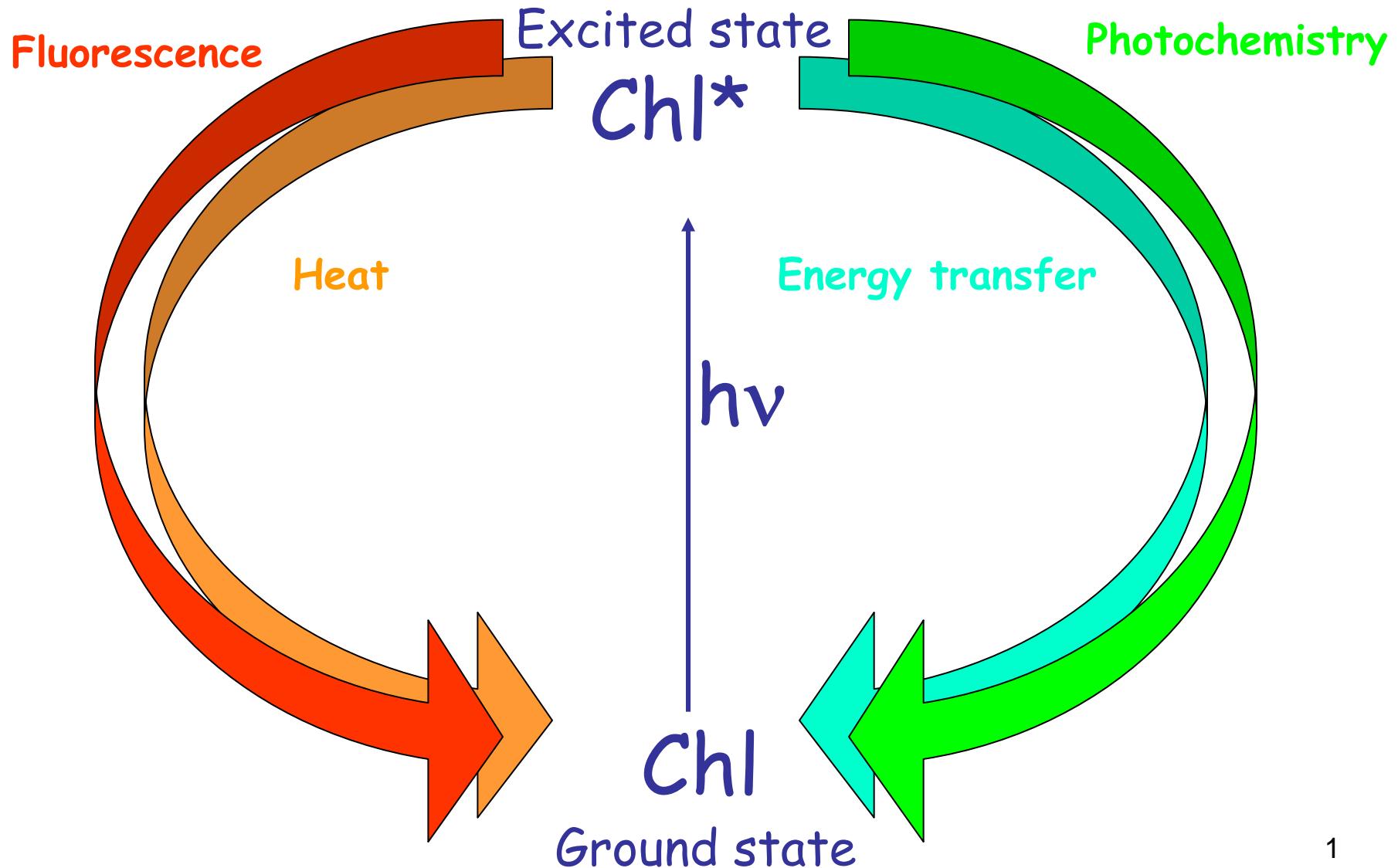
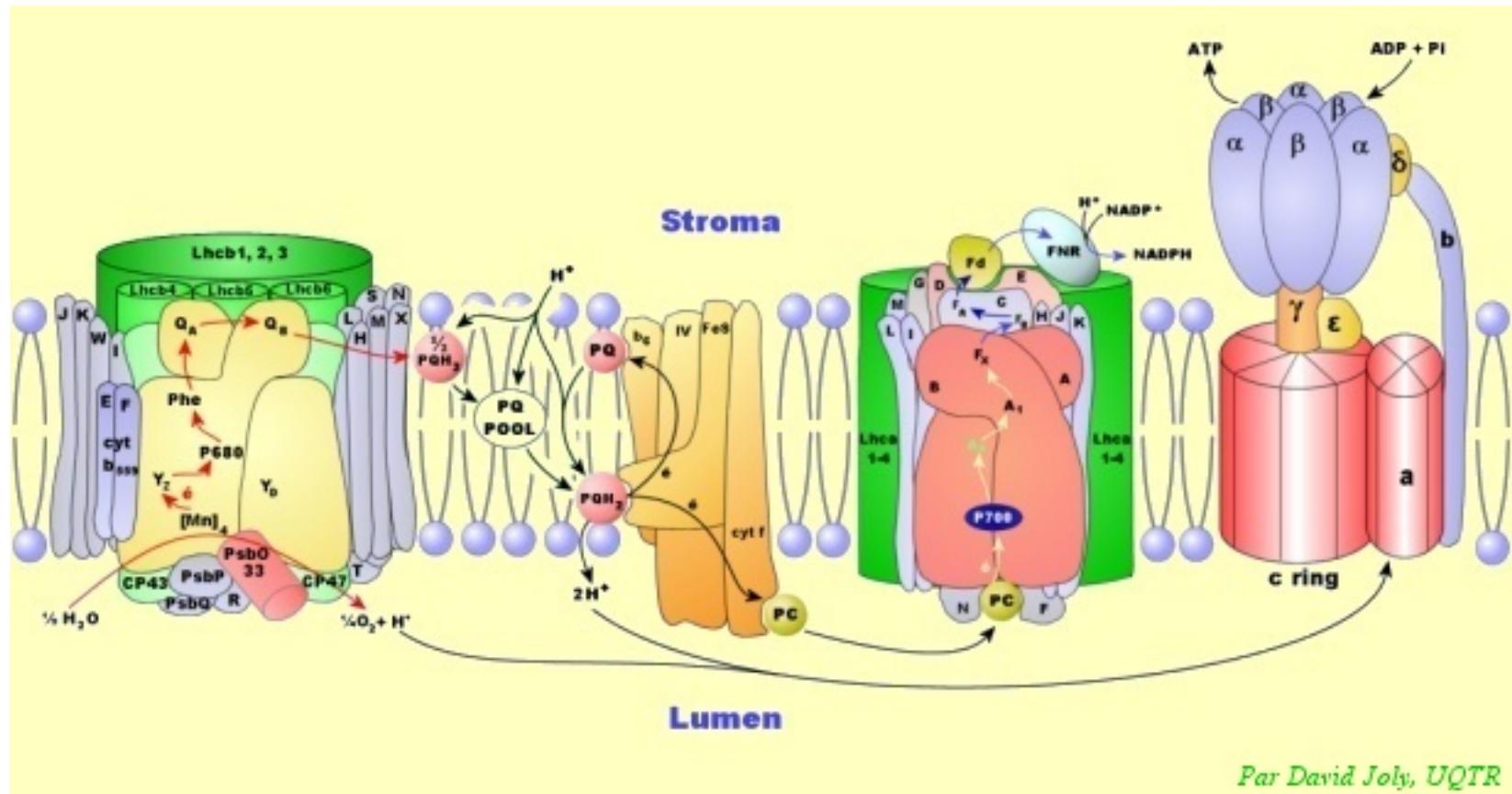


De-excitation of Chlorophyll



Quenching of Fluorescence

Photochemical Quenching



Par David Joly, UQTR

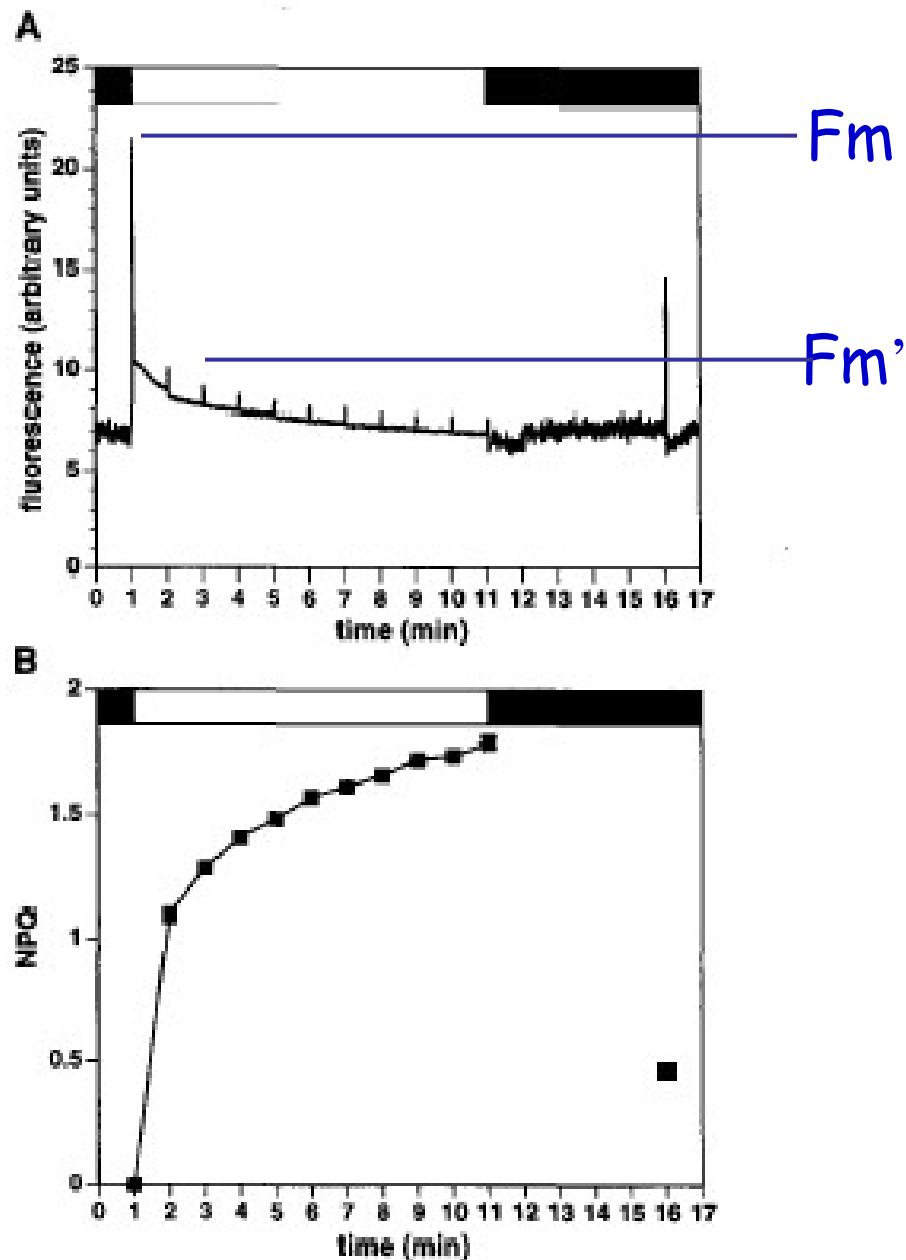
Photochemical Quenching via:
 Linear electron transfer to NADP+
 Mehler reaction (water-water cycle, pseudo-cyclic electron transfer)

Quenching of Fluorescence

Photochemical Quenching

Non-Photochemical Quenching: NPQ

Measurements of NPQ in Chlamydomonas



Decrease of Chl Fluorescence Results from:

State transition

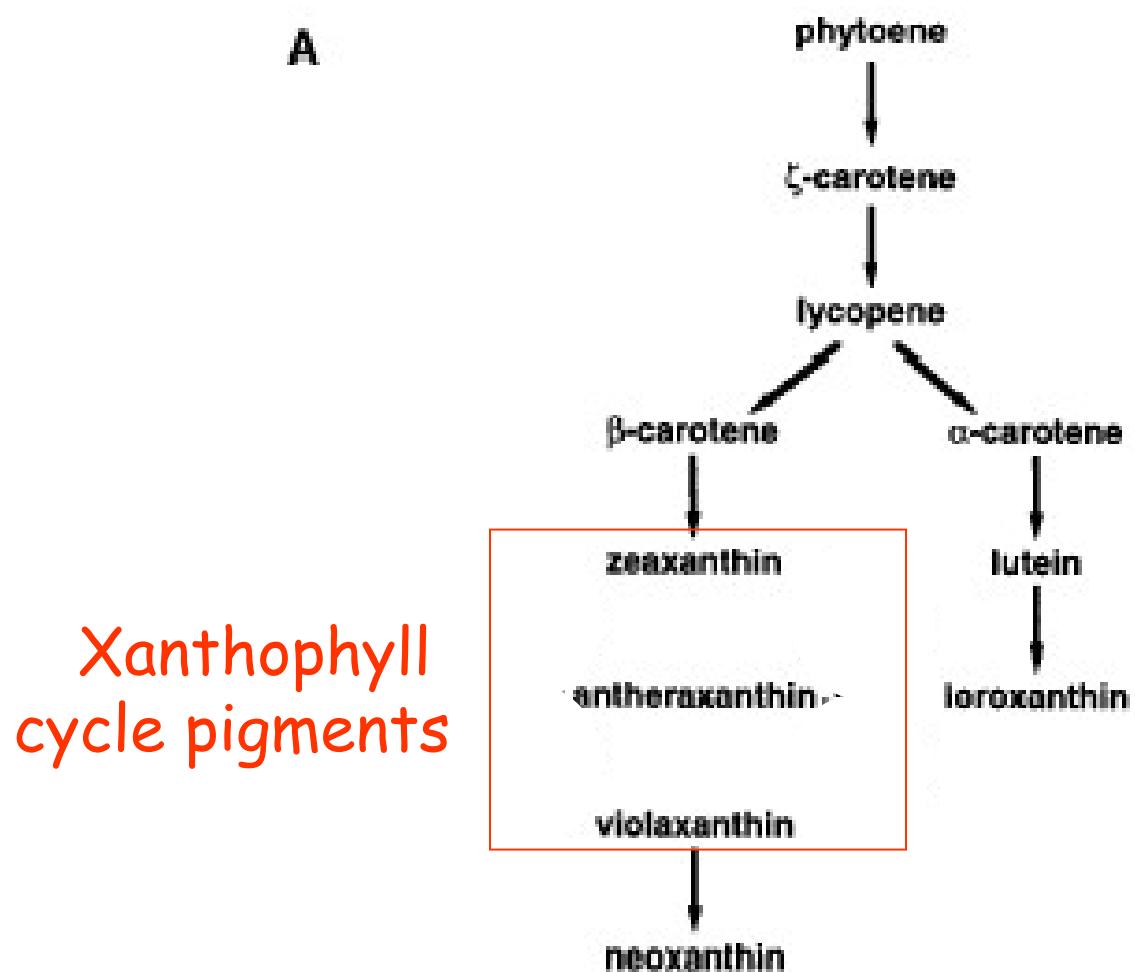
Photoinhibition (q_I)

NPQ (q_E)

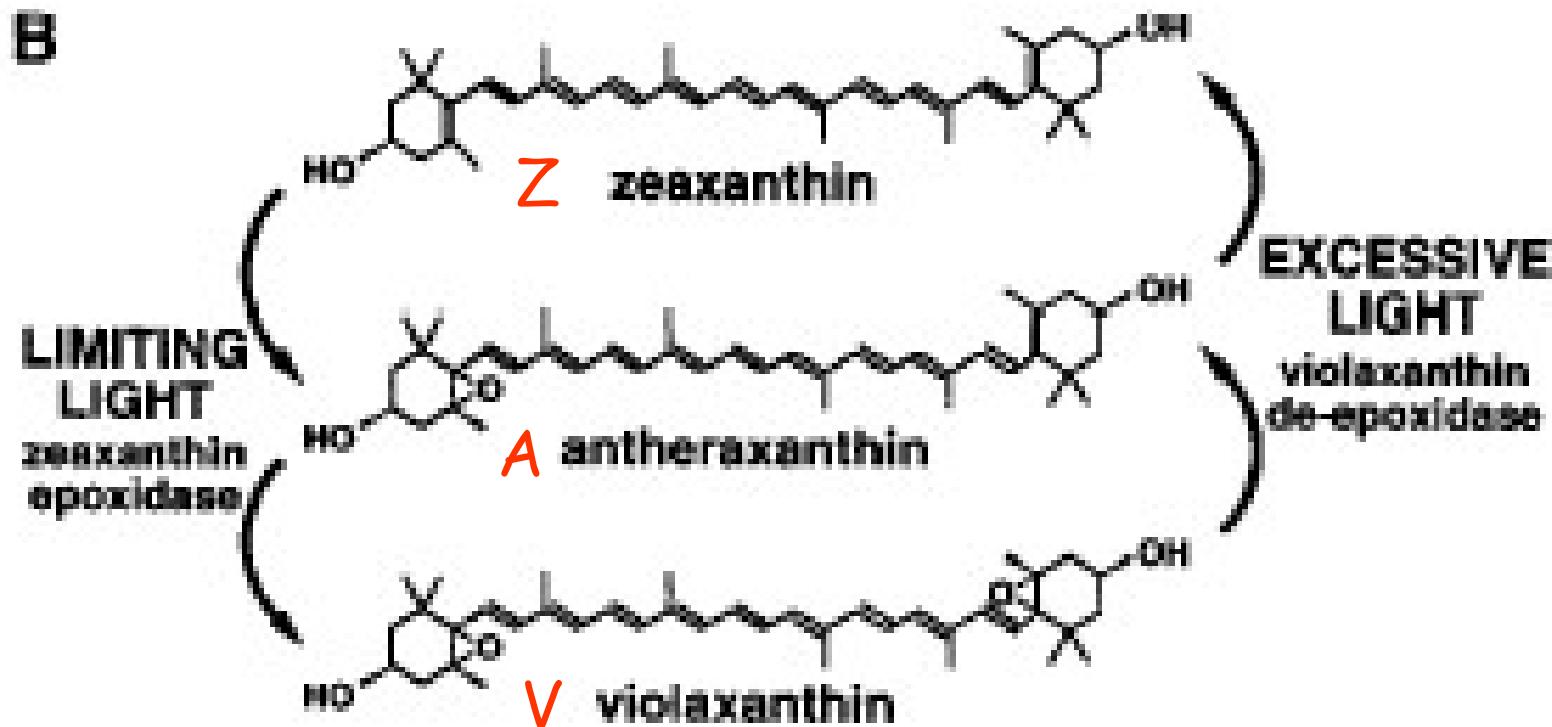
ΔpH^+ is required for NPQ

Specific Carotenoids are involved in NPQ

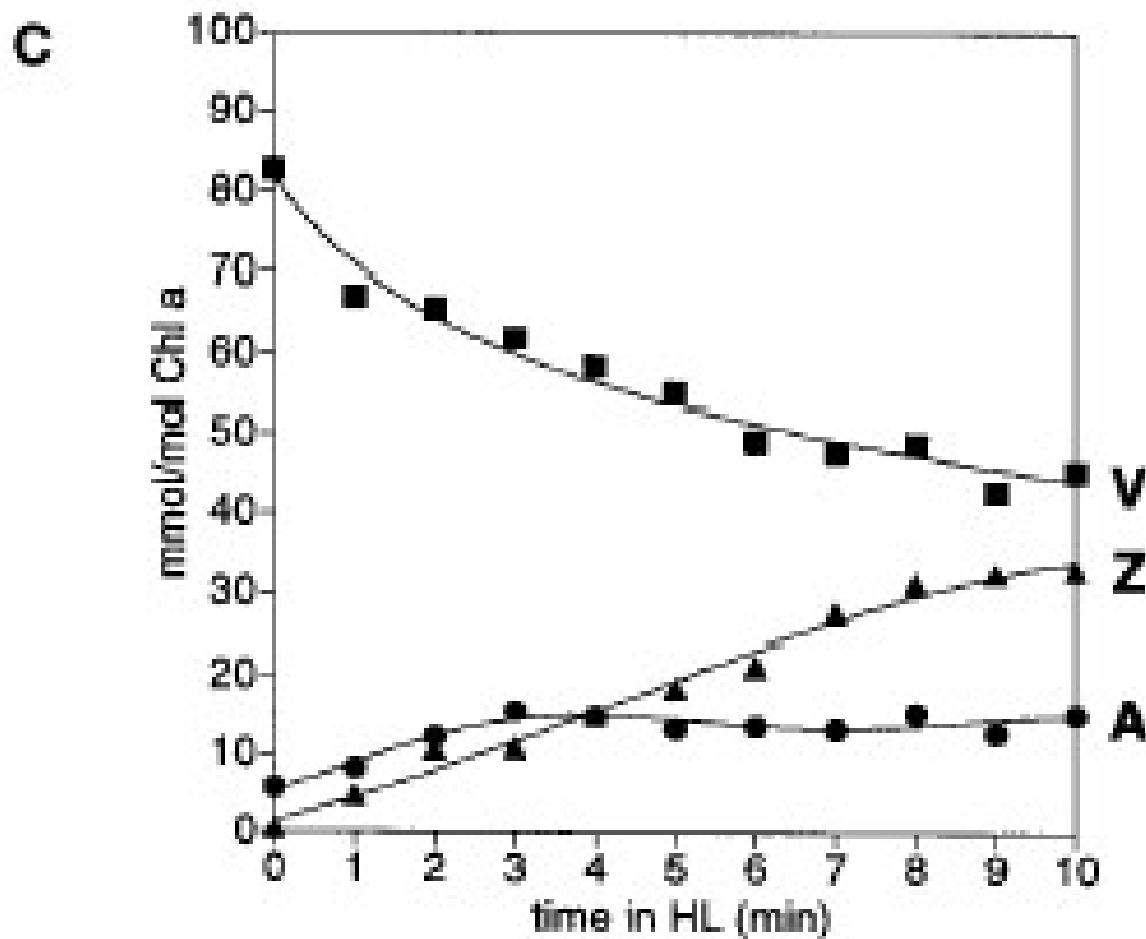
Biosynthesis of Carotenoids



Xanthophyll Cycle Pigments



Analysis of Xanthophyll Cycle Pigments in Chlamy following High Light Treatment



Analysis of Xanthophyll Cycle Pigments in Chlamy following High Light Treatment

Table 1. HPLC Analysis of the Xanthophyll Cycle Pigments in Wild-Type *Chlamydomonas* Cells under Different Growth Conditions*

Growth Medium	Incident PFD	mmol/mol Chlorophyll a			V + A + Z	(A + Z)/(V + A + Z)
		V	A	Z		
TAP	5	118	4	0	122	0.03
TAP	50	175	8	3	186	0.06
TAP	300	127	51	109	287	0.56
HS	50	123	3	0	126	0.02
HS	350	148	46	83	277	0.47

*Cells of an Arg⁺ control strain wt, derived from CC-425, were grown at the indicated PFD ($\mu\text{mol photons m}^{-2} \text{ sec}^{-1}$) for 9 days. Cells were grown photoheterotrophically on agar medium containing acetate as a carbon source (TAP) or photoautotrophically on minimal medium lacking acetate (HS). Values are the averages of duplicate determinations. V, violaxanthin; A, antheraxanthin; Z, zeaxanthin.

The Plant Cell, Vol. 9, 1369–1380, August 1997 © 1997 American Society of Plant Physiologists

Chlamydomonas Xanthophyll Cycle Mutants Identified by Video Imaging of Chlorophyll Fluorescence Quenching

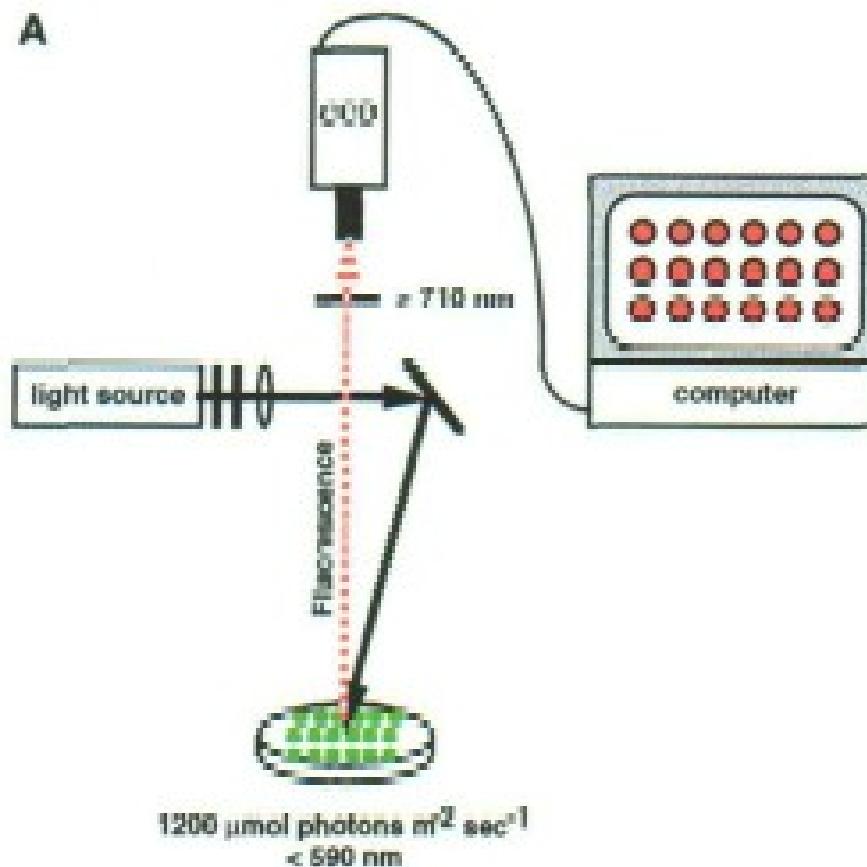
Krishna K. Niyogi,¹ Olle Björkman, and Arthur R. Grossman

The Plant Cell, Vol. 10, 1121–1134, July 1998, www.plantcell.org © 1998 American Society of Plant Physiologists

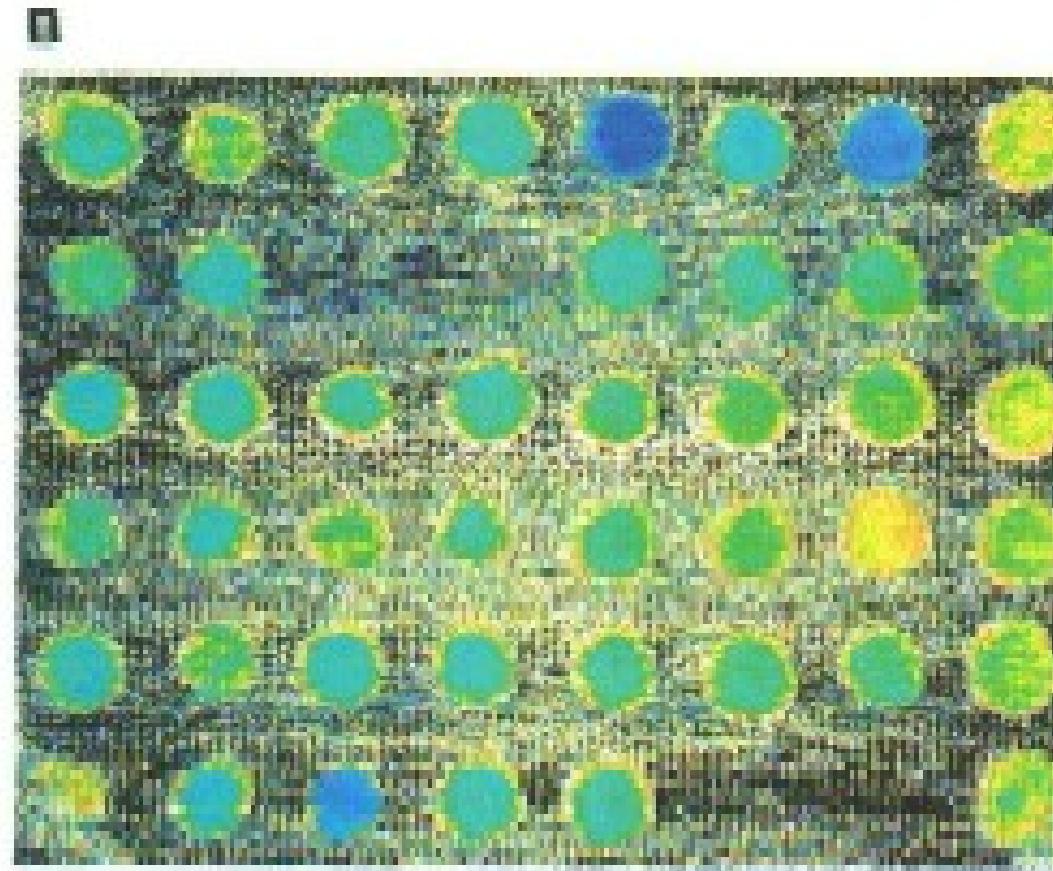
Arabidopsis Mutants Define a Central Role for the Xanthophyll Cycle in the Regulation of Photosynthetic Energy Conversion

Krishna K. Niyogi,¹ Arthur R. Grossman, and Olle Björkman

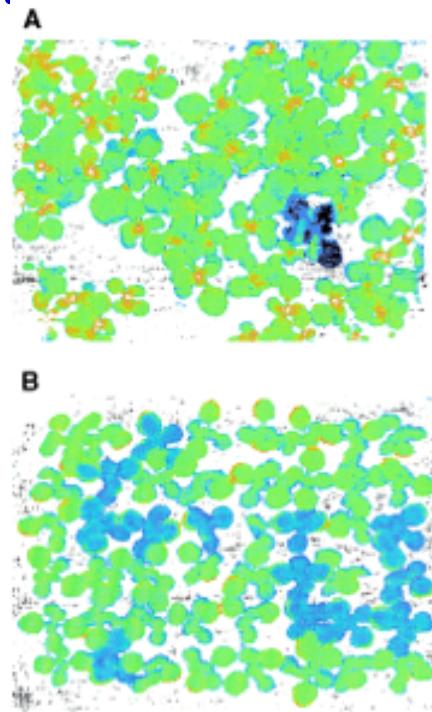
Screening for NPQ Mutants



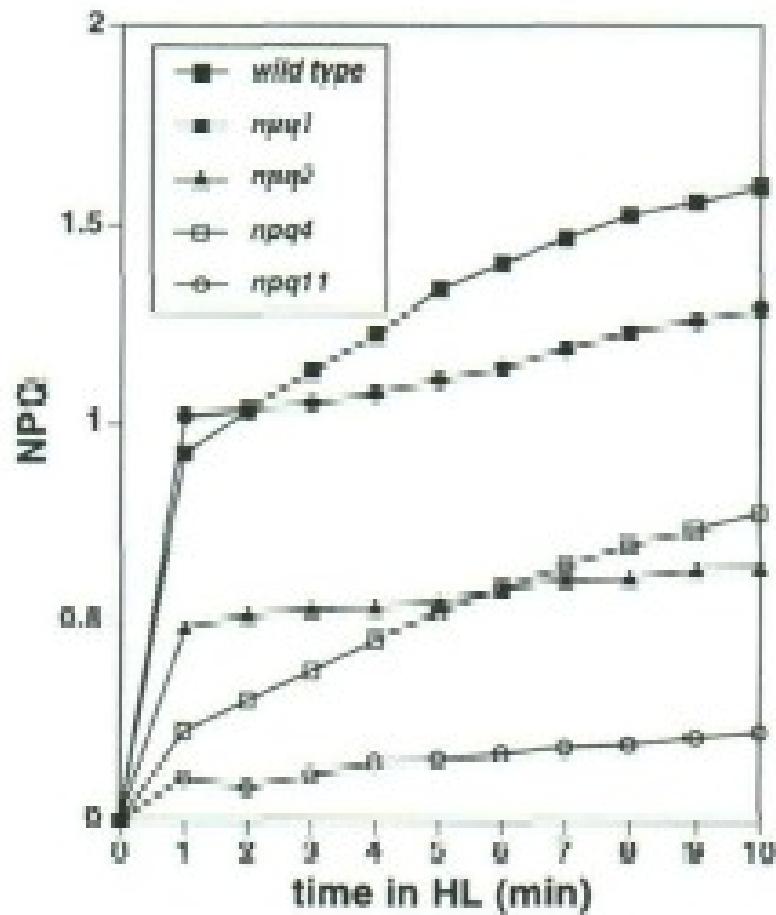
Chlamydomonas NPQ mutants



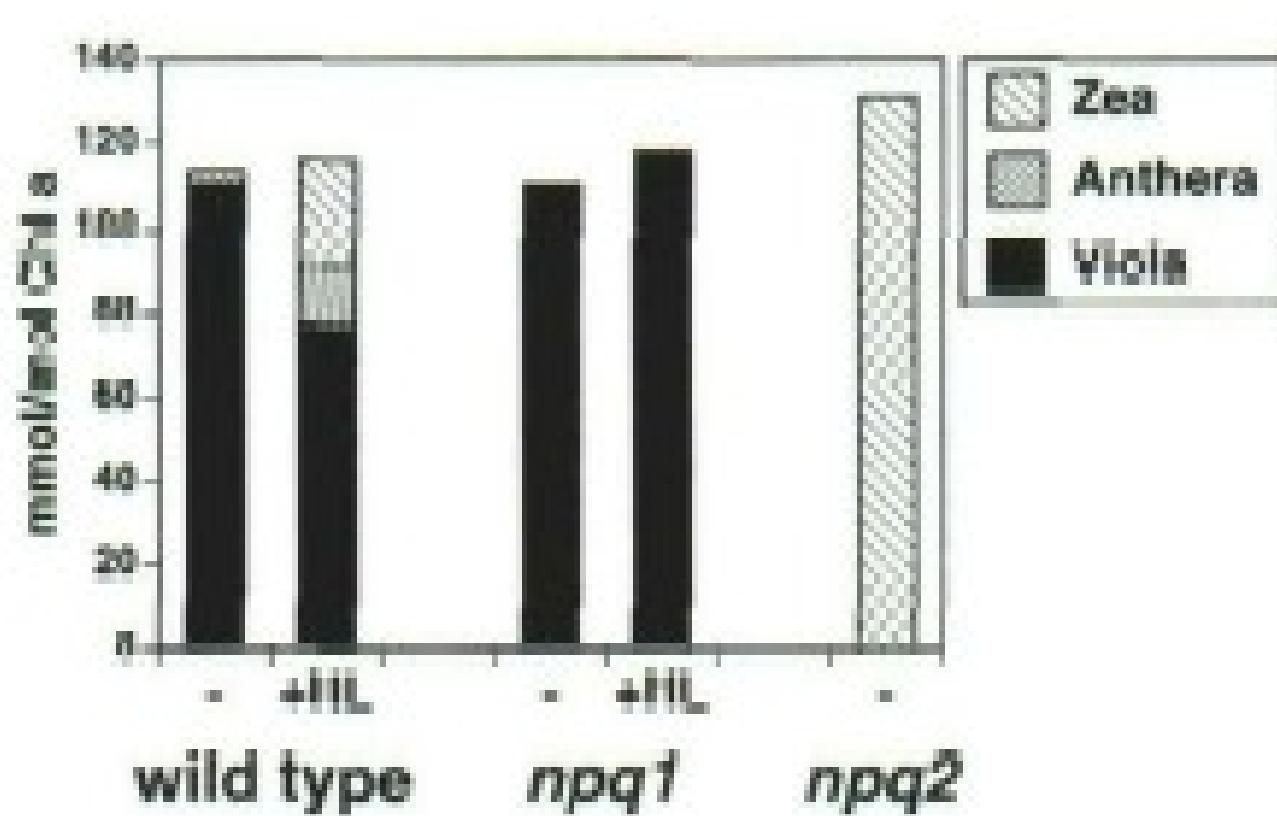
Arabidopsis NPQ mutants



NPQ in Wild Type and Mutants

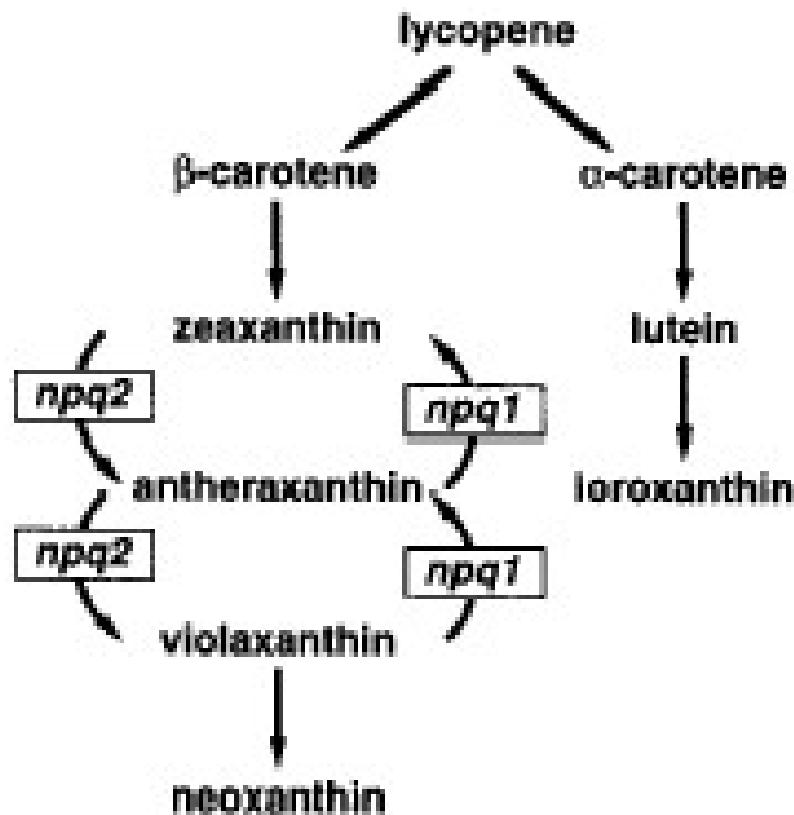


Composition of Xanthophyll Cycle Pigments in WT and Mutants following High Light treatment

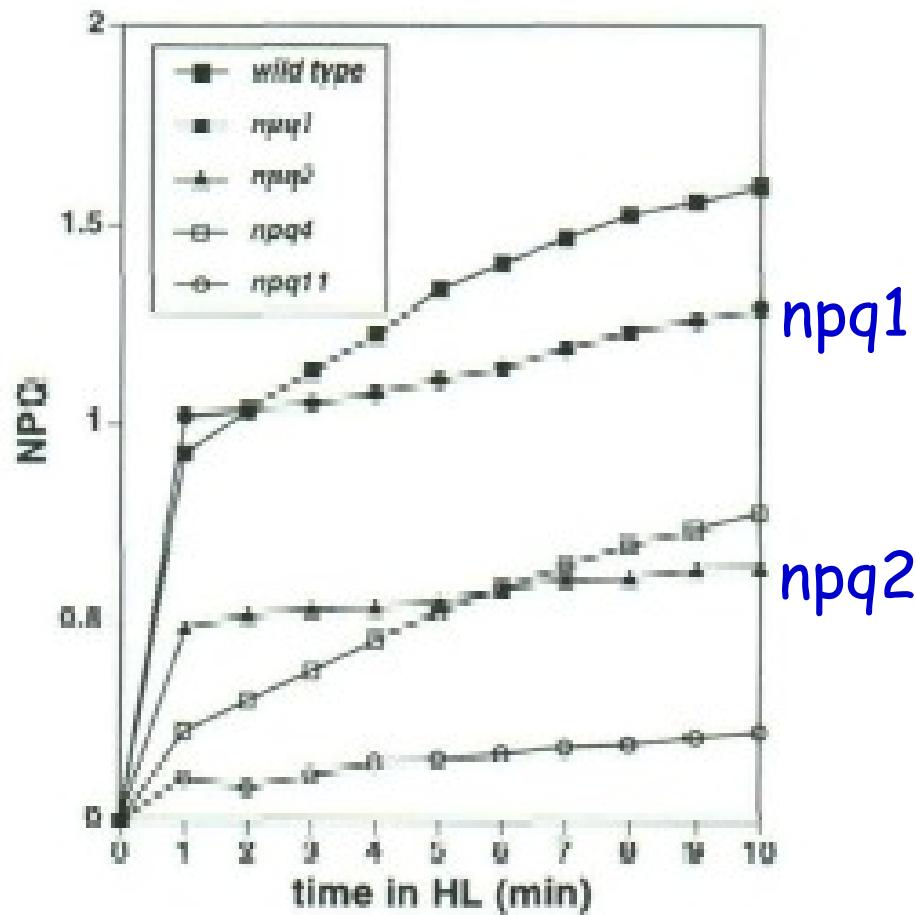


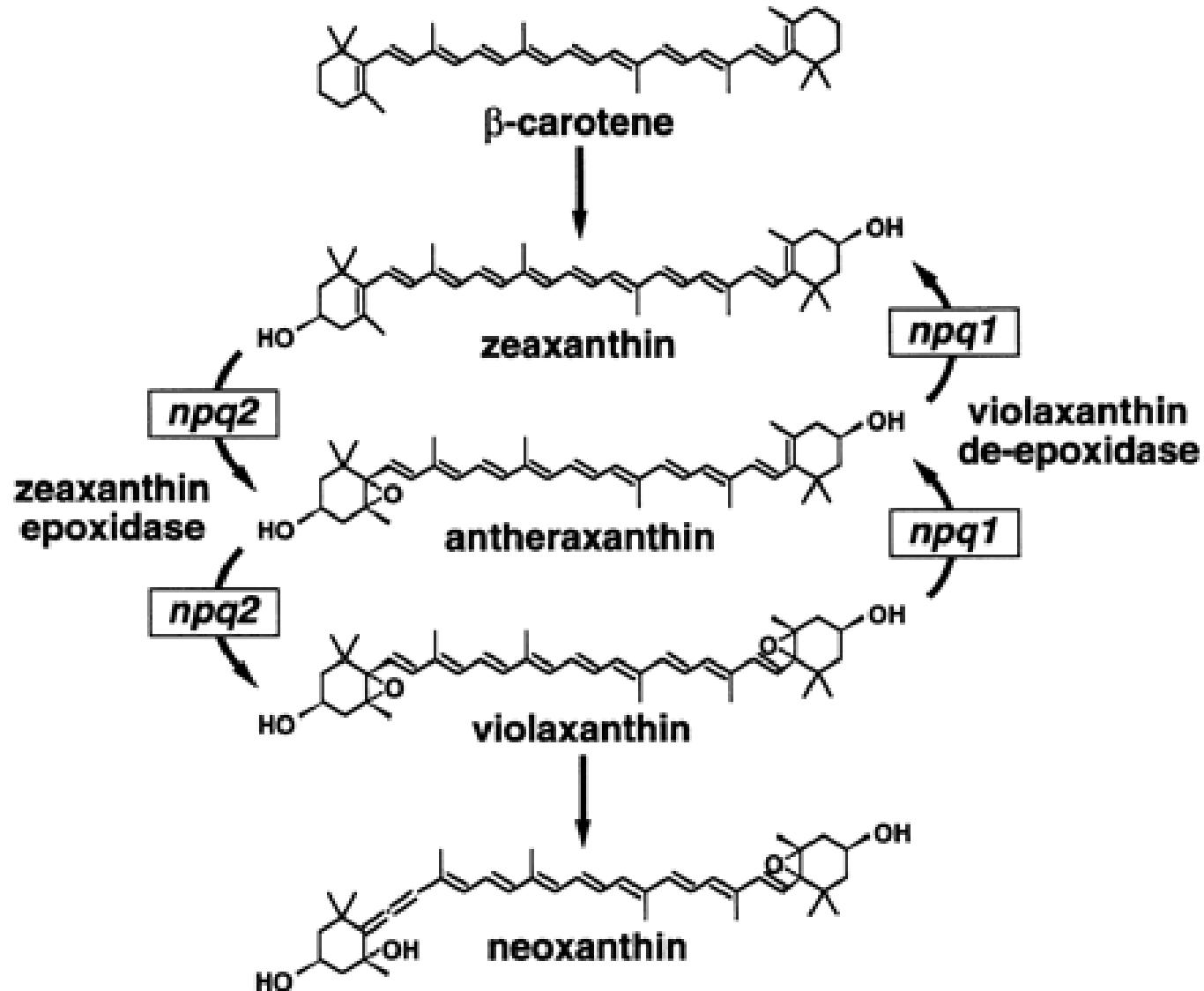
npq1 is impaired in violaxanthin de-epoxidase (VDE)

npq2 is impaired in zeaxanthin epoxidase

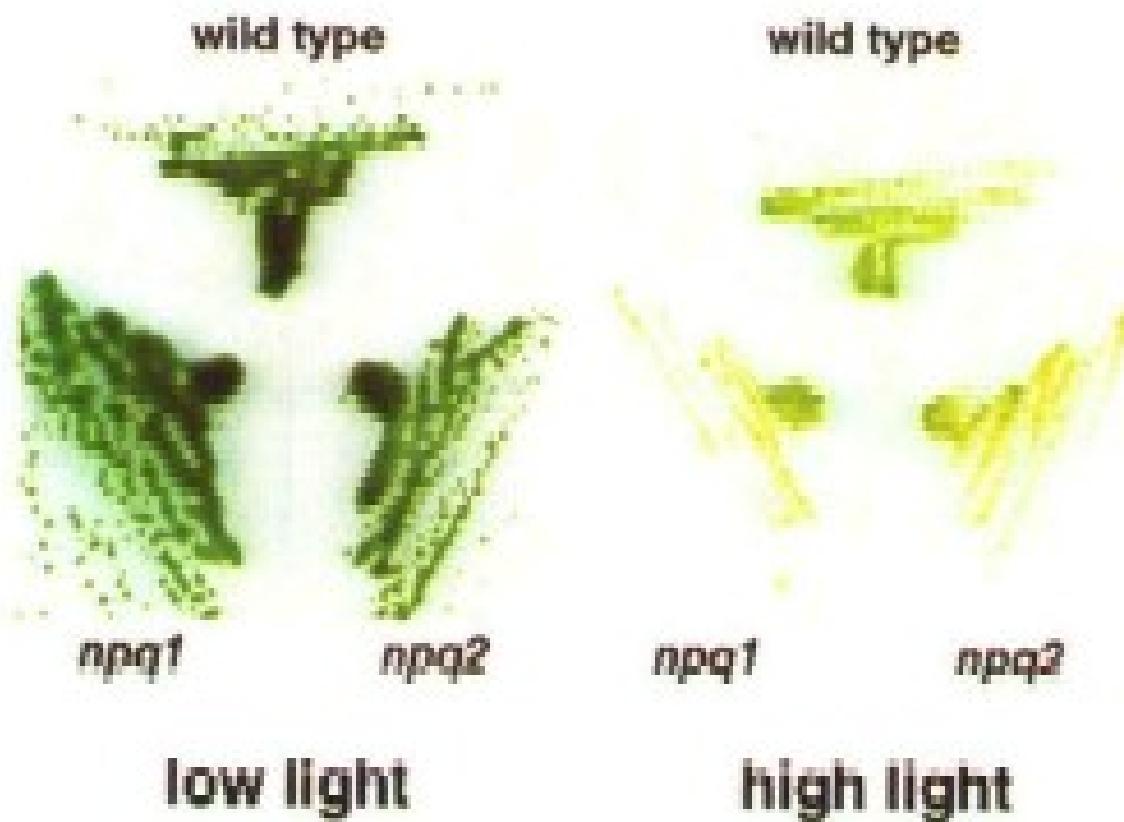


NPQ in Wild Type and Mutants

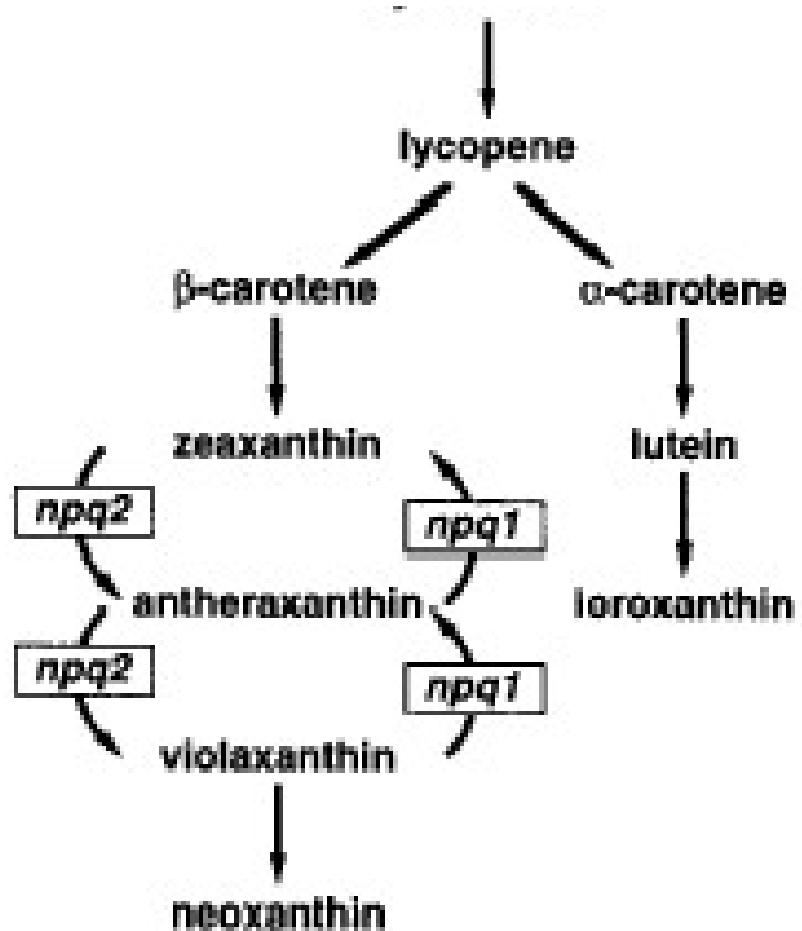




Growth during High Light Illumination of WT and NPQ Mutants



Double mutants impaired in npq1 and biosynthesis of lutein/loroxanthin are High Light Sensitive



Xanthophyll cycle pigments in wild-type and *npq4* leaves

mmol per mol chlorophyll a

		V	A	Z	V + A + Z	(A + Z)/(V + A + Z)
Wild type	before HL	53 ± 8	16 ± 2	13 ± 1	82 ± 9	0.36 ± 0.03
	after HL	18 ± 1	8 ± 1	43 ± 9	70 ± 10	0.73 ± 0.03
<i>npq4-1</i>	before HL	54 ± 2	12 ± 1	6 ± 1	73 ± 2	0.26 ± 0.02
	after HL	17 ± 1	8 ± 1	43 ± 5	68 ± 6	0.75 ± 0.02

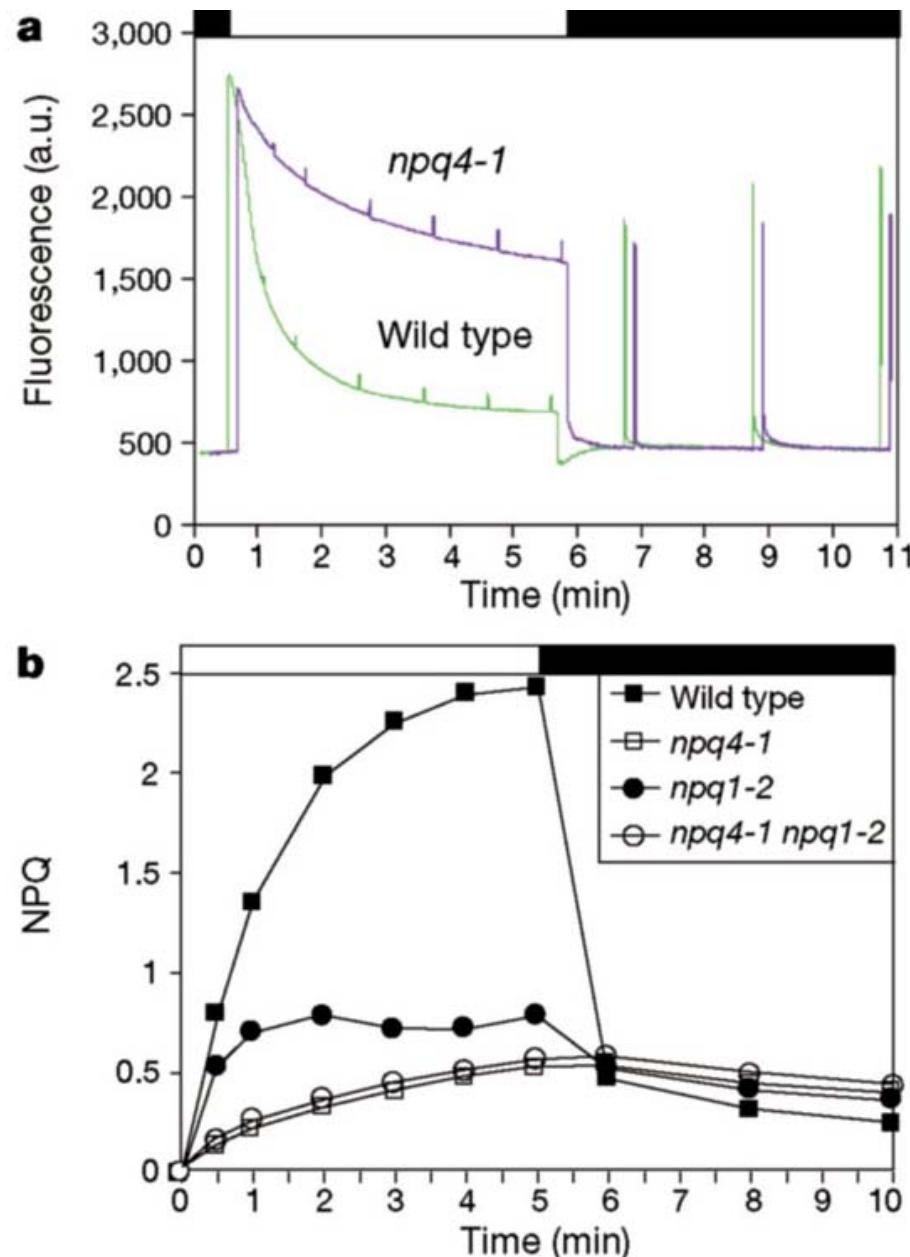
Pigment composition of leaves was measured by high-performance liquid chromatography either before or after exposure to high light (HL, $640 \mu\text{mol photons m}^{-2} \text{ sec}^{-1}$) for 30 min. Values are means ± s.e. ($n = 4$).

A pigment-binding protein essential for regulation of photosynthetic light harvesting

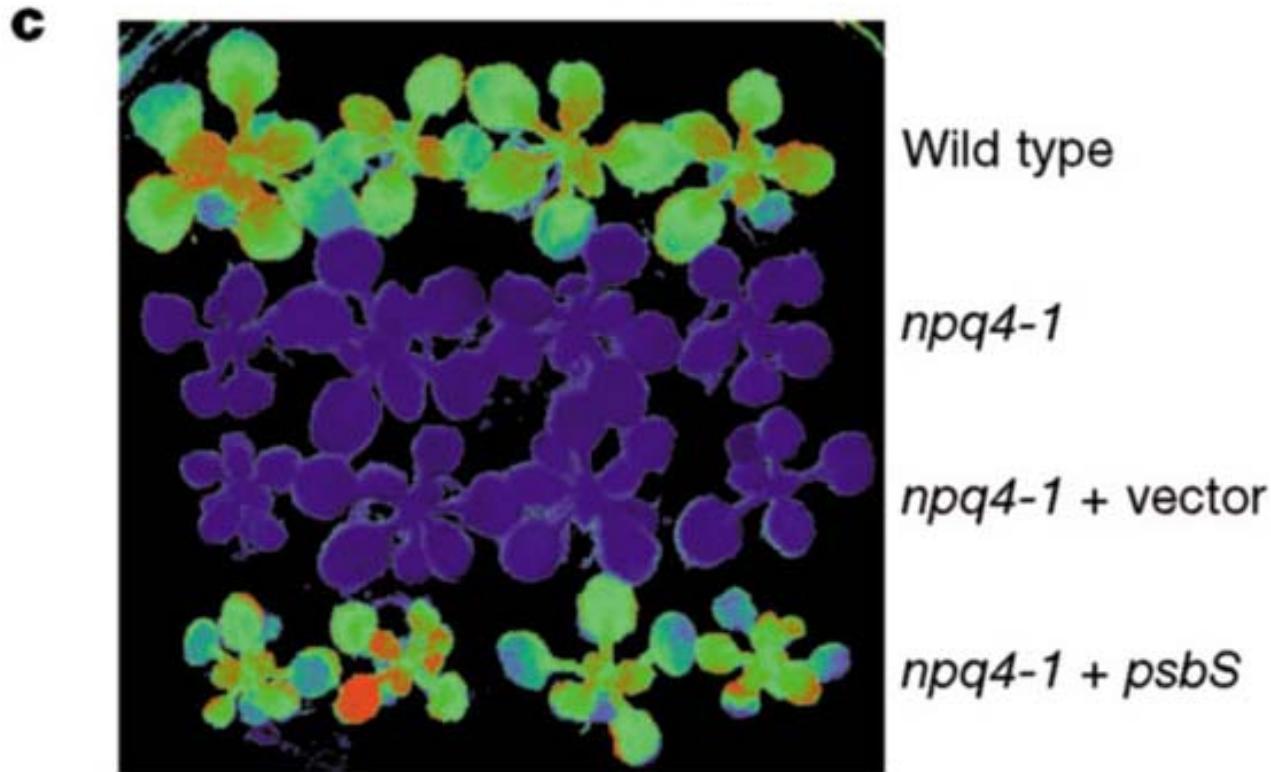
Xiao-Ping Li⁺, Olle Björkman[†], Connie Shih[†], Arthur R. Grossman[†], Magnus Rosenquist^{‡§}, Stefan Jansson[‡] & Krishna K. Niyogi⁺

NATURE | VOL 403 | 27 JANUARY 2000 | www.nature.com

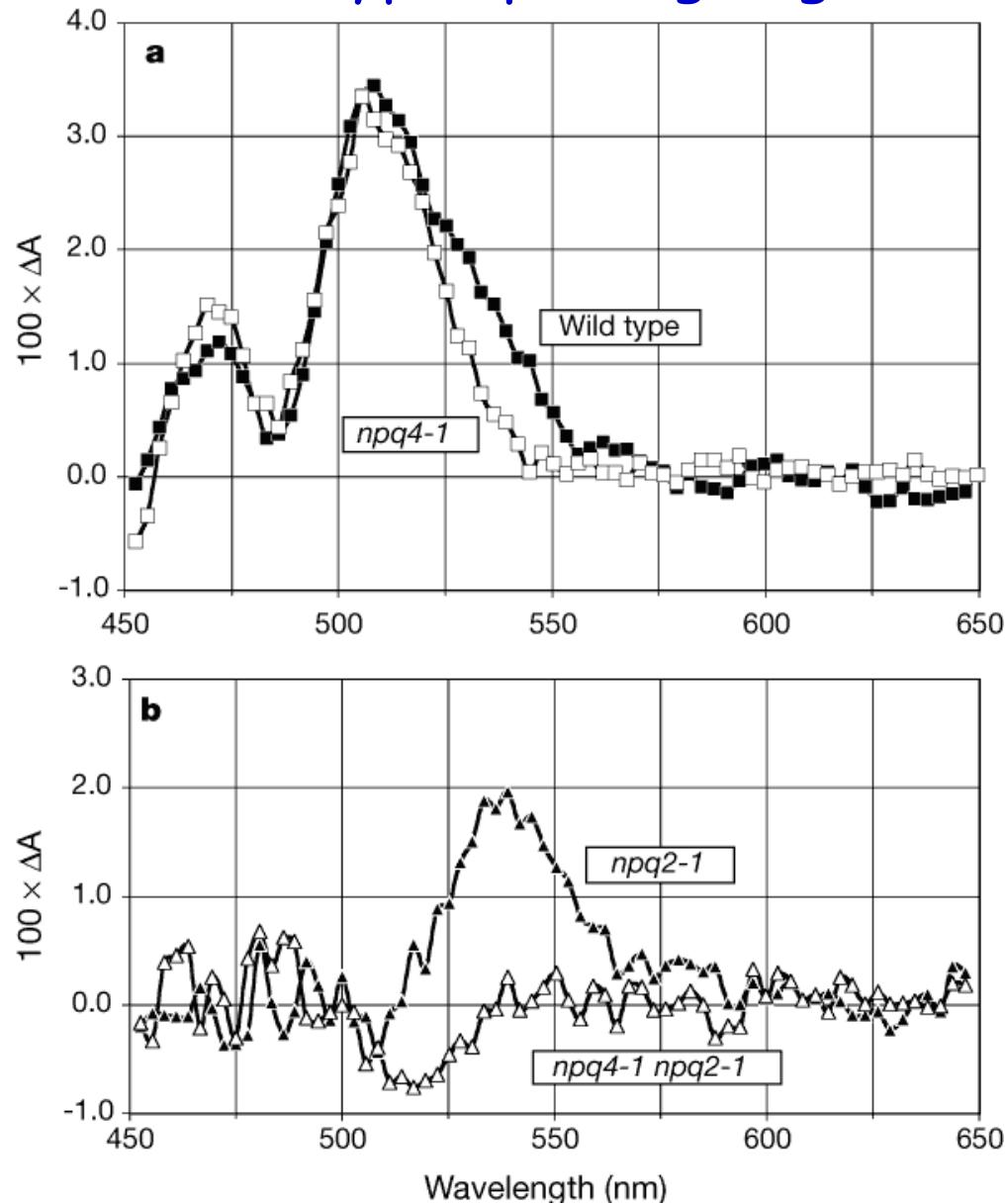
Mutant *npq4* exhibits abnormal NPQ



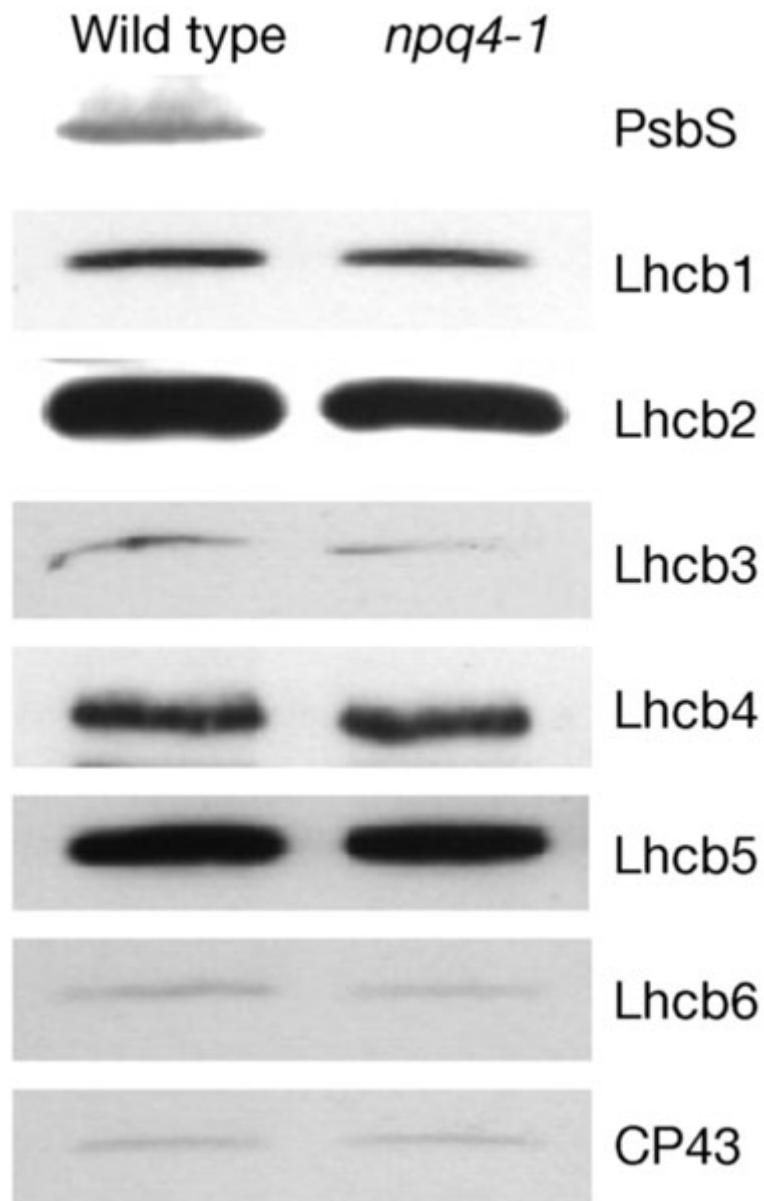
Mutant *npq4* is complemented by *PsbS*



Mutant npq4 does not exhibit the typical spectral changes observed in the wild type upon high light illumination



Content of LHC protein in wild type and mutant *npg4*



Photosynthetic parameters of wild type and *npq4* leaves

Parameter	Wild type	<i>npq4-1</i>
$\Phi(O_2)$	$0.0818 \pm 0.0024 (n = 5)$	$0.0812 \pm 0.0033 (n = 5)$
P_{\max}	$18.15 \pm 1.06 (n = 5)$	$18.87 \pm 0.92 (n = 5)$
F_v/F_m	$0.837 \pm 0.003 (n = 8)$	$0.836 \pm 0.004 (n = 8)$

$\Phi(O_2)$, apparent quantum yield of O_2 evolution (O_2 evolved per incident photon); P_{\max} , maximum rate of O_2 evolution ($\mu\text{mol } O_2 \text{ m}^{-2} \text{ sec}^{-1}$); F_v/F_m , maximum quantum yield of photosystem II electron transport. Values are means \pm s.e.

PsbS in specifically involved in NPQ and not in light harvesting

PsbS is required for the conformational change essential for NPQ