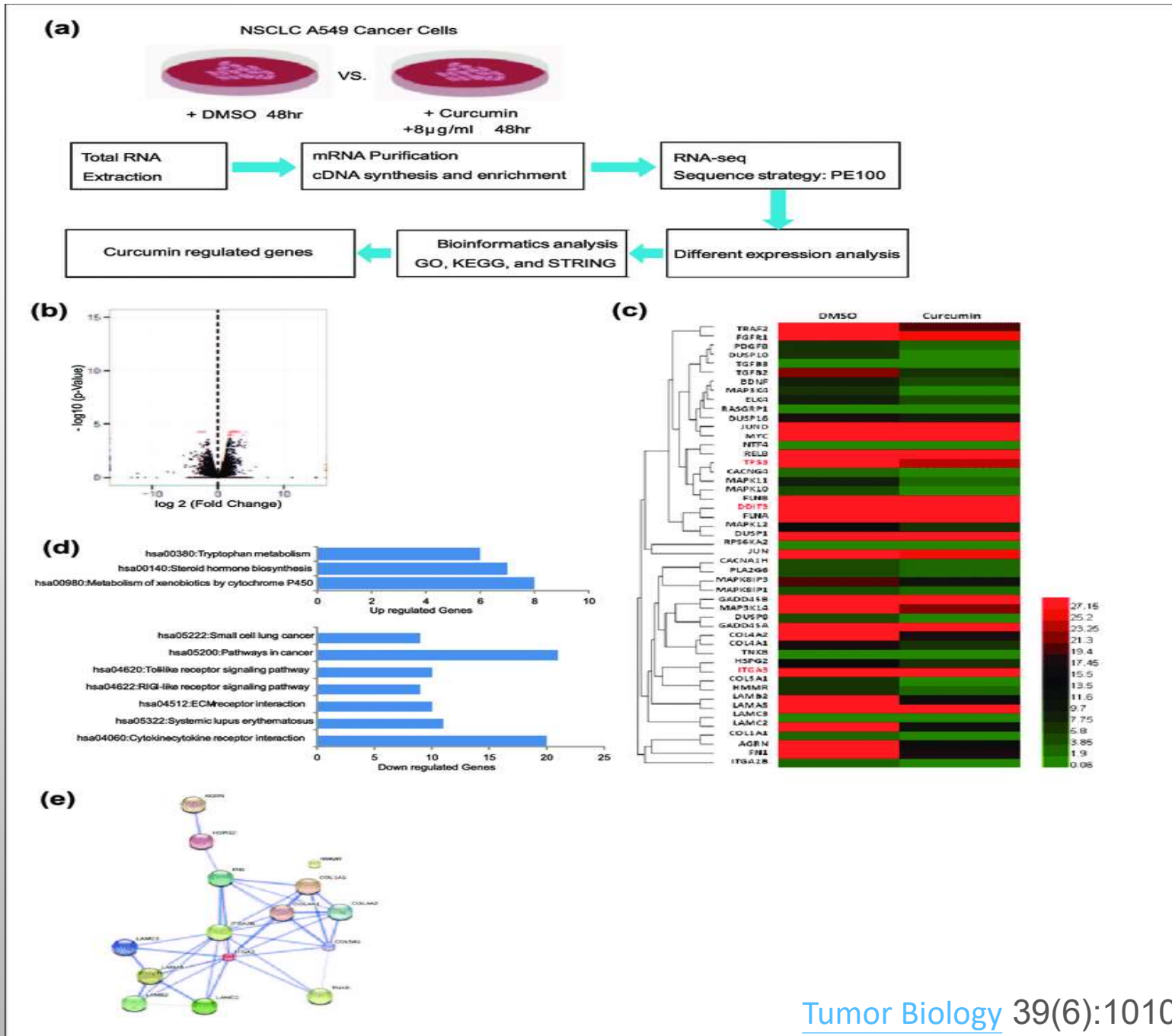
潮健康

(2) 轉錄體定序(RNA-seq)

抽取RNA → 轉成雙股DNA並切成小片段 → 霰彈式定序 → 對應到基因組相對位置

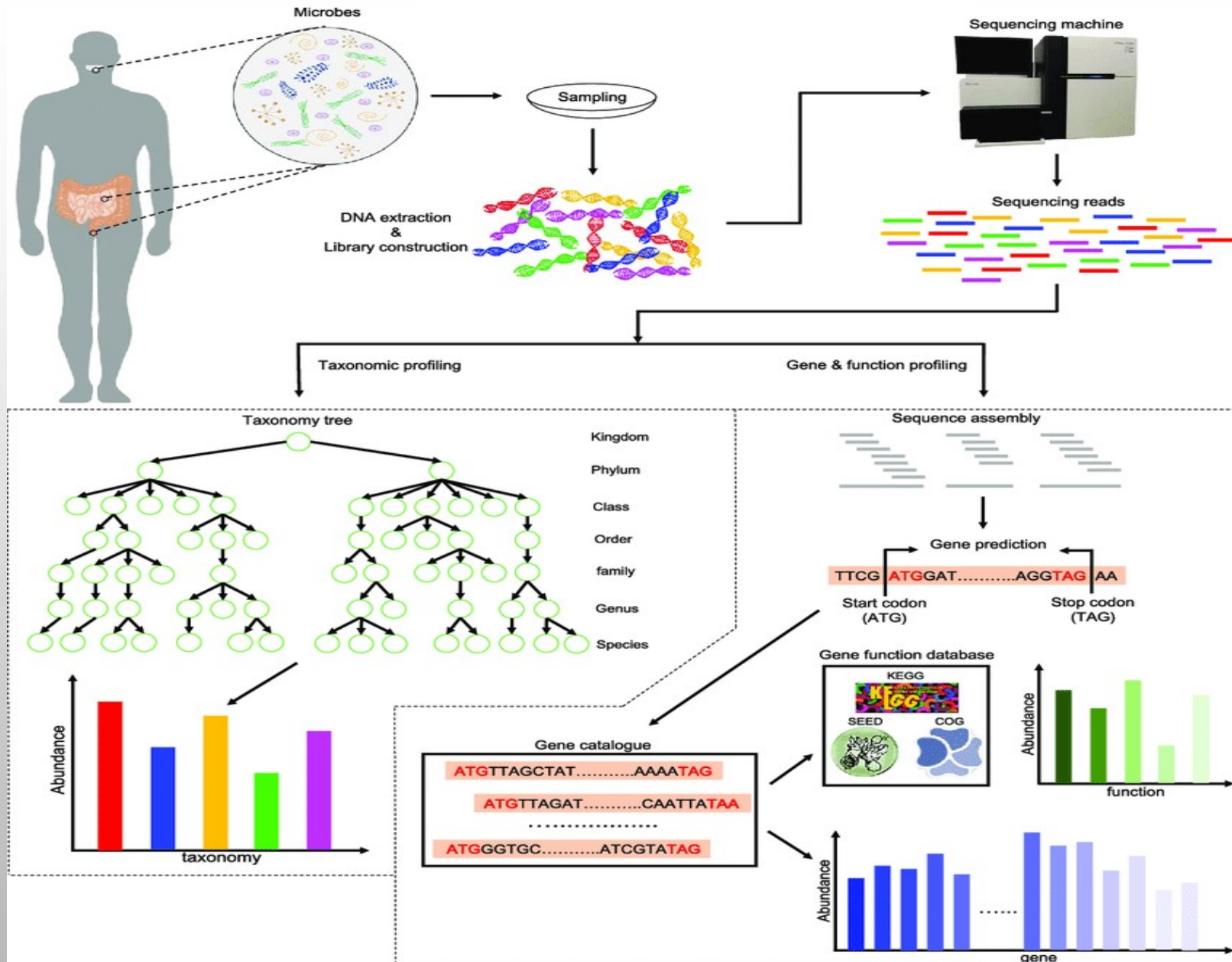


Transcriptome alteration analysis with RNA-seq in A549 NSCLC



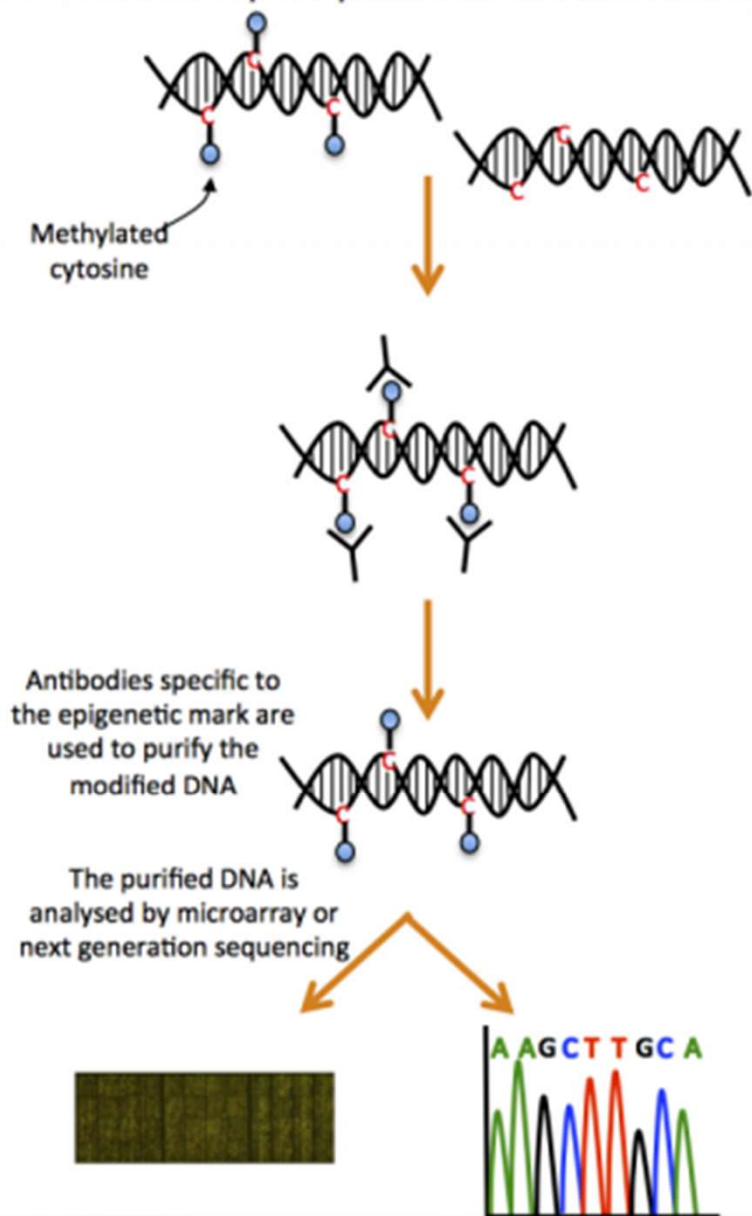
(3) 微生物検定(DNA-seq)

Metagenomic sequencing (Bacterial DNA sequence)

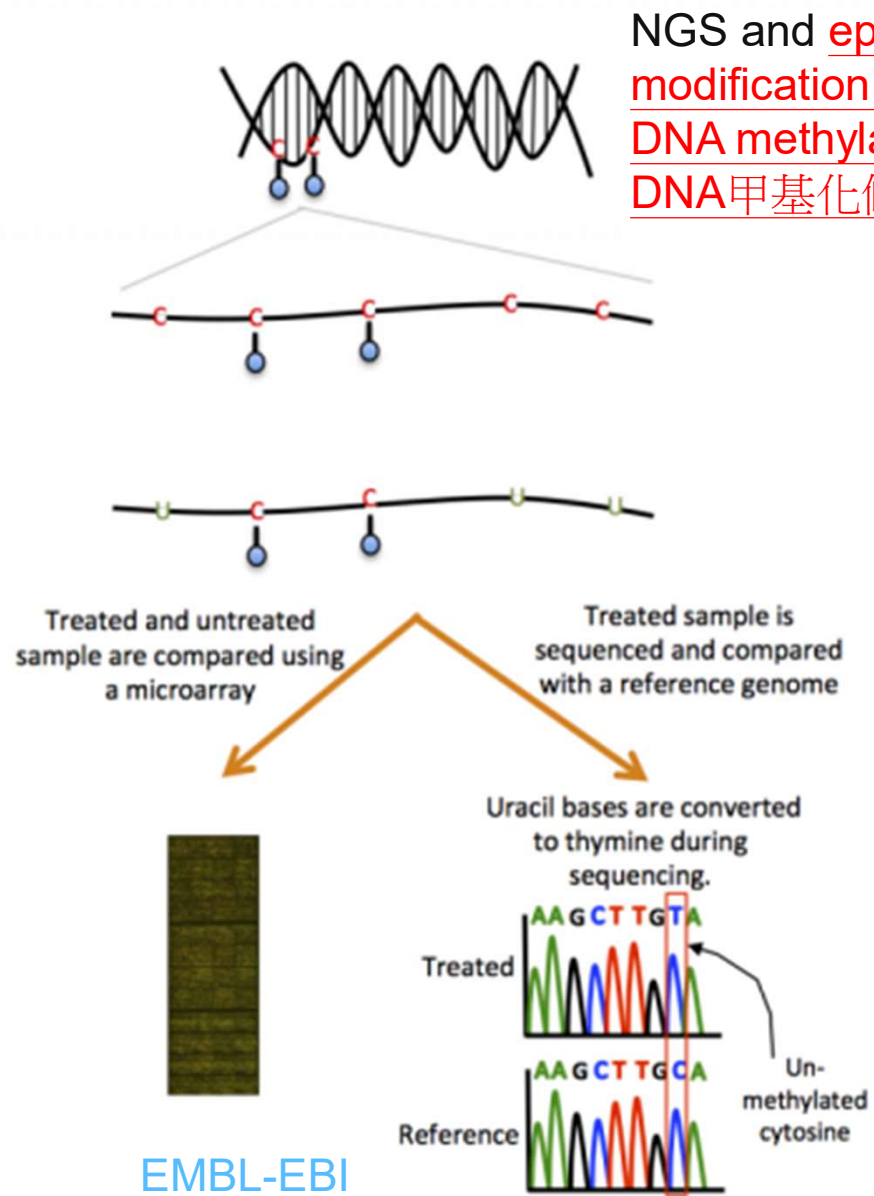


(4) 後遺傳基因修飾(DNA-seq)

Immunoprecipitation based methods

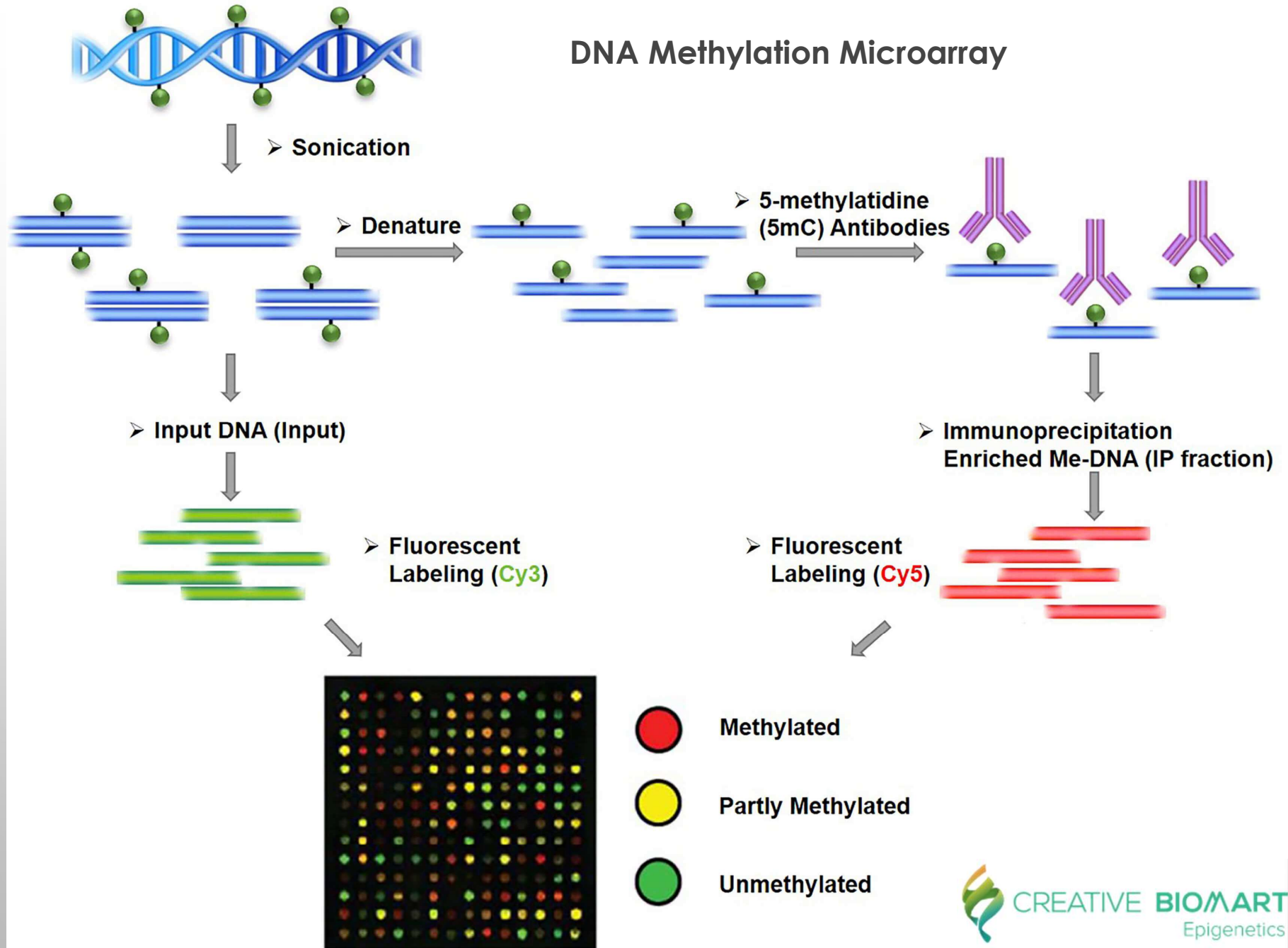


Bisulfite based methods



NGS and epigenetic modification (ex. DNA methylation-DNA甲基化修飾)

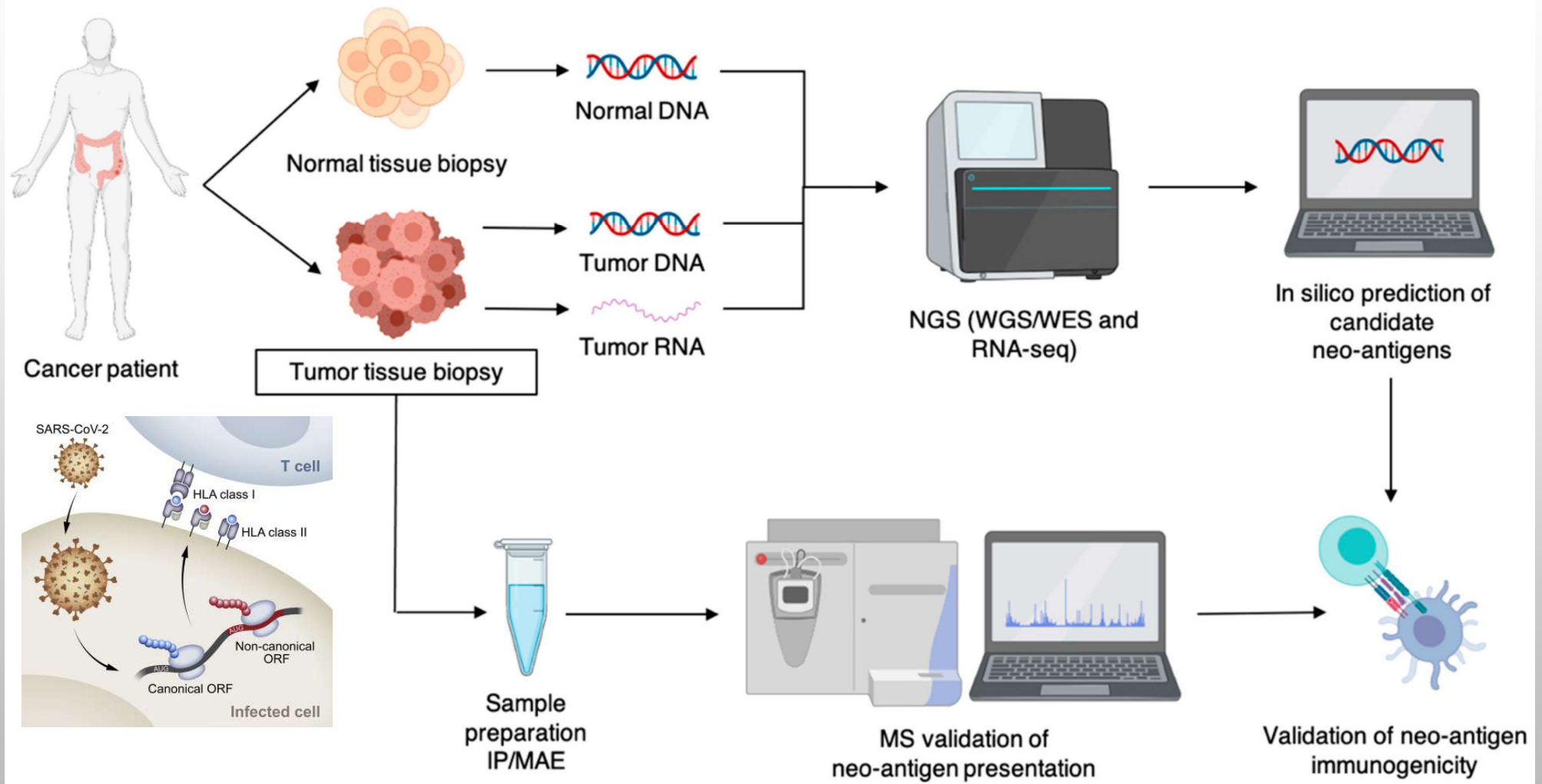
DNA Methylation Microarray



(5) 疫苗抗原篩選

Neo-Antigen mRNA Vaccines

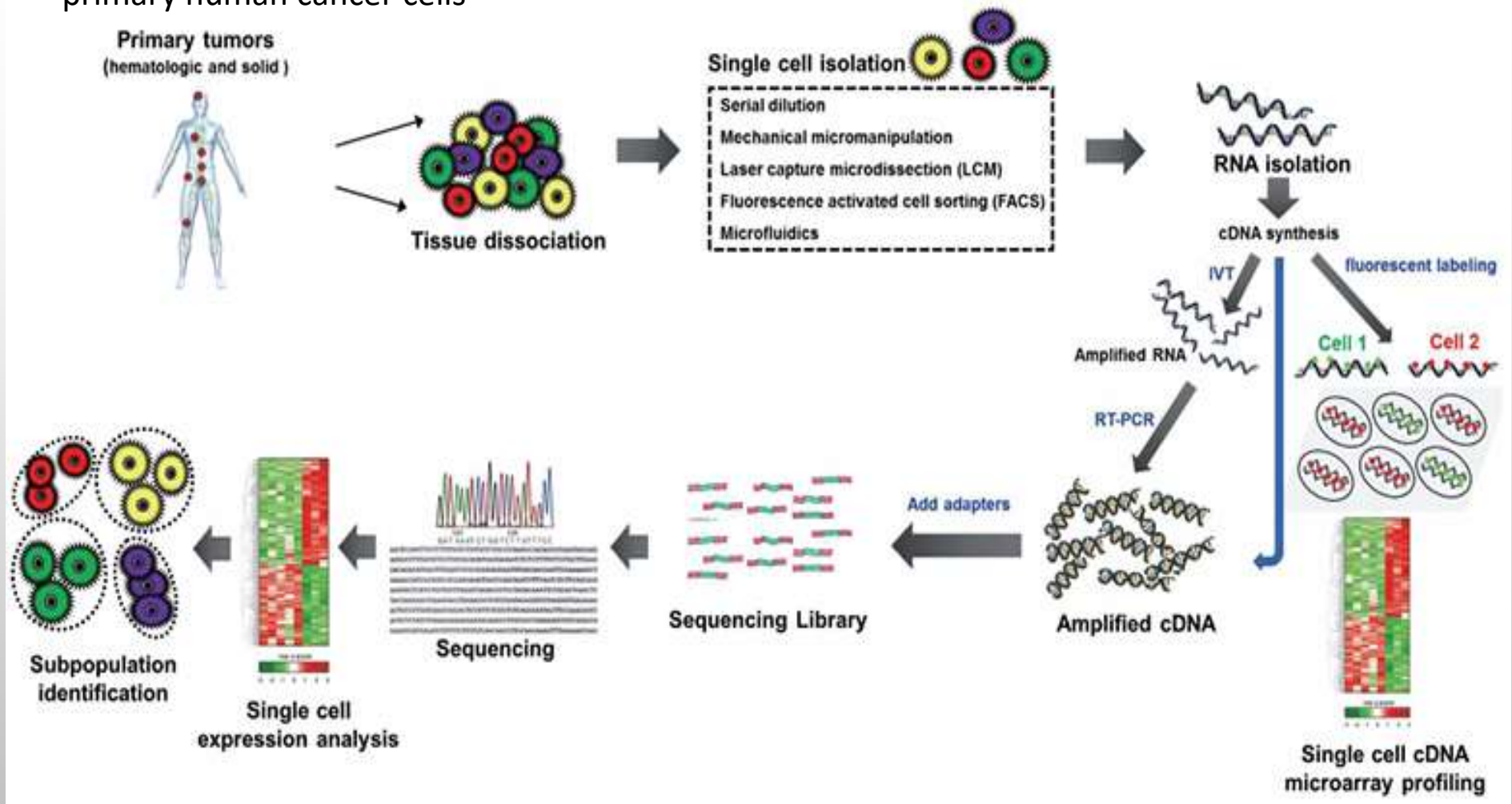
assessment of both **human and pathogen genomes**, their transcriptomes, as well as examination of **host immune responses**, such as **T and B cell diversity**. NGS can be used to assess the quality of vaccine stocks, **the diversity of HLA polymorphisms** in large populations, and also for detection of **new pathogenic strains** in mixed samples.



Vaccines 2020, 8(4), 776

(6)單細胞定序 (Single cell sequencing)

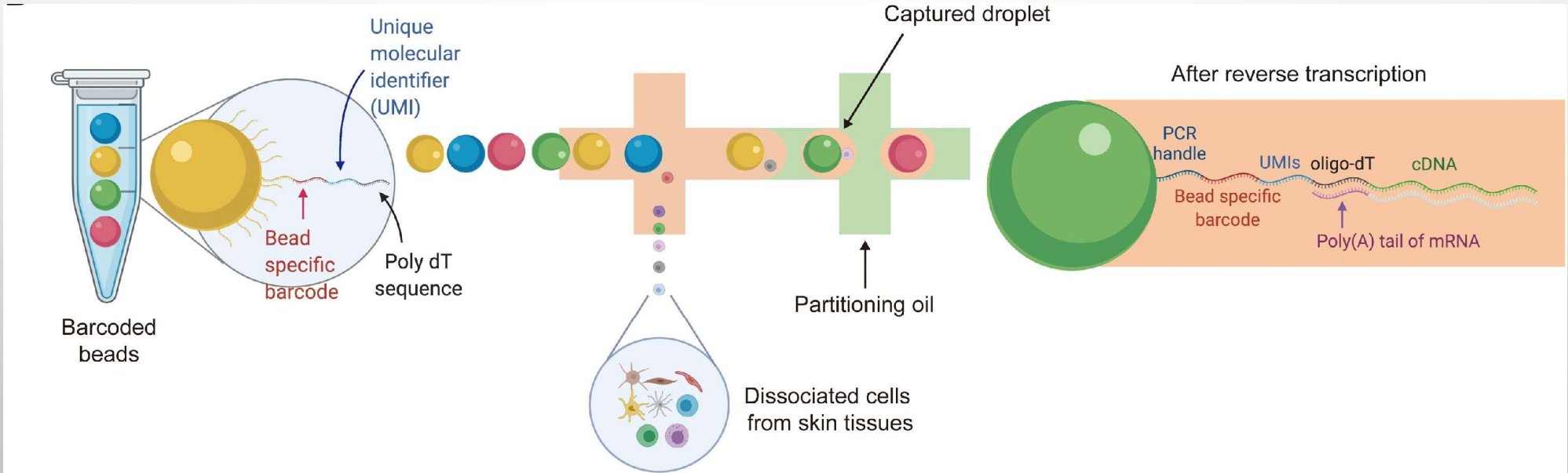
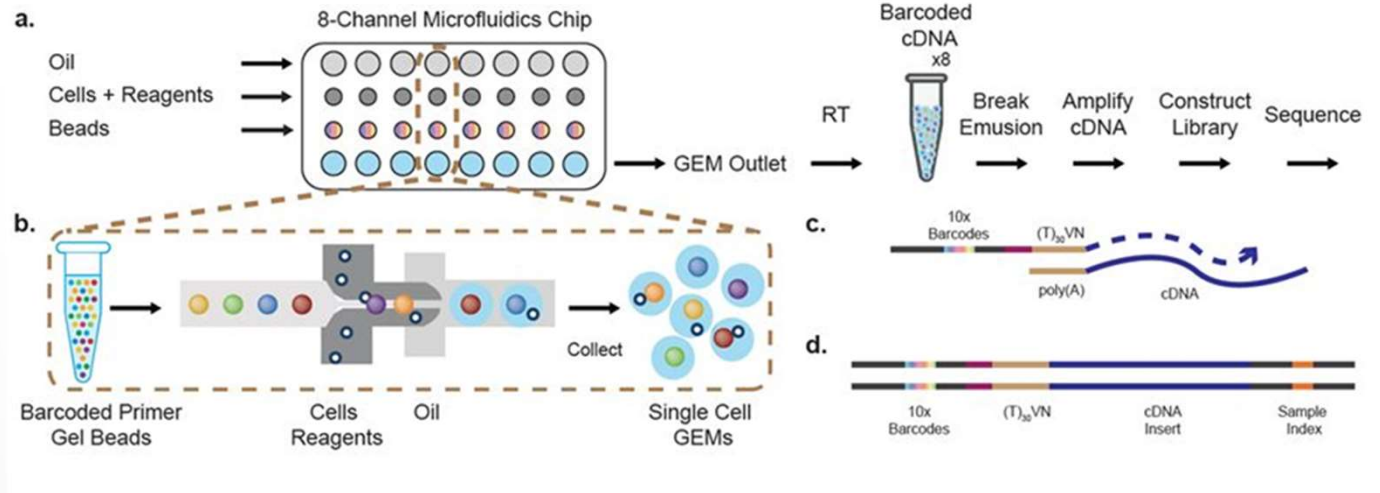
primary human cancer cells



Journal of Liver Cancer 2018;18(1):1-8.

單細胞解析系統原理

Single cell RNA sequencing experiments routinely target the retention of 500 to 10,000 cells which require an input of 800 to 16,000 cells.

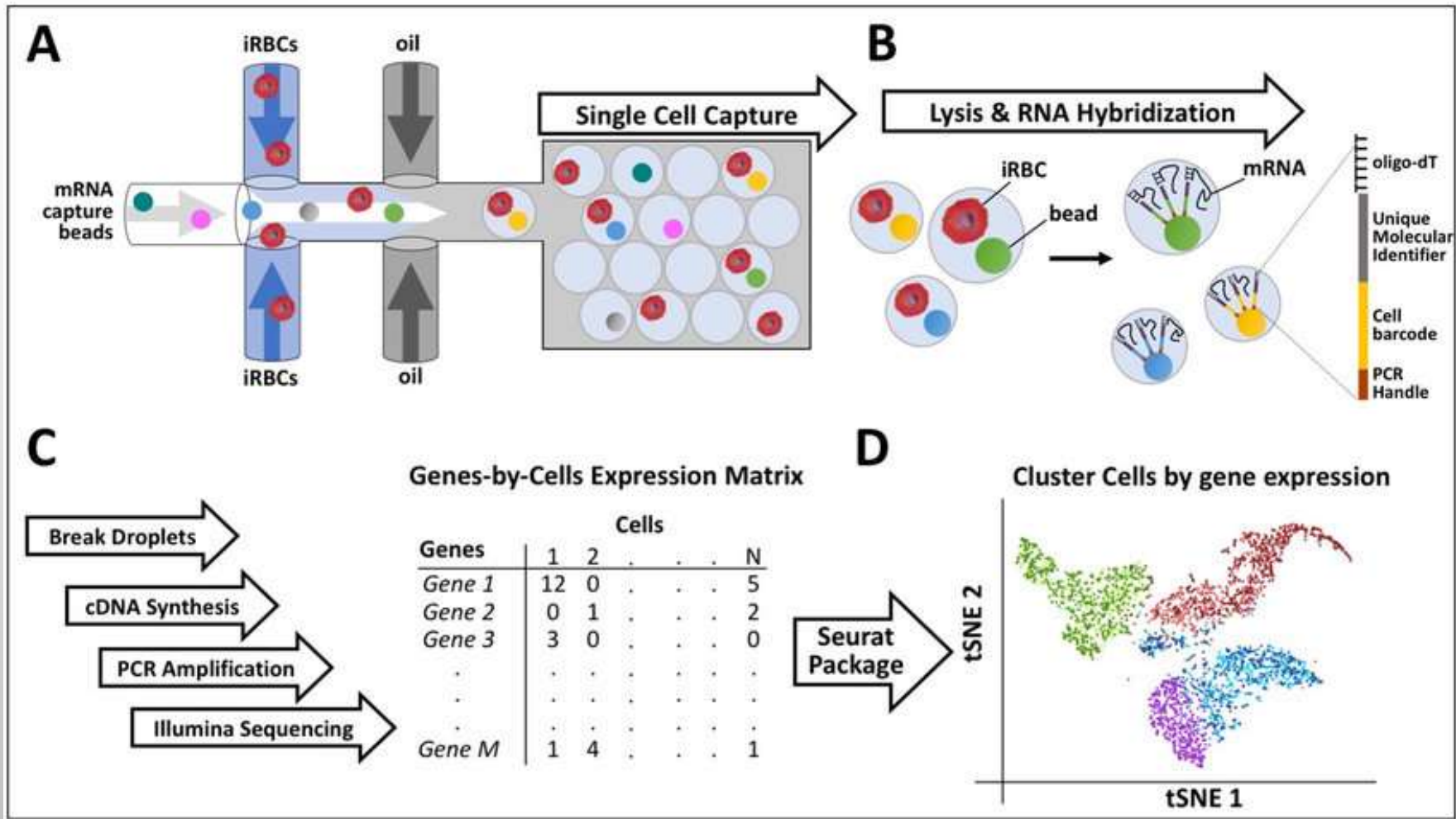


 <https://youtu.be/Rqz1QrmWcSM>

 <https://youtu.be/Q5pkJITEgxo>

 <https://www.youtube.com/watch?v=dbE1UlpxzHQ>

<https://doi.org/10.1016/j.jdermsci.2020.06.002>



•November 2017 [Nature](#) 551(7678)

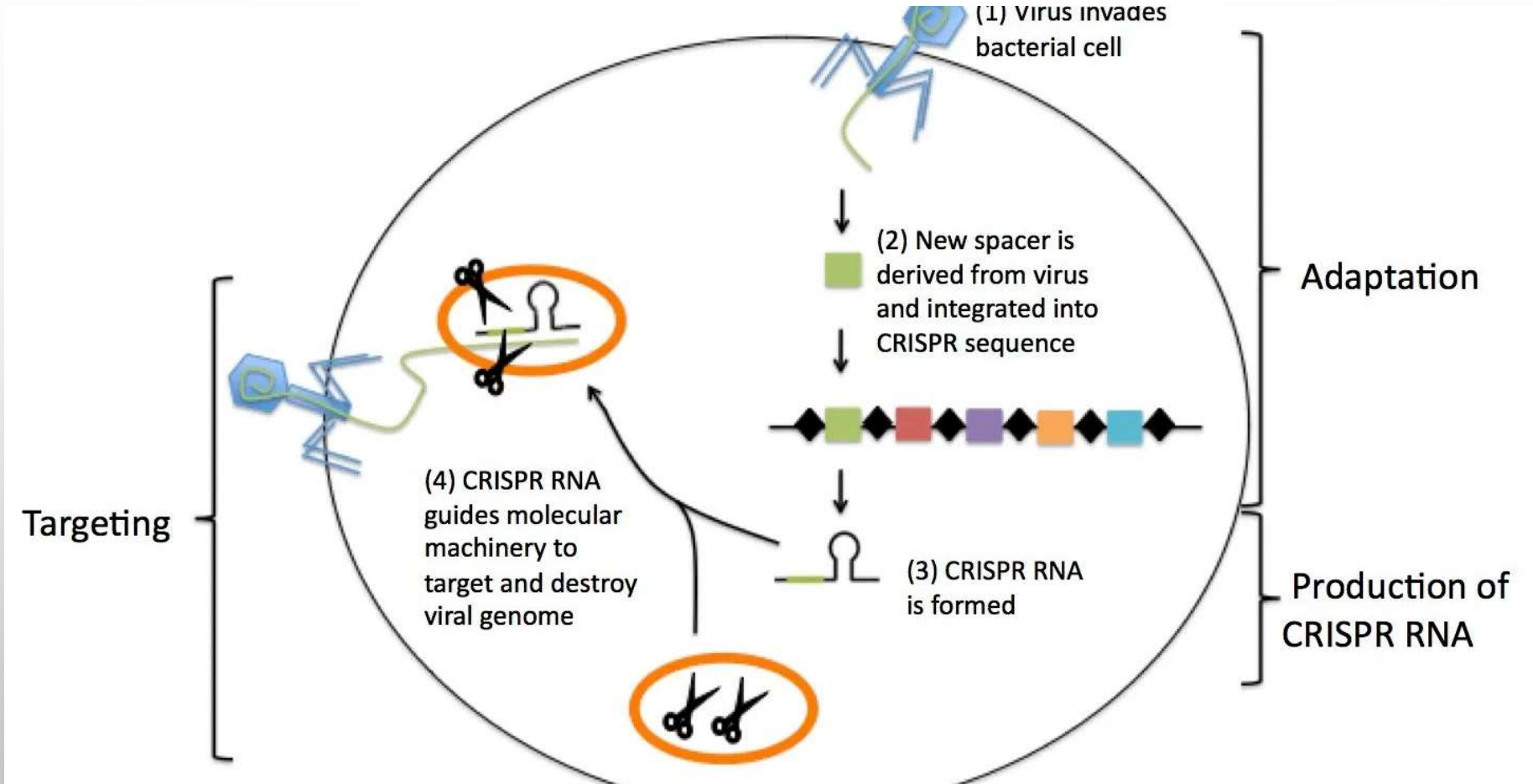


CRISPR/cas9 genome editing (基因編輯技術)

CRISPR-CAS9(Clustered regularly interspaced short palindromic repeat / CRISPR associated protein 9)

- The Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) and CRISPR- associated (Cas) system is the latest addition to the genome editing toolbox, offering a simple, rapid, and efficient solution.
- Derived from components of a simple bacterial immune system, the CRISPR-Cas9 system permits targeted gene cleavage and gene editing in a variety of eukaryotic cells, and because the endonuclease cleavage specificity in CRISPR-Cas9 system is guided by RNA sequences, editing can be directed to virtually any genomic locus by engineering the guide RNA sequence and delivering it along with the Cas endonuclease to your target cell.

Bacterial immune system



細菌的Cas9 酵素

Nobel Prize in Chemistry (2020)

”

2020 NOBEL PRIZE
IN CHEMISTRY GOES
TO EMMANUELLE
CHARPENTIER AND
JENNIFER DOUDNA



for discovering the
CRISPR/Cas9
technology

 eurofins | Genomics



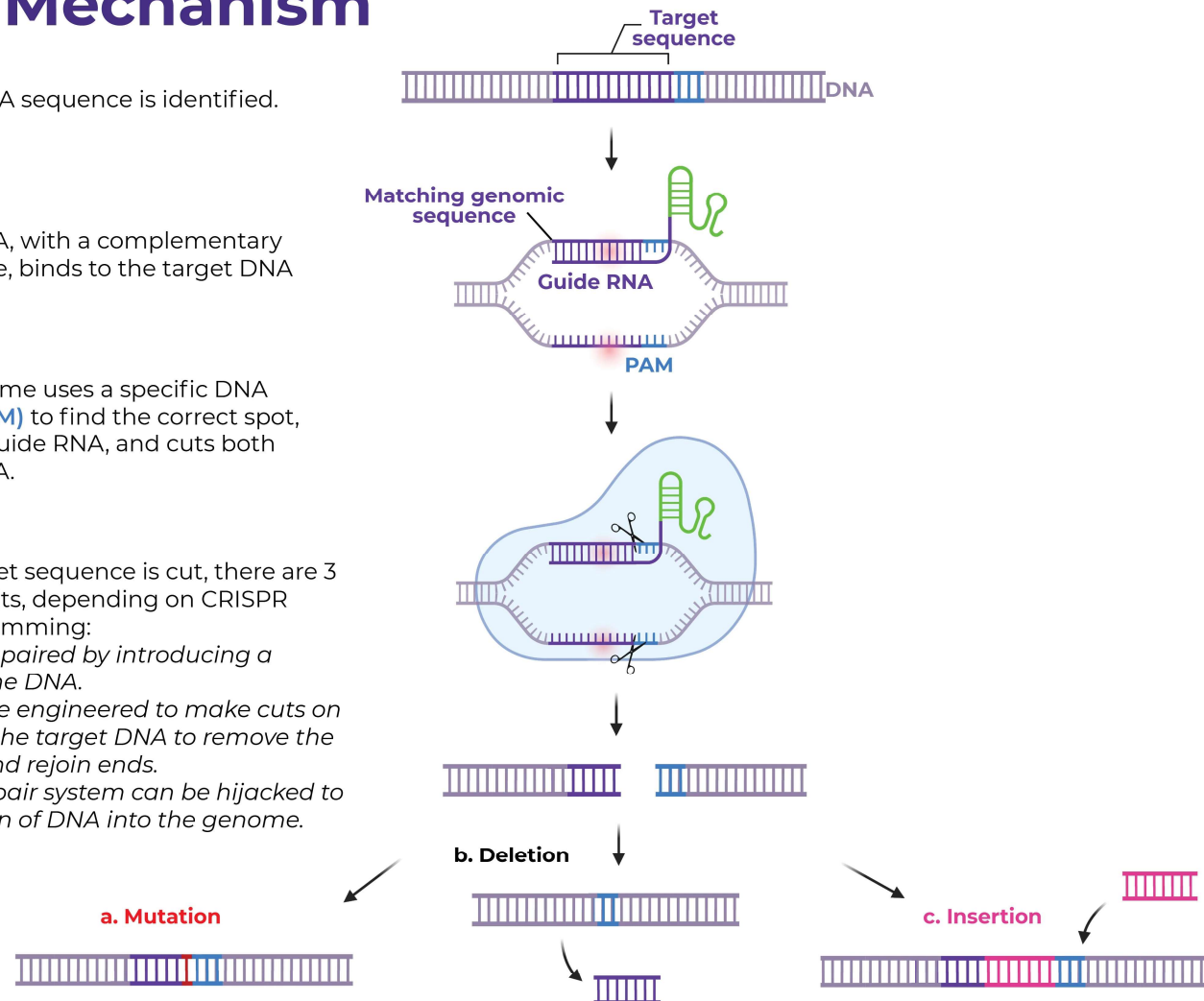
© Nobel Media. Ill. Niklas
Elmehed.
Emmanuelle
Charpentier
Prize share: 1/2

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Elmehed.
Jennifer A. Doudna
Prize share: 1/2

<https://mcbblog.nsfbio.com>

CRISPR Mechanism

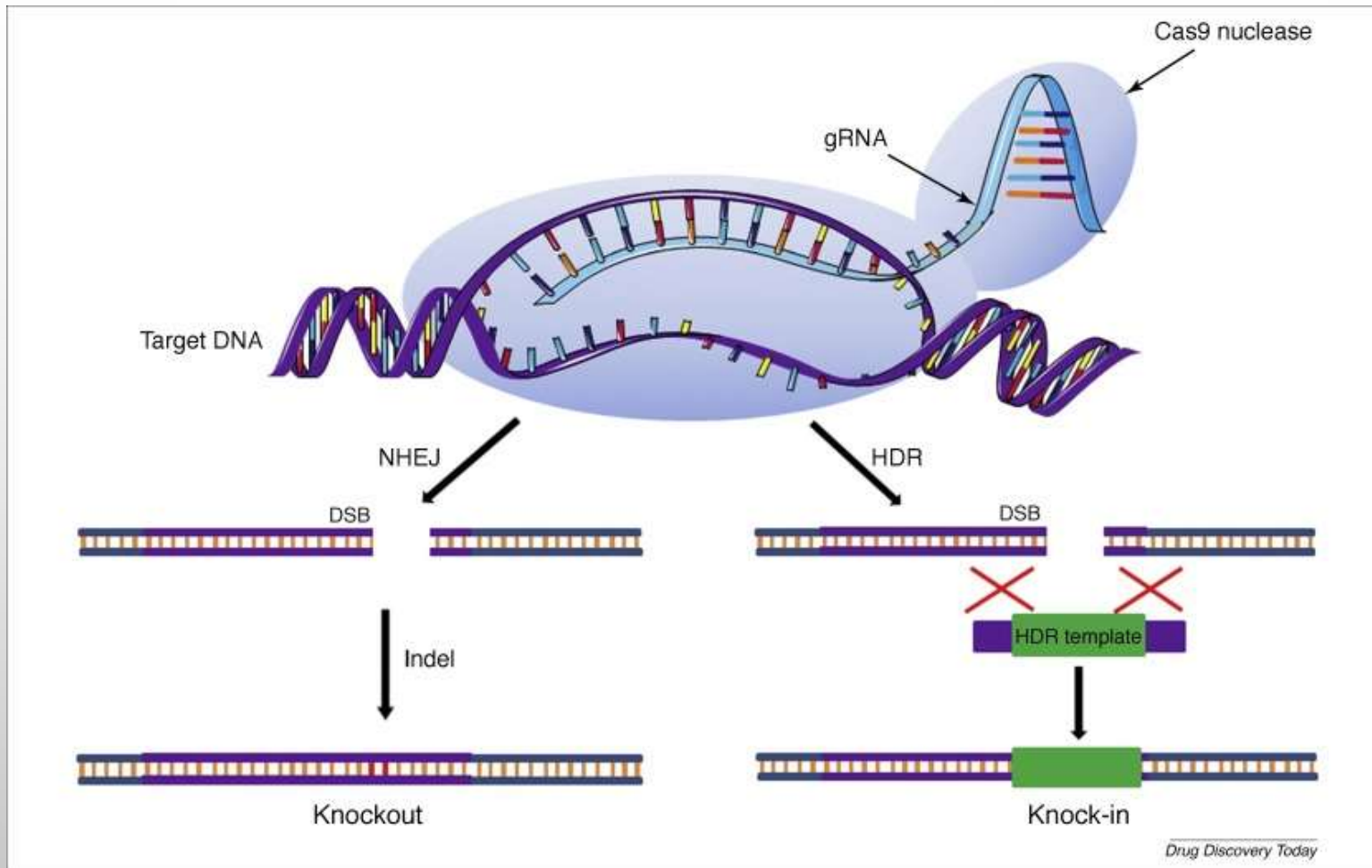
- ① The target DNA sequence is identified.
- ② The guide RNA, with a complementary DNA sequence, binds to the target DNA sequence.
- ③ The Cas9 enzyme uses a specific DNA sequence (**PAM**) to find the correct spot, binds to the guide RNA, and cuts both strands of DNA.
- ④ Once the target sequence is cut, there are 3 potential results, depending on CRISPR system programming:
 - a. The cut is repaired by introducing a mutation in the DNA.
 - b. Enzymes are engineered to make cuts on either side of the target DNA to remove the target DNA and rejoin ends.
 - c. The DNA repair system can be hijacked to insert a section of DNA into the genome.



<https://www.youtube.com/watch?v=UKbrwPL3wXE>

<https://www.umassmed.edu/rti/biology/crispr-cas9/>

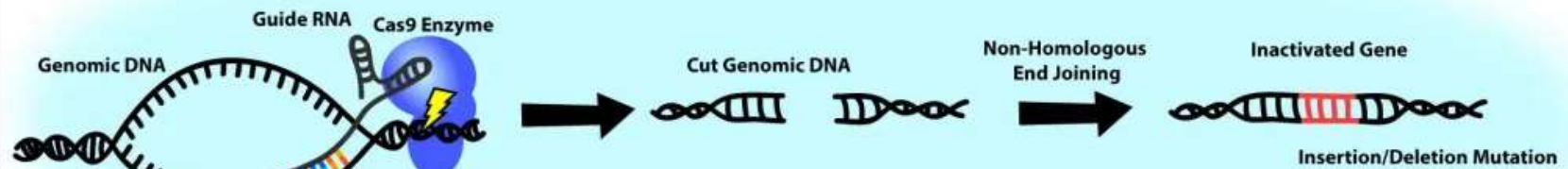
Functions of CRISPR-cas9



<https://www.youtube.com/watch?v=g7bkE1krgFM>

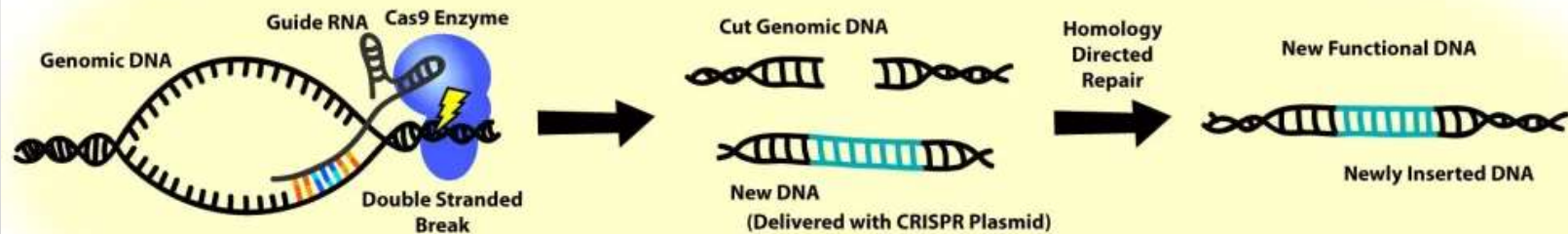


Gene Silencing with CRISPR



After the Cas9 enzyme is guided to the right location in the genome and cuts the DNA, the cell's natural repair mechanisms take over. The cut is fixed by non-homologous end joining. This process is error-prone and does not perfectly replace the cut DNA, often resulting in an insertion or deletion mutation which silences the gene.

Gene Insertion with CRISPR



To insert a gene, the new gene is added into the original CRISPR plasmid. It is designed to line up perfectly with the cut DNA strands, so the cell uses a different technique, homology directed repair, to incorporate a new stretch of DNA into the genome.

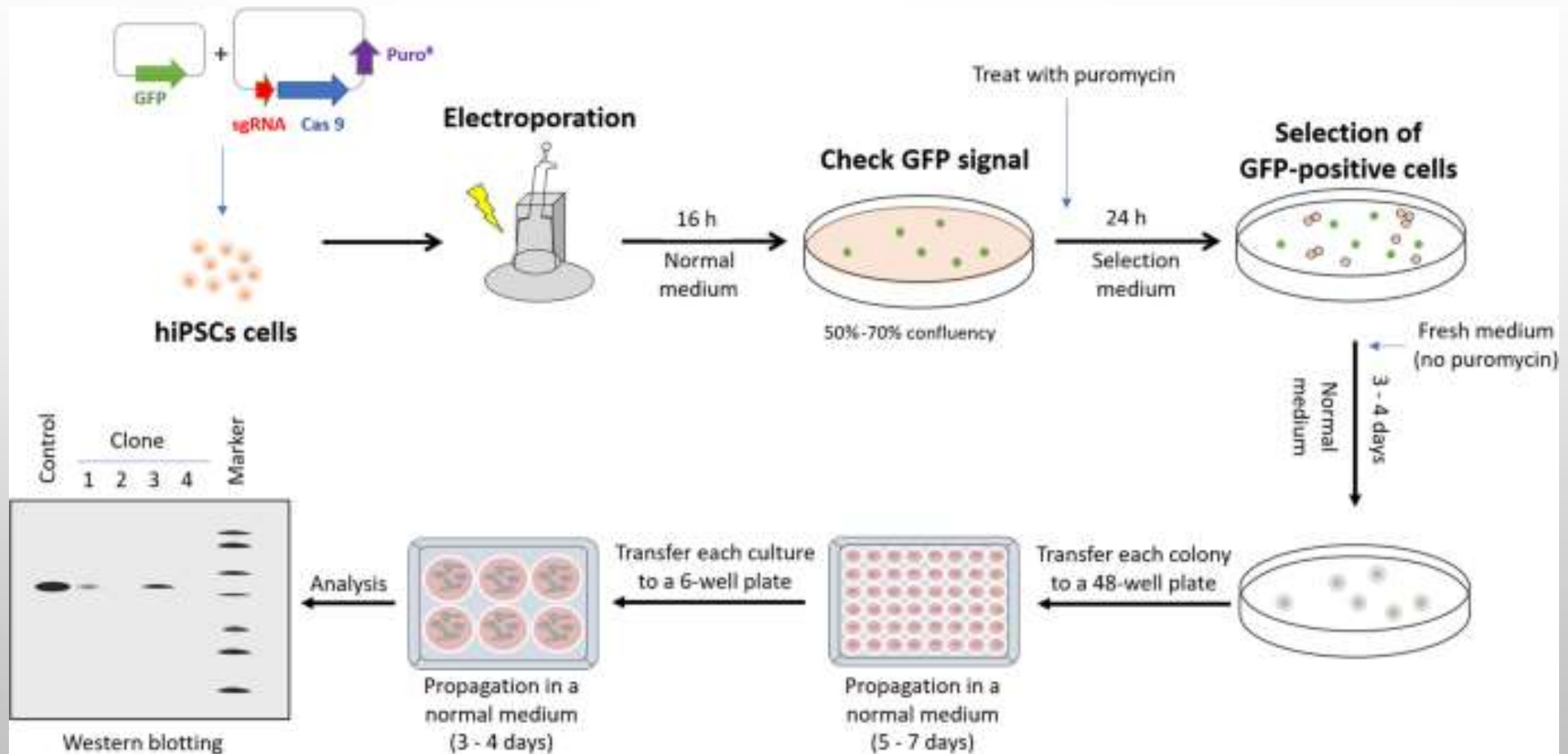


https://www.youtube.com/watch?v=pNseb_U6gu0

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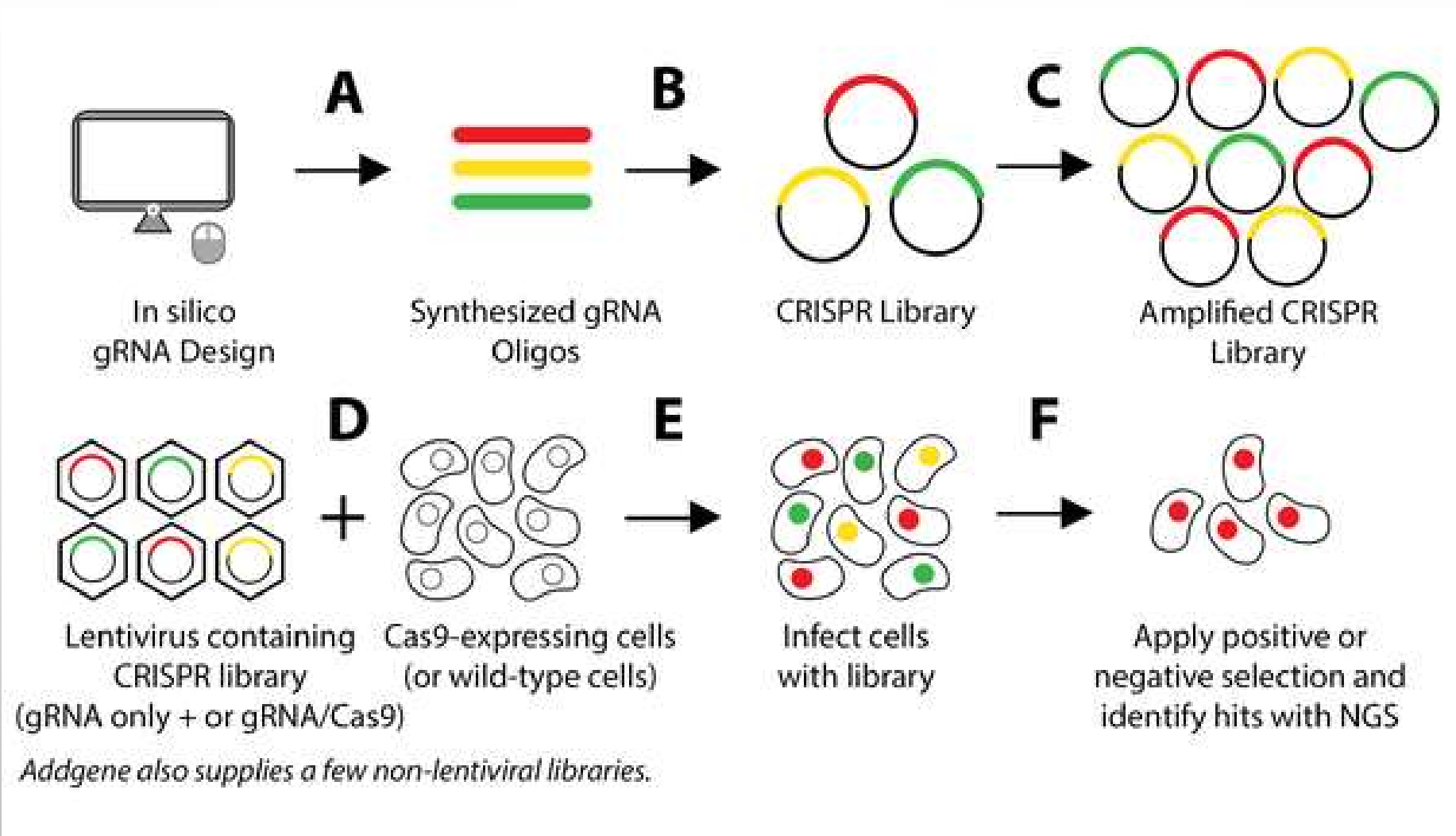
CRISPR protocol

(a) single sgRNA



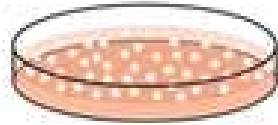
Acta Pharmacologica Sinica volume 41, pages1427–1432 (2020)

(b) CRISPR sgRNA library screening

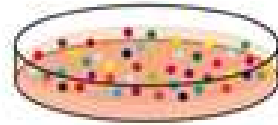


A

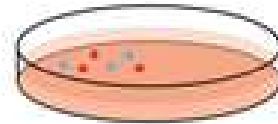
Infect cell line with pooled sgRNA library lentiviral particles



Screen plates for response of phenotype of interest

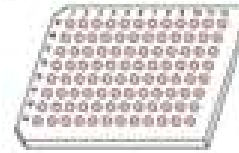


PCR and/or sequence individual sgRNA(s)

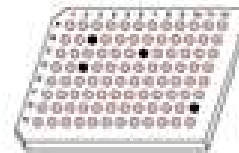
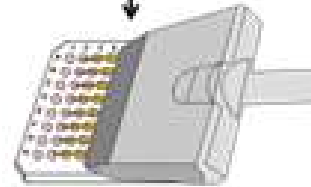


B

Infect cell line in single wells with individual sgRNA library lentiviral particles



Screen plates for response of phenotype of interest



Identity of each individual sgRNA in each well is known without PCR or sequencing

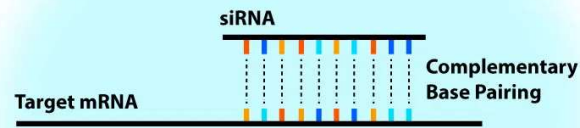
Compared to RNAi

CRISPER VERSUS RNAi siRNA

CRISPER	RNAi
The hallmark of a bacterial defense system that forms the basis for CRISPR-Cas9 genome editing technology	A biological process in which RNA molecules inhibit gene expression or translation, by neutralizing targeted mRNA molecules
Naturally occurs in prokaryotes	Naturally occurs in many eukaryotes
A genome editing technology, involving in the knocking out of genes	A form of post-transcriptional regulation of gene expression, involving in the knocking down of gene expression
Applicable in the DNA level	Applicable in the RNA level
Silences genes permanently	Silences genes temporarily
High cost	Low cost
Low off-target effects	High off-target effects
	Visit www.PEDIAA.com

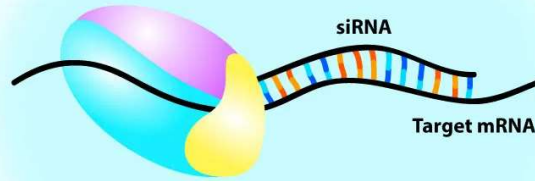
<https://cn.weblogographic.com/what-is-difference-between-crispr>

RNAi

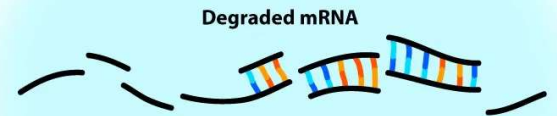


1) Small interfering RNA (siRNA) is designed to specifically target mRNA that's derived from a particular gene.

RISC Protein Complex

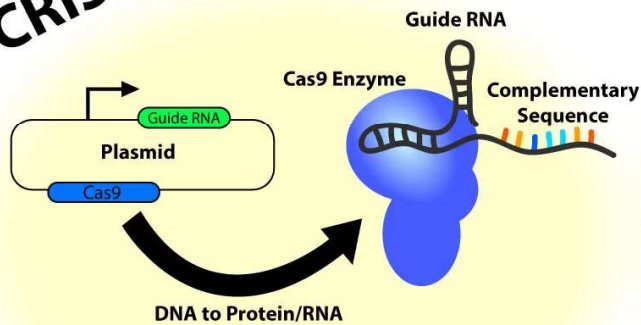


2) Once it's been introduced into a cell, siRNA binds to target mRNA with the help of a piece of cellular machinery called the RISC protein complex.

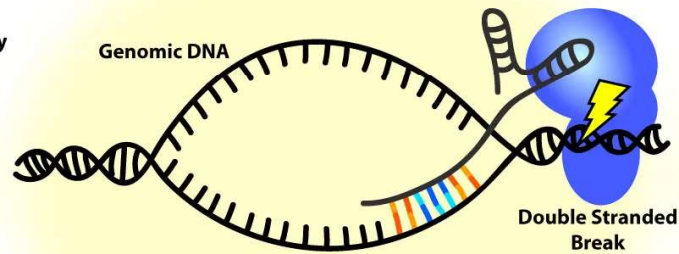


3) After binding, the siRNA promotes degradation of the target mRNA, resulting in translational repression and lower levels of the corresponding protein.

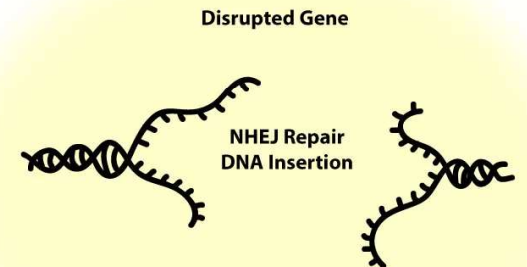
CRISPR



1) A plasmid, a circular piece of DNA, encodes for both the Cas9 enzyme and a "guide RNA" that's complementary to the target gene's DNA sequence.



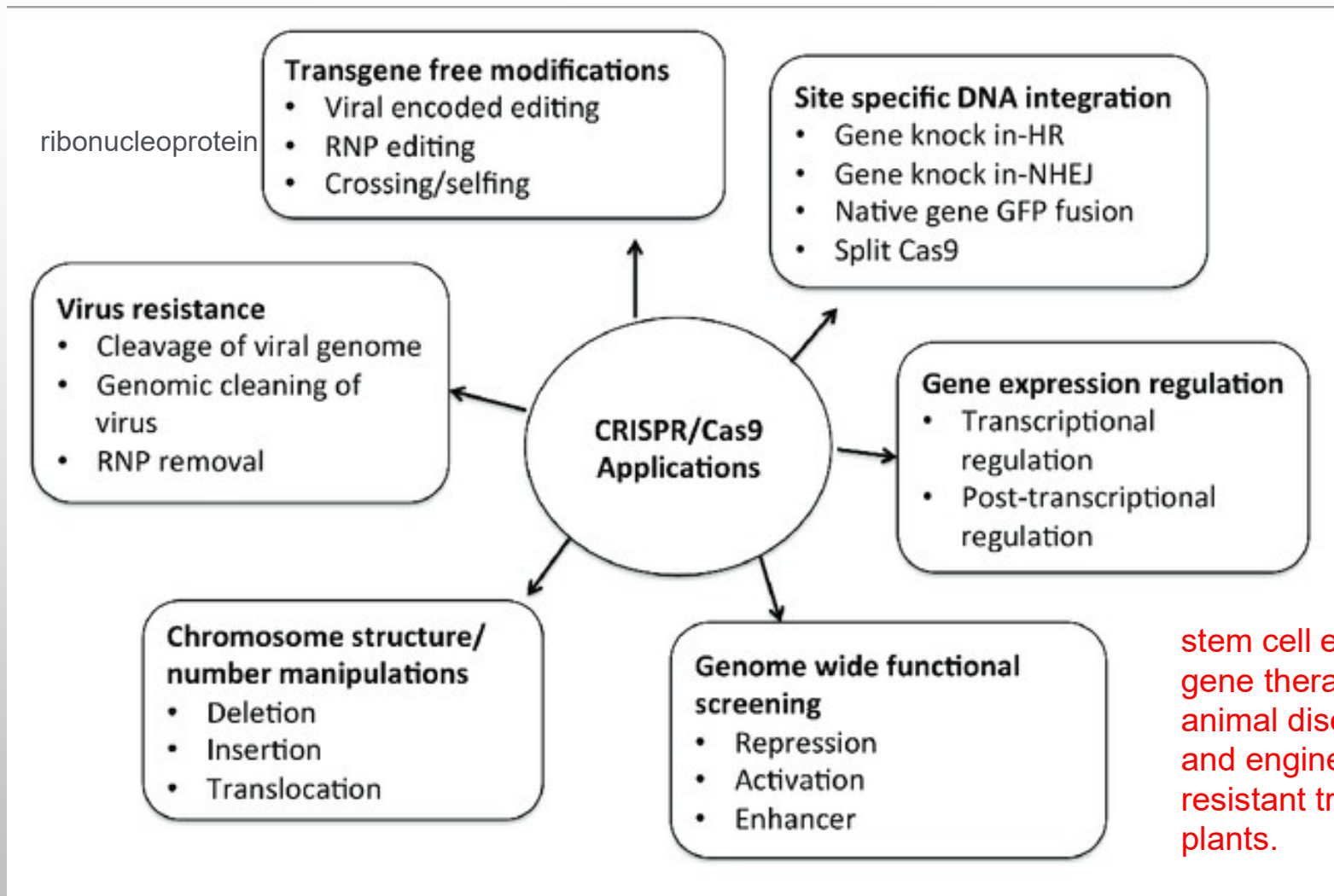
2) After the plasmid enters the cell, the resulting Cas9/guide RNA complex binds the target DNA sequence and Cas9 cuts both strands of DNA.



3) The cell repairs the DNA break via non-homologous end joining (NHEJ), which is prone to errors that can silence the gene, or homology directed repair, which enables high-precision editing of the gene.

by Christopher Gerry

Applications of CRISPR system



Genome analysis (基因體分析)

(1) Open reading frame(ORF): 開放讀序框架

- 基因組中能被轉譯成氨基酸序列的DNA片段
- 一條DNA序列共含有六種可能性的讀取框架(reading frame)
- 框架讀取：每3個鹼基為一組
- 若讀取框架中，含有起始(ATG, Methionine)及終止碼(TAA, TAG, TGA),且該讀取框架夠長，則其可能為一個基因
- 若讀取框架片段沒有包含起始或終止碼，則該片段則有可能是某基因片段

AAACCATGCTAAATCTGGTAGGCAAGCAGTTGTGAAAA

Frame 1 -> K P C - I W - A S S C E

Frame 2 -> N H A K S G R Q A V V K

Frame 3 -> T M L N L V G K Q L - K

Frame 4 -> G H - I Q Y A L L Q S F <- Frame 4

Frame 5 -> V M S F R T P L C N H F <- Frame 5

Frame 6 -> F W A L D P L C A T T F <- Frame 6

Q：假設你有一條來自大腸桿菌序列如下，請問你要如何以尋找ORF的方式預測該片段是否含有基因片段？

aaacggctcatcgtcttaaaggcgtatttgccatgctaaatctggtacccggcaagcagttat
gtgaaacgctggaacatctgattcgtgagaaggatgttccaggaatagaaaaatacatcag
cgacattgacagttatgtcaagagcttgctgtagcaaggtagcctattacatgaacaatatg
aacgtaattattgccgatgaccatccgatagcttctggtattcgcaaatcacttgagcaa
attgagtgggtgaatggtgtcggcgaatttgaagactctacagcactgatcaacaacctgcc
gaaactggatgcgcatgtgttgattaccgatctctccatgcctggcgataagtacggcgatg
gcattaccttaatcaagtacatcaagcgccatttccaagcctgtcgatcattgttctgactat
gaacaacaaccggcgattcttagtgcggtattggatctggatatcgaagggatcgtgctga
aacaaggtgcaccgaccgatctgccgaaagctctcgccgcgctgcagaaagggaagaaat
ttaccccggaagcgtttctcgccctgttggaaaaaatcagtgctggtggttacggtgacaag
cgtctctcgccaaaagagagtgaagttctgcgcctgtttgcggaaggcttcctggtgaccga
gatcgctaaaaagctgaaccgcagtattaaaccatcagtagccagaagaaatctgcgatg
atgaagctgggtgtcgagaacgatatcgccctgctgaattatctctcttcagtgaccttaagt
ccggcagataaagactaatcacctgtaggccagat

ORF預測軟體 — TRANSLATE TOLLS IN EXPASY

<https://web.expasy.org/translate/>

Expasy

Translate

Home | Contact

Programmatic access ↓

Translate is a tool which allows the translation of a nucleotide (DNA/RNA) sequence to a protein sequence.

DNA or RNA sequence

```
cgaatttgaagactctacagcactgatcaacaacctgccgaaactggatgcccattgtgttattaccg  
atctctccatgcttggcgataagtagcggcgatggcattaccttaatacaagtacatcaagcgcatttc  
ccaagcctgtcgatcattgttctgactatgaacaacaaccggcgattcttagtgcggtattggatct  
ggatcgcgaaggatcgtgctgaacaaggtagcaccgaccgatctgccgaaagctctcggcgcgctgc  
agaaggggaagaaatttaccgggaaagcgtttctcgcctgttgaaaaaatcagtgcgtggtgttac  
ggtgacaagcgtctctcgcgcaaaagagagtgaagtctcgcgctgtttgcggaaggcttctggtgac  
cgagatcgttaaaaagctgaaccgcagtattaaaaccatcagtagccagaagaatctgccgatgatga  
agctgggtgtcgagaacgatatcgcctgctgaattatctctctcagtgaccttaagtcggcagat  
aaagactaatcacctgtaggccagat
```

待預測序列

Output format

- Verbose: Met, Stop, spaces between residues
- Compact: M, -, no spaces
- Includes nucleotide sequence
- Includes nucleotide sequence, no spaces

DNA strands

- forward
- reverse

Genetic codes - See NCBI's genetic codes

Standard

reset

TRANSLATE!

開始預測

ORF預測結果

5'3' Frame 1

KRLIVLKAYLPC-IWYPASSYVKRWNI-FVRRMFQE-KNTSATLTVMSRACCSKVAYYMNNMNVIIADDHPIVLFGIRKSLEQIEWVNVVGEFEDSTAL
INNLPKLDHAVLITDLSMPGDKYGDGITLIKIYIKRHFPKLSIIIVLTMNNNPAILSAVLDLDIEGIVLKQGAPTDLPKALAALQKGGKFTPEVSRLLEK
ISAGGYGDKRLSPKESEVLRLFAEGFLVTEIAKKNLRSIKTISSQKKSAMMGLGVENDIALLLNYLSSVTLSPADKD-SPVGQ

5'3' Frame 2

NGSSS-RRICHAKSGTRQAVM-NAGTSDS-EGCSRNRKIHQRH-QLCQELAVAR-PIT-TI-T-LLPMTIR-SCSVFANHLSKLSG-MLSANLKTLOH-
STTCRNWMRMC-LPISPCLAISTAMALP-SSTSSAISQACRSLF-L-TTTRRFLVRYWIWISKGSC-NKVHRPICRKLSPRCRKRGNLPRKAFLACWKK
SVLVVTVTSVSRQKRVKFCACLRKASW-PRSLKS-TAVLKPSVARRNLR--SWVSRTISPC-IISLQ-P-VRQIKTNHL-AR

5'3' Frame 3

TAHRLKGVFAMLNLVPGKQLCETLEHLIREKDVPKIEKYISDIDSYVKSLL-QGSLLEHQYERNYCR-PSDSLVRYSQIT-AN-VGECRRRI-RLYSTD
QQPAETGCACVDYRSLHAWR-VRRWHYLNQVHQAPFPKVDHCSDEYQQPGDS-CGIGSGYRRDRAETRCTDRSAESSRRAAEREIYPGKRFSPVGKN
QCWWLR-QASLAKRE-SAPVCGRLPGDRDR-KAEPQY-NHQ-PEEICDDEAGCRERYRPAELSLFSDLKSGR-RLITCRPD

3'5' Frame 1

IWPTGD-SLSAGLKVTEER-FSRAISFSTPSFIIADFFWLLMVLILRFSFLAISVTRKPSANRRRTSLSFGERRLSP-PPALIFSNRRETLSGVNFFPF
CSAARAFGRSVGAPCFSTIPSISSRNTALRIAGLLFIVRTMIDRLGKWRMLYLIKVMPSPYLSPGMERSVINTCASSFGRLISAVESSNSPTTFTHSI
CSSDLRIPNKTIGWSSAIITFILEM--ATLLQQALDITVNVADVFFYSWNILLTNQMFQRF-LLAGYQI-HGKYAFKMSR

3'5' Frame 2

SGLQVISLILPDLRSLKRDNSAGRYRSRHPASSQISSGY-WF-YCGSAF-RSRSPGSLPQTGAELHSLLDACHRNHQH-FFPTGKRFPG-ISLSL
AARRELSADRSVHLVSARSLRYPDPIPH-ESPGCCS-SEQ-STGLGNGA-CT-LR-CHRRTYRQAWRDR-STHAHPVSAGC-SVL-SLQIRRHQSPTQF
AQVICEYRTRLSDGHRQ-LRSYCSCNRLPCYSKLLT-LSMSLMYFSIPGTSFSRIRCSSVSHNCLPGTRFSMANTPLRR-AV

3'5' Frame 3

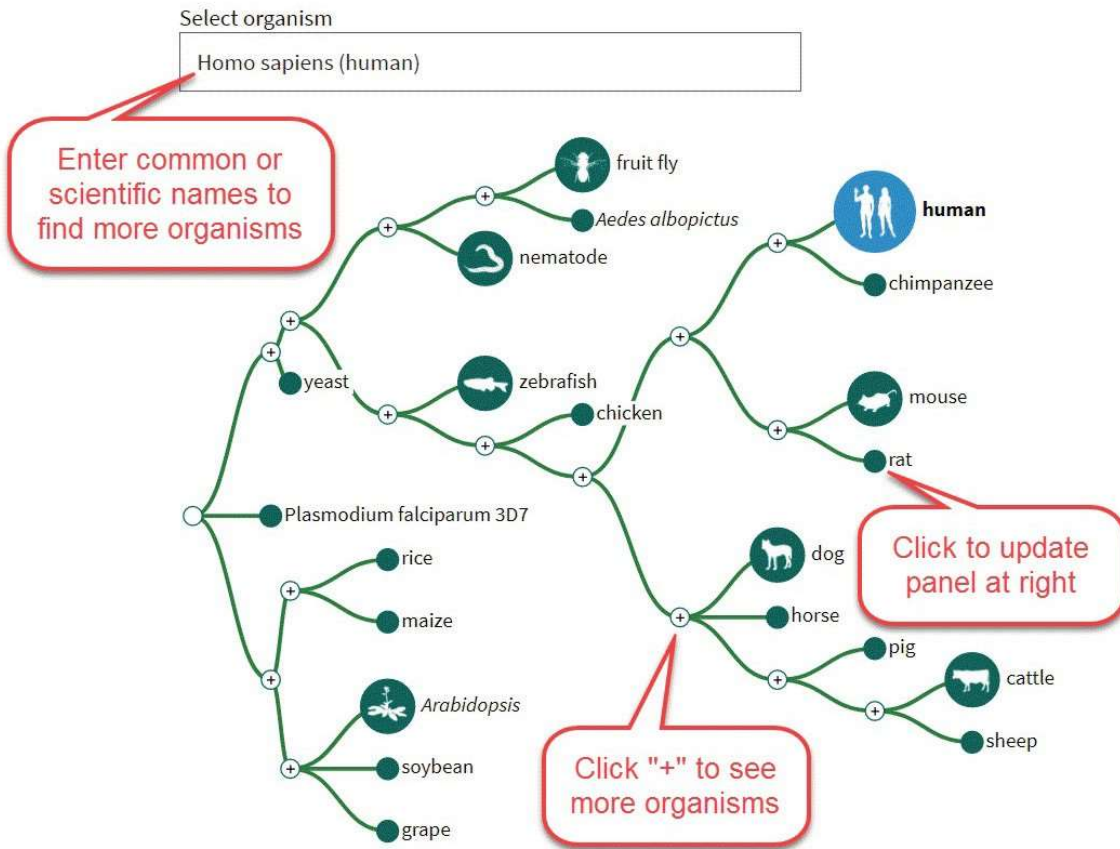
LAYR-LVFI CRT-GH-REIIQQGDIVLDTQLHHRFFLLATDGFNTAVQLFSDLGHQEAFRKQAQNFLLFWRETIVTVTTSTDDFFQQARNAFRGKFLPFL
QRGESFRQIGRCTLFQHDPDFDIQIQYRTKNRRVVHVSQNNDRQAWEMALDVLG-GNAIAVLIARHGEIGNQHMRIQFRQVVDQCCRVFKFADNIHPLNL
LK-FANTEQDYRMVIGNNYVHIVHIVIGYLATASS-HNCQCR-CIFLFLEHPSHESDVPAFHITACRVPLAWQIRL-DDEPF

(2) NCBI genome data viewer

<https://www.ncbi.nlm.nih.gov/gdv/>

Genome Data Viewer

GDV is a genome browser supporting the exploration and analysis of more than 540 eukaryotic RefSeq genome assemblies. ⓘ



Homo sapiens (human) genome

Search within selected assembly

Search in genome
Location, gene or phenotype

Examples: TP53, chr17:7667000-7689000, rs334, DNA repair

Assembly
GRCh38.p11

Browse genome BLAST genome

Select assembly version

Assembly details

Name	GRCh38.p11
RefSeq accession	GCF_000001405.37
GenBank accession	GCA_000001405.26
Download via FTP	RefSeq, GenBank
Submitter	Genome Reference Consortium
Level	Chromosome

Annotation details

Annotation Release 108
Release date

Homo sapiens (human) genome



Search in genome

aquaporin



Search "Aquaporin"

Genes

Other

Name	Location
AQP4	Chr18: 26.85M - 26.87M
AQP1	Chr7: 30.91M - 30.93M
AQP2	Chr12: 49.95M - 49.96M
AQP3	Chr9: 33.44M - 33.45M
AQP5	Chr12: 49.96M - 49.97M
AQP9	Chr15: 58.14M - 58.19M
AQP8	Chr16: 25.22M - 25.23M

result

Examples: [TP53](#), [chr17:7667000-7689000](#), [rs334](#), [DNA repair](#)

Assembly

GRCh38.p12

Chromosome 7

location: 7p14.3
(cytological map)

Transcript ID

Current position
(physical map)

Genome Data Viewer

Homo sapiens: GRCh38.p12 (GCF_000001405.38) Chr 7 (NC_000007.14): 30,910,312 - 30,926,899

Reset All Share this page FAQ Help Browser Agreement Version 4.5

Region: AQP1 NM_001329872.1
Gene Transcript
Exon cds range: 30,911,910 - 30,912,293, range: 30,911,694 - 30,912,293

NC_000007.14

Genes, NCBI Homo sapiens Annotation Release 109, 2018...
AQP1 [+4]

Genes, Ensembl release 93
000240583 ... ENSP00000421315.2

dbSNP Build 151 (Homo sapiens Annotation Release 108) all data

Cited Variants, dbSNP Build 150 (Homo sapiens Annotat...
1 1 2 1 1 1 1

RNA-seq exon coverage, aggregate (filtered), NCBI Homo sapiens Annotation Release 109 - log base 2 scaled
104171

RNA-seq intron-spanning reads, aggregate (filtered), NCBI Homo sapiens Annotation Release 109 - log
40884

Feedback

Gene structure

Search result

(3) Restriction Enzyme Tool-NEBcutter

<http://nc2.neb.com/NEBcutter2/>



NEBcutter V2.0



This tool will take a DNA sequence and find the large, non-overlapping open reading frames using the E.coli genetic code and the sites for all Type II and commercially available Type III restriction enzymes that cut the sequence just once. By default, only enzymes available from NEB are used, but other sets may be chosen. Just enter your sequence and "submit". Further options will appear with the output. **The maximum size of the input file is 1 MByte, and the maximum sequence length is 300 KBases.**

[What's new in V2.0](#) [Citing NEBcutter](#)

Local sequence file: 選擇檔案 未選擇任何檔案

GenBank number: [\[Browse GenBank\]](#)

Standard sequences:
Plasmid vectors ▾
Viral + phage ▾

or paste in your DNA sequence: (plain or FASTA format)

```
gagttcgcgg cgggtggcgg cgggtggcgc acgcgcgacc tagggatcga tctggagggg  
61 cttggggagc gtcagagac ctctagctcg agcgcgaggg acctccgcc  
gggatgcctg  
121 gggagcagat ggaccctact ggaagtcagt tggattcaga tttctctcag  
caagatactc  
181 cttgctgat aattgaagat tctcagcctg aaagccaggt tctagaggat  
gattctggtt  
241 ctcacttcag tatgctatct cgacaccttc ctaatctcca gacgcacaaa
```

The sequence is: Linear Circular

Enzymes to use:
 NEB enzymes
 All commercially available specificities
 All specificities
 All + defined oligonucleotide sequences
 Only defined oligonucleotide sequences
[\[define oligos\]](#)

Minimum ORF length to display: a.a.

Name of sequence: (optional)

Earlier projects:

Note: Your earlier projects will be deleted 2 days after they were last accessed.

NEBcutter V2.0

This tool will take a DNA sequence and find the large, non-overlapping open reading frames using the E.coli genetic code and the sites for all Type II and commercially available Type III restriction enzymes that cut the sequence just once. By default, only enzymes available from NEB are used, but other sets may be chosen. Just enter your sequence and "submit". Further options will appear with the output. **The maximum size of the input file is 1 MByte, and the maximum sequence length is 300 KBases.**

[What's new in V2.0](#) [Citing NEBcutter](#)

Local sequence file: <input type="text" value="選擇檔案 未選擇任何檔案"/>	Standard sequences:
GenBank number: <input type="text"/> [Browse GenBank]	# Plasmid vectors ▾
or paste in your DNA sequence: <i>(plain or FASTA format)</i>	# Viral + phage ▾
<div style="border: 1px solid gray; height: 100px; width: 100%;"></div>	<input type="button" value="Submit"/>
The sequence is: <input type="radio"/> Linear <input checked="" type="radio"/> Circular	<input type="button" value="More options"/>
Enzymes to use:	<input type="button" value="Set colors"/>
<input checked="" type="radio"/> NEB enzymes	
<input type="radio"/> All commercially available specificities	
<input type="radio"/> All specificities	
<input type="radio"/> All + defined oligonucleotide sequences	
<input type="radio"/> Only defined oligonucleotide sequences	
[define oligos]	
Minimum ORF length to display: <input type="text" value="100"/> a.a.	
Name of sequence: <input type="text"/> <i>(optional)</i>	
Earlier projects:	

Note: Your earlier projects will be deleted 2 days after they were last accessed.

Nucleotide

Nucleotide

P53

Search

Help



Using Nucleotide

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[GenBank FTP](#)

[RefSeq FTP](#)

- Homo sapiens cellular tumor antigen p53
- Homo sapiens tumor protein p53
- Mus musculus cellular tumor antigen p53
- Cricetulus griseus cellular tumor antigen p53
- Cricetulus griseus tumor protein p53
- Canis lupus familiaris cellular tumor antigen p53
- Canis lupus familiaris tumor protein p53
- Xenopus laevis cellular tumor antigen p53
- Xenopus laevis tumor protein p53 L homeolog
- Rattus norvegicus cellular tumor antigen p53
- Rattus norvegicus tumor protein p53
- Danio rerio cellular tumor antigen p53
- Danio rerio tumor protein p53
- Macaca mulatta cellular tumor antigen p53
- Macaca mulatta tumor protein p53
- Bos taurus cellular tumor antigen p53
- Bos taurus tumor protein p53
- Drosophila melanogaster p53
- Pan troglodytes cellular tumor antigen p53
- Homo sapiens tumor protein p53 binding protein 1

GenBank, RefSeq, TPA and
research and discovery.

You are here: NCBI > DNA & RNA > Nucleotide Database

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Nucleotide

Nucleotide

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Homo sapiens tumor protein p53 binding protein 1 (TP53BP1), transcript variant 1, mRNA

NCBI Reference Sequence: NM_001141980.3

[FASTA](#) [Graphics](#)

Go to

Analyze this sequence

Run BLAST

Pick Primers

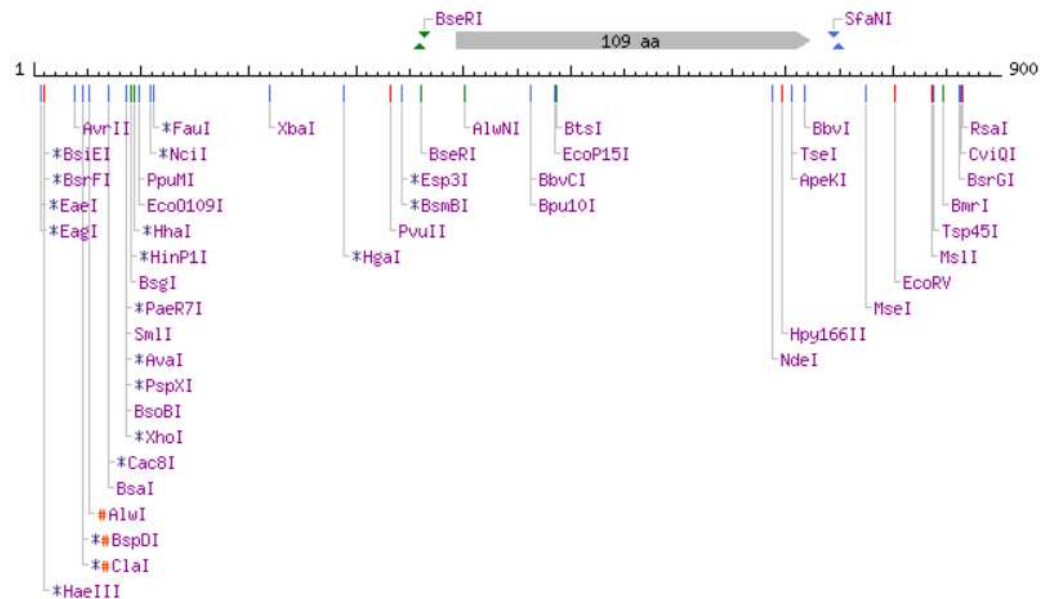
Highlight Sequence Features

LOCUS NM_001141980 10369 bp mRNA linear PRI 08-SEP-2019
 DEFINITION Homo sapiens tumor protein p53 binding protein 1 (TP53BP1), transcript variant 1, mRNA.
 ACCESSION NM_001141980
 VERSION NM_001141980.3
 KEYWORDS RefSeq; RefSeq Select.
 SOURCE Homo sapiens (human)
 ORGANISM [Homo sapiens](#)
 Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglires; Primates; Haplorrhini; Catarrhini; Hominidae; Homo.
 REFERENCE 1 (bases 1 to 10369)
 AUTHORS Hurley RM, Wahner Hendrickson AE, Visscher DW, Ansell P, Harrell MI, Wagner JM, Negron V, Goergen KM, Maurer MJ, Oberg AL, Meng XW, Flatten KS, De Jonge MJA, Van Herpen CD, Gietema JA, Koonstra RHT, Jager A, den Hollander MW, Dudley M, Shepherd SP, Swisher EM and Kaufmann SH.
 TITLE 53BP1 as a potential predictor of response in PARP inhibitor-treated homologous recombination-deficient ovarian cancer
 JOURNAL Gynecol. Oncol. 153 (1), 127-134 (2019)
 PUBMED [30686551](#)
 REMARK GeneRIF: 53BP1 deletion increased HR in BRCA1-mutant COV362 cells

```

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/ inference="alignment:Splign:2.1.0"
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exon
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/ inference="alignment:Splign:2.1.0"
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ORIGIN
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61 cttggggagc gtcagagac cttctagctc agcgcgaggg acctcccgcc gggatgcctg
121 gggagcagat ggaccctact ggaagtcagt tggattcaga ttctctcag caagatactc
181 cttgcctgat aattgaagat tctcagcctg aaagccaggt tctagaggat gattctggtt
241 ctcacttcag tatgctatct cgacaccttc ctaatctcca gacgcacaaa gaaaatcctg
301 tgttggatgt tgtgtccaat cctgaacaaa cagctggaga agaacgagga gacggaata
361 gtgggttcaa tgaacattg aaagaaaaca aggttcgaga ccctgtggat tcttctaact
421 tggacacatg tggttccatc agtcaggtca ttgagcagtt acctcagcca aacaggacaa
481 gcagttttct gggaatgtca gtggaatctg ctcctgctgt ggggaaagag aagggaagag
541 agttggaaca gaaggagaaa gagaaggaag aagatacttc aggcataact acacattccc
601 ttggtgctga agatactgcc tcatcacagt tgggttttgg ggttctggaa ctctccaga
661 gccaggatgt tgaggaaaat actgtgccat atgaagtgga caaagagcag ctacaatcag
721 taaccaccaa ctctggttat accaggctgt ctgatgtgga tgctaatact gcaattaagc
781 atgaagaaca gtccaacgaa gatatcccca tagcagaaca gtccagcaag gacatccctg
841 tgacagcaca gcccgtaag gatgtacatg ttgtaaaaga gcaaaatcca ccacctgcaa
901 ggtcagagga catgctttt agcccaaaag catctgttgc tgctatggaa gcaaaaagac
961 agttgtctgc acaagaaact atggaagtg gactgcagat tcagaagta ccaagcctg
1021 aggttttgtc aactcaggaa gacttgtttg accagagcaa taaaacagta tcttctgatg
1081 gttgctctac tcttcaagg gaggaaagtg ggtgttcttt ggcttccact cttgccacca
1141 cttctgatct cctgcagctc tctggtcaga ggtcccttgt tcaggacagt tcttccaga
1201 attcttcaga tcttgttct ctttctcctg atgctttcag atctactctt tttatcgttc
1261 ctgacagctc cacagagcaa gaaggagac aagataagcc aatggacacg tcagtgattat
1321 ctgaagaagg aggagagcct tttcagaaga aacttcaag tggtaacca gtggagttag
1381 aaaaaccccc tctctgctc gagtccaact taccaccaca agcctcaaca ccaatattct
1441 agagcacacc agtcttccct cctgggtcac ttcctatccc atcccagcct cagttttctc
1501 atgacatttt tattccttcc ccaagtctgg aagaacaatc aaatgatggg aagaaagatg
1561 gagatatgca tagttcatct ttgacagttg agtgttctaa aacttcagag attgaaccaa
1621 agaattcccc tgaggatctt gggctatctt tgacagggga tcttgcagag ttgatgcttt
1681 ctacaagtga atatagtcag tccccaaaga tggagagcct gagttctcac agaattgatg
1741 aagatggaga aaacacacag attgaggata cggaacccat gtctccagtt tcaattctta
1801 aatttgttcc tgctgaaatg gatagtatcc tgatgatacc agcaccagat ggtgaagtc
    
```

Cleavage code	Enzyme name code
▼ blunt end cut	Available from NEB
▼ 5' extension	Has other supplier
▼ 3' extension	Not commercially available
▼ cuts 1 strand	*: cleavage affected by CpG meth. #: cleavage affected by other meth. (enz.name): ambiguous site



Main options	Availability	Display	Zoom	List
New DNA Custom digest View sequence ORF summary Save project Print	All commercial All	2 cutters 3 cutters	Zoom in More...	0 cutters 1 cutters All sites Save all sites Flanking enzymes
Minimum ORF length to display: <input type="text" value="100"/> aa. <input type="button" value="OK"/>				

Double digest finder

NEBcloner v1.3.13

FEEDBACK ? HELP HISTORY

Home Page / RE Digest

Restriction Enzyme Single/Double Digestion

Select Enzyme

Select 2nd Enzyme [✕ clear 2nd selection](#)

Please select an enzyme to view the protocol.

[Show Detailed Protocol](#)

Name	Cat #	Temp °C	Supplied Buffer	Add SAM	% Activity in NEBuffer			
					1.1	2.1	3.1	CutSmart ®

Name	Time-Saver™	Heat Inactivation (°C)	Methylation Sensitivity

Restriction Digest Resources

- [Tool: Enzyme Finder](#)
- [Tool: NEBcutter](#)
- [Info: Troubleshooting Guide](#)
- [Info: Restriction Enzyme Tips](#)
- [Info: Double Digests](#)
- [Info: RE Performance Chart](#)
- [Info: Dam-Dcm and CpG Methylation](#)
- [Video: Cloning with Restriction Enzymes](#)

Search for product name/number

EcoRI

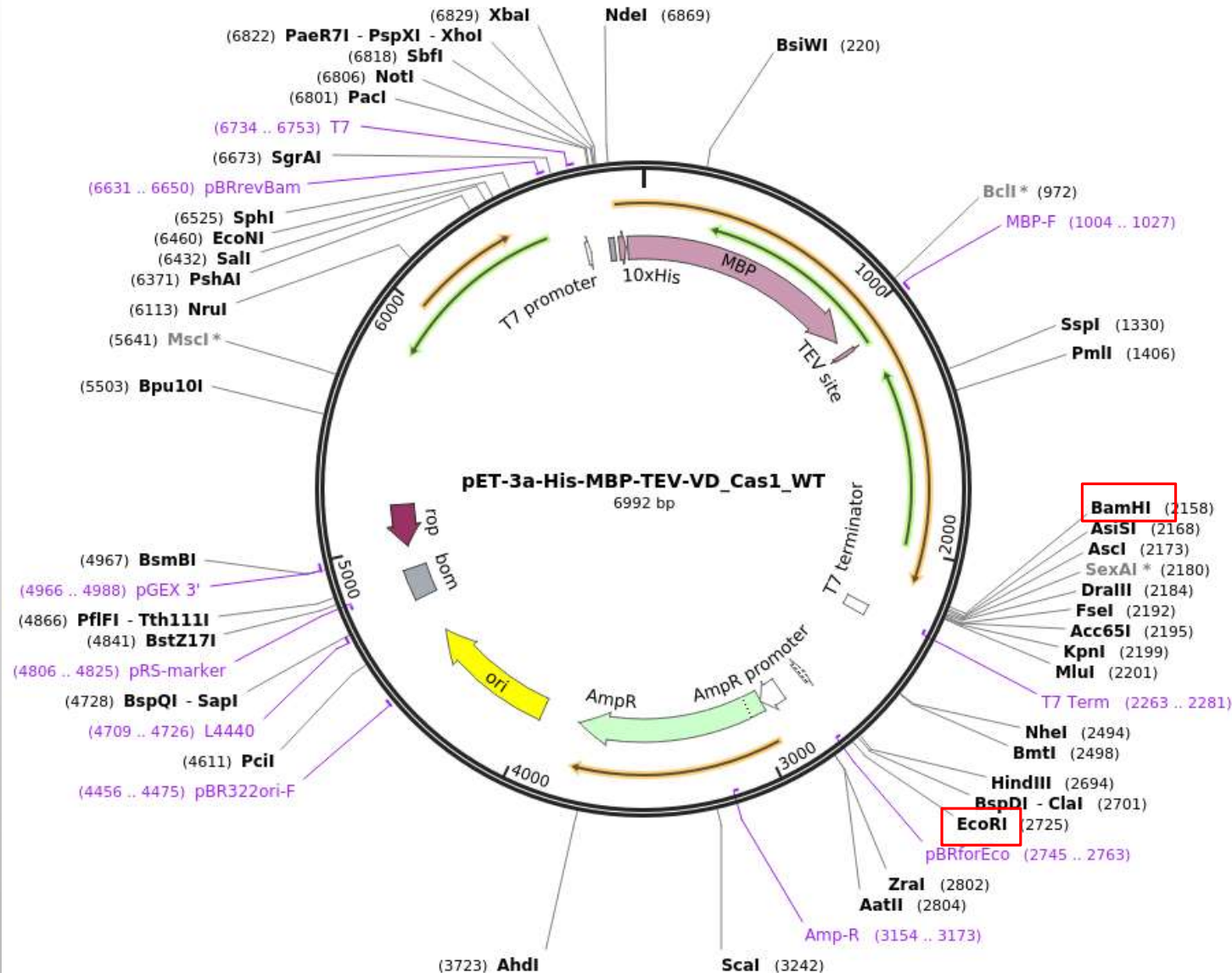
No results

Learn about traditional cloning

- [Info: Getting Started / Cloning Guidelines](#)
- [Info: Traditional Cloning Workflow](#)
- [Video: Traditional Cloning](#)

PET3A PLASMID

Created with SnapGene®



Two Fragments:

4128 bp

512 bp

Search for product name/number

No results

Learn about traditional cloning



[Info: Getting Started / Cloning Guidelines](#)



[Info: Traditional Cloning Workflow](#)



[Video: Traditional Cloning](#)

[Home Page](#) / RE Digest

Restriction Enzyme Single/Double Digestion

[✕ clear 2nd selection](#)

Digest in NEBuffer 3.1

[Show Detailed Protocol](#)

Name	Cat #	Temp °C	Supplied Buffer	Add SAM	% Activity in NEBuffer			
					1.1	2.1	3.1	CutSmart®
EcoRI	R0101	37	NEBuffer EcoRI	No	25	100*	50	50*
BamHI	R0136	37	NEBuffer 3.1	No	75*	100*	100	100*

* May exhibit star activity in this buffer.

Name	Time-Saver™	Heat Inactivation (°C)	Methylation Sensitivity
EcoRI	Yes	65	cpg (Blocked by Some Combinations of Overlapping)
BamHI	Yes	No	None

Notes:

1. Digest in NEBuffer 3.1 (or NEBuffer 3 + BSA) at 37 °C.
2. At least one enzyme has < 100% activity in this buffer, so additional units of enzyme and/or longer incubation time may be necessary.
3. EcoRI has a High Fidelity version EcoRI-HF. BamHI has a High Fidelity version BamHI-HF. High Fidelity (HF) Restriction Enzymes have been engineered for reduced star activity and have 100% activity in CutSmart Buffer which may simplify your double digest.

MORE OPTIONS



NEBcutter V2.0



This tool will take a DNA sequence and find the large, non-overlapping open reading frames using the E.coli genetic code and the sites for all Type II and commercially available Type III restriction enzymes that cut the sequence just once. By default, only enzymes available from NEB are used, but other sets may be chosen. Just enter your sequence and "submit". Further options will appear with the output. **The maximum size of the input file is 1 MByte, and the maximum sequence length is 300 KBases.**

[What's new in V2.0](#) [Citing NEBcutter](#)

Local sequence file: 未選擇任何檔案

GenBank number: [\[Browse GenBank\]](#)

or paste in your DNA sequence: (plain or FASTA format)

```
gagttcgcgg cgggtggcgg cgggtggcgc acgcgcgacc tagggatcga tctggagggg
61 cttggggagc gtcagagac ctctagctcg agcgcgaggg acctccgcc
gggatgcctg
121 gggagcagat ggaccctact ggaagtcagt tggattcaga tttctctcag
caagatactc
181 cttgcctgat aattgaagat tctcagcctg aaagccaggt tctagaggat
gattctggtt
241 ctcacttcag tatgctatct cgacaccttc ctaatctcca gacgcacaaa
```

The sequence is: Linear Circular

Enzymes to use: NEB enzymes
 All commercially available specificities
 All specificities
 All + defined oligonucleotide sequences
 Only defined oligonucleotide sequences
[\[define oligos\]](#)

Minimum ORF length to display: a.a.

Name of sequence: (optional)

Earlier projects:

Note: Your earlier projects will be deleted 2 days after they were last accessed.

Standard sequences:
Plasmid vectors ▾
Viral + phage ▾

A **nicking enzyme** (or **nicking endonuclease**) is an **enzyme** that cuts one strand of a double-stranded **DNA** at a specific recognition **nucleotide** sequences

In addition to Type II and commercially available Type III enzymes, also look for:

- Type I & III enzymes
- Homing endonucleases**
- Nicking enzymes

Ignore CpG methylation Ignore EcoBI methylation
 Ignore Dam methylation Ignore EcoKI methylation
 Ignore Dcm methylation

Genetic code to use when searching for ORFs:

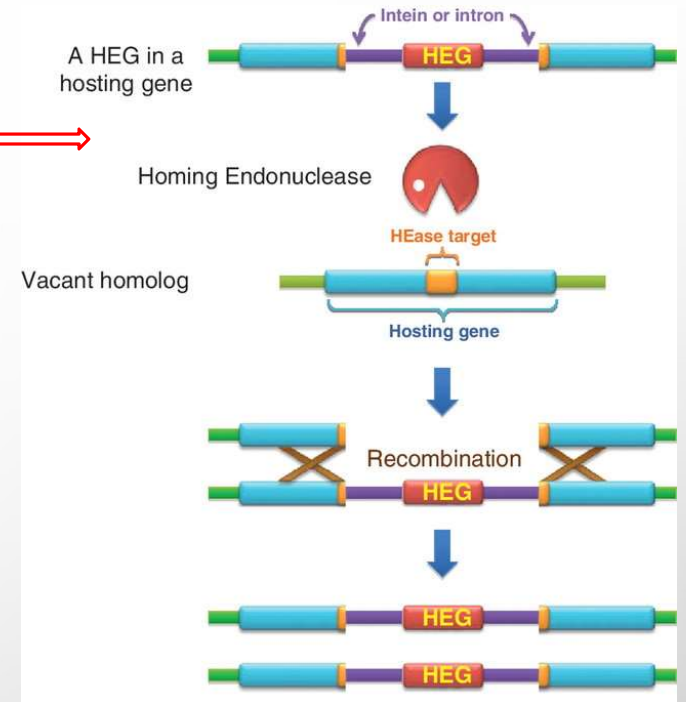
11. Bacterial and Plant Plastid

Sequence is a fragment

Process this region only: - bp

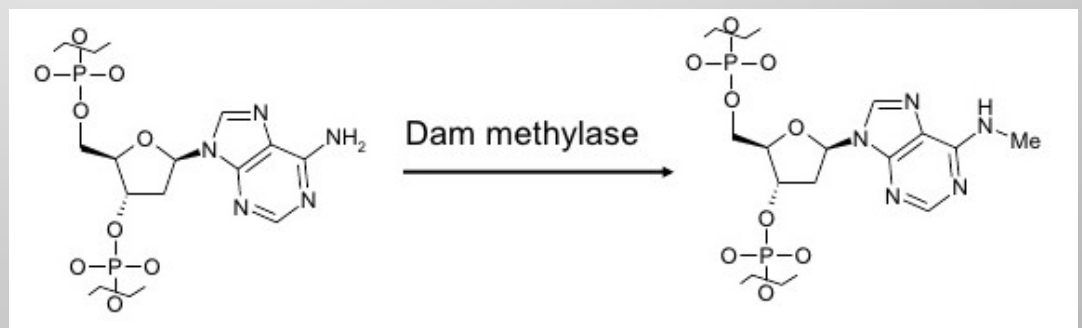
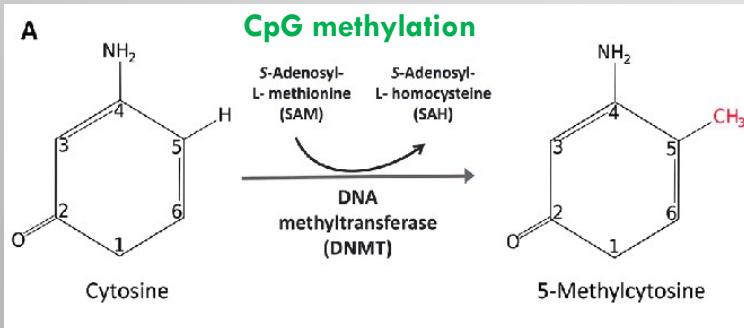
OK

Cancel



EcoKI or **EcoBI** methyltransferases modify adenine residues within their respective recognition sequences:

AAC(N)6GTGC for **EcoKI** and TGA(N)8TGCT for **EcoBI**



Set Colors



NEBcutter V2.0



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[What's new in V2.0](#) [Citing NEBcutter](#)

Local sequence file: 未選擇任何檔案

GenBank number: [\[Browse GenBank\]](#)

or paste in your DNA sequence: (plain or FASTA format)

```
gagttcggg cgggtggcgg cgggtggcgc agcggcgacc tagggatcga tctggagggg
61 cttggggagc gtcagagac ctctagctcg agcgcgaggg acctccgcc
gggatgcctg
121 gggagcagat ggaccctact ggaagtcagt tggattcaga tttctctcag
caagatactc
181 cttgctgat aattgaagat tctcagcctg aaagccaggt tctagaggat
gattctggtt
241 ctcacttcag tatgctatct cgacaccttc ctaatctcca gacgcacaaa
```

The sequence is: Linear Circular

Enzymes to use: NEB enzymes
 All commercially available specificities
 All specificities
 All + defined oligonucleotide sequences
 Only defined oligonucleotide sequences
[\[define oligos\]](#)

Minimum ORF length to display: a.a.

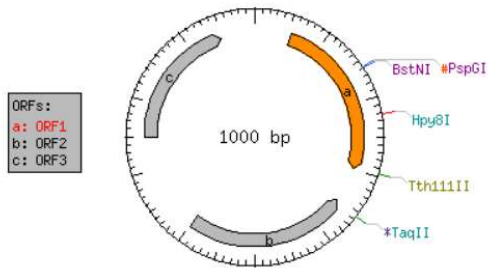
Standard sequences:
Plasmid vectors ▾
Viral + phage ▾

Name of sequence: (optional)

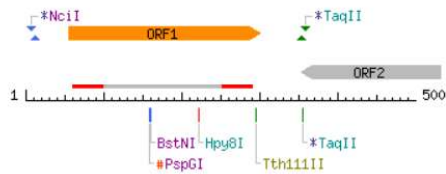
Earlier projects:

Note: Your earlier projects will be deleted 2 days after they were last accessed.

Cleavage code	Enzyme name code
⌞ blunt end cut	Available from NEB
⌞ 5' extension	Has other supplier
⌞ 3' extension	Not commercially available
⌞ cuts 1 strand	*: cleavage affected by CpG meth.
	#: cleavage affected by other meth.
	(enz.name): ambiguous site



ORFs:
a: ORF1
b: ORF2
c: ORF3



ORF1 N-term Met HpyCH4V *Hpy99I
Gln Lys Ser Ser Ser

Set Colors

[Help](#) [Comments](#)

Element	Color	Change
Scale	black	Change
Cut site - blunt	red	Change
Cut site - 5' ext.	royalblue	Change
Cut site - 3' ext.	green	Change
Cut site - one strand	black	Change
Cut site - unknown	black	Change
Cut site - multiple overlapping	darkmagenta	Change
Guide line	silver	Change
Supplier code - NEB	darkmagenta	Change
Supplier code - other commercial	darkcyan	Change
Supplier code - none	olive	Change
MS - CpG	darkslateblue	Change
MS - other	orangered	Change
Basepair hilite - unique	darkred	Change
Basepair hilite - degenerate	darkblue	Change
a.a. name	black	Change
a.a. background	yellow	Change
ORF body	mediumgray	Change
ORF outline (linear)	mediumgray	Change
ORF outline (circular)	black	Change
ORF name	black	Change
Highlighted ORF body	darkorange	Change
Highlighted ORF outline (linear)	darkorange	Change

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[What's new in V2.0](#) [Citing NEBcutter](#)

Local sequence file: 選擇檔案 未選擇任何檔案

GenBank number: [\[Browse GenBank\]](#)

or paste in your DNA sequence: (plain or FASTA format)

The sequence is: Linear Circular

Enzymes to use: NEB enzymes
 All commercially available specificities
 All specificities
 All + defined oligonucleotide sequences
 Only defined oligonucleotide sequences
[\[define oligos\]](#)

Minimum ORF length to display: a.a.

Name of sequence: (optional)

Earlier projects:

Standard sequences:

- # Plasmid vectors
- # Plasmid vectors
- PURExpr_DHFR_ctrl
- PURExpr_pmr
- LITMUS-U
- LITMUS28
- LITMUS28i
- LITMUS29
- LITMUS38
- LITMUS38i
- LITMUS39
- pACP-GPI
- pACPm
- pACYC177
- pACYC184
- pAd2-AvrII
- pAd2-BsaBI
- pASi3
- pBC4
- pBeloBAC11
- pBR322

NEBcutter V2.0

[Program Guide](#)

[Help](#)

[Comments](#)

This tool will take a DNA sequence and find the large, non-overlapping open reading frames using the E.coli genetic code and the sites for all Type II and commercially available Type III restriction enzymes that cut the sequence just once. By default, only enzymes available from NEB are used, but other sets may be chosen. Just enter your sequence and "submit". Further options will appear with the output. **The maximum size of the input file is 1 MByte, and the maximum sequence length is 300 KBases.**

[What's new in V2.0](#) [Citing NEBcutter](#)

Local sequence file: 未選擇任何檔案

GenBank number: [\[Browse GenBank\]](#)

or paste in your DNA sequence: (plain or FASTA format)

The sequence is: Linear Circular

Enzymes to use:

- NEB enzymes
- All commercially available specificities
- All specificities
- All + defined oligonucleotide sequences
- Only defined oligonucleotide sequences

[\[define oligos\]](#)

Minimum ORF length to display: a.a.

Name of sequence: (optional)

Earlier projects:

Standard sequences:

- # Plasmid vectors ▾
- # Viral + phage ▾
- # Viral + phage
- Ad2
- Lambda
- Lambda_gt11
- Lambda_NEB
- M13KE
- M13KO7
- M13mp18
- PhiX174
- PhiX174_NEB
- SV40
- T3
- T7